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(54) Title: CORYNEBACTERIUM GLUTAMICUM GENES ENCODING PROTEINS INVOLVED IN HOMEOSTASIS AND ADAPTATION

(57) Abstract: Isolated nucleic acid molecules, designated HA nucleic acid molecules, which encode novel HA proteins from *Corynebacterium glutamicum* are described. The invention also provides antisense nucleic acid molecules, recombinant expression vectors containing HA nucleic acid molecules, and host cells into which the expression vectors have been introduced. The invention still further provides isolated HA proteins, mutated HA proteins, fusion proteins, antigenic peptides and methods for the improvement of production of a desired compound from *C. glutamicum* based on genetic engineering of HA genes in this organism.

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***CORYNEBACTERIUM GLUTAMICUM* GENES ENCODING PROTEINS
INVOLVED IN HOMEOSTASIS AND ADAPTATION**

Related Applications

- 5 This application claims priority to prior filed U.S. Provisional Patent Application
Serial No. 60/141031, filed June 25, 1999. This application also claims priority to prior
filed German Patent Application No. 19931636.8, filed July 8, 1999, German Patent
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Application No. 19941391.6, filed August 31, 1999, and German Patent Application No.
25 19942088.2, filed September 3, 1999. The entire contents of all of the aforementioned
applications are hereby expressly incorporated herein by this reference.

Background of the Invention

- 30 Certain products and by-products of naturally-occurring metabolic processes in
cells have utility in a wide array of industries, including the food, feed, cosmetics, and
pharmaceutical industries. These molecules, collectively termed 'fine chemicals',
include organic acids, both proteinogenic and non-proteinogenic amino acids,

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nucleotides and nucleosides, lipids and fatty acids, diols, carbohydrates, aromatic compounds, vitamins and cofactors, and enzymes. Their production is most conveniently performed through the large-scale culture of bacteria developed to produce and secrete large quantities of one or more desired molecules. One particularly useful

5 organism for this purpose is *Corynebacterium glutamicum*, a gram positive, nonpathogenic bacterium. Through strain selection, a number of mutant strains have been developed which produce an array of desirable compounds. However, selection of strains improved for the production of a particular molecule is a time-consuming and difficult process.

10

Summary of the Invention

The invention provides novel bacterial nucleic acid molecules which have a variety of uses. These uses include the identification of microorganisms which can be used to produce fine chemicals, the modulation of fine chemical production in *C.*

15 *glutamicum* or related bacteria, the typing or identification of *C. glutamicum* or related bacteria, as reference points for mapping the *C. glutamicum* genome, and as markers for transformation. These novel nucleic acid molecules encode proteins, referred to herein as homeostasis and adaptation (HA) proteins.

C. glutamicum is a gram positive, aerobic bacterium which is commonly used in

20 industry for the large-scale production of a variety of fine chemicals, and also for the degradation of hydrocarbons (such as in petroleum spills) and for the oxidation of terpenoids. The HA nucleic acid molecules of the invention, therefore, can be used to identify microorganisms which can be used to produce fine chemicals, *e.g.*, by fermentation processes. Modulation of the expression of the HA nucleic acids of the

25 invention, or modification of the sequence of the HA nucleic acid molecules of the invention, can be used to modulate the production of one or more fine chemicals from a microorganism (*e.g.*, to improve the yield or production of one or more fine chemicals from a *Corynebacterium* or *Brevibacterium* species).

The HA nucleic acids of the invention may also be used to identify an organism

30 as being *Corynebacterium glutamicum* or a close relative thereof, or to identify the presence of *C. glutamicum* or a relative thereof in a mixed population of microorganisms. The invention provides the nucleic acid sequences of a number of *C.*

glutamicum genes; by probing the extracted genomic DNA of a culture of a unique or mixed population of microorganisms under stringent conditions with a probe spanning a region of a *C. glutamicum* gene which is unique to this organism, one can ascertain whether this organism is present. Although *Corynebacterium glutamicum* itself is nonpathogenic, it is related to species pathogenic in humans, such as *Corynebacterium diphtheriae* (the causative agent of diphtheria); the detection of such organisms is of significant clinical relevance.

The HA nucleic acid molecules of the invention may also serve as reference points for mapping of the *C. glutamicum* genome, or of genomes of related organisms. Similarly, these molecules, or variants or portions thereof, may serve as markers for genetically engineered *Corynebacterium* or *Brevibacterium* species.

The HA proteins encoded by the novel nucleic acid molecules of the invention are capable of, for example, performing a function involved in the maintenance of homeostasis in *C. glutamicum*, or in the ability of this microorganism to adapt to different environmental conditions. Given the availability of cloning vectors for use in *Corynebacterium glutamicum*, such as those disclosed in Sinskey *et al.*, U.S. Patent No. 4,649,119, and techniques for genetic manipulation of *C. glutamicum* and the related *Brevibacterium* species (*e.g.*, *lactofermentum*) (Yoshihama *et al.*, *J. Bacteriol.* 162: 591-597 (1985); Katsumata *et al.*, *J. Bacteriol.* 159: 306-311 (1984); and Santamaria *et al.*, *J. Gen. Microbiol.* 130: 2237-2246 (1984)), the nucleic acid molecules of the invention may be utilized in the genetic engineering of this organism to make it a better or more efficient producer of one or more fine chemicals. This improved production or efficiency of production of a fine chemical may be due to a direct effect of manipulation of a gene of the invention, or it may be due to an indirect effect of such manipulation.

There are a number of mechanisms by which the alteration of an HA protein of the invention may directly affect the yield, production, and/or efficiency of production of a fine chemical from a *C. glutamicum* strain incorporating such an altered protein. For example, by engineering enzymes which modify or degrade aromatic or aliphatic compounds such that these enzymes are increased or decreased in activity or number, it may be possible to modulate the production of one or more fine chemicals which are the modification or degradation products of these compounds. Similarly, enzymes involved in the metabolism of inorganic compounds provide key molecules (*e.g.* phosphorous,

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sulfur, and nitrogen molecules) for the biosynthesis of such fine chemicals as amino acids, vitamins, and nucleic acids. By altering the activity or number of these enzymes in *C. glutamicum*, it may be possible to increase the conversion of these inorganic compounds (or to use alternate inorganic compounds) to thus permit improved rates of

5 incorporation of inorganic atoms into these fine chemicals. Genetic engineering of *C. glutamicum* enzymes involved in general cellular processes may also directly improve fine chemical production, since many of these enzymes directly modify fine chemicals (e.g., amino acids) or the enzymes which are involved in fine chemical synthesis or secretion. Modulation of the activity or number of cellular proteases may also have a

10 direct effect on fine chemical production, since many proteases may degrade fine chemicals or enzymes involved in fine chemical production or breakdown.

Further, the aforementioned enzymes which participate in aromatic/aliphatic compound modification or degradation, general biocatalysis, inorganic compound metabolism or proteolysis are each themselves fine chemicals, desirable for their activity

15 in various *in vitro* industrial applications. By altering the number of copies of the gene for one or more of these enzymes in *C. glutamicum* it may be possible to increase the number of these proteins produced by the cell, thereby increasing the potential yield or efficiency of production of these proteins from large-scale *C. glutamicum* or related bacterial cultures.

20 The alteration of an HA protein of the invention may also indirectly affect the yield, production, and/or efficiency of production of a fine chemical from a *C. glutamicum* strain incorporating such an altered protein. For example, by modulating the activity and/or number of those proteins involved in the construction or rearrangement of the cell wall, it may be possible to modify the structure of the cell wall

25 itself such that the cell is able to better withstand the mechanical and other stresses present during large-scale fermentative culture. Also, large-scale growth of *C. glutamicum* requires significant cell wall production. Modulation of the activity or number of cell wall biosynthetic or degradative enzymes may allow more rapid rates of cell wall biosynthesis, which in turn may permit increased growth rates of this

30 microorganism in culture and thereby increase the number of cells producing the desired fine chemical.

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By modifying the HA enzymes of the invention, one may also indirectly impact the yield, production, or efficiency of production of one or more fine chemicals from *C. glutamicum*. For example, many of the general enzymes in *C. glutamicum* may have a significant impact on global cellular processes (e.g., regulatory processes) which in turn have a significant effect on fine chemical metabolism. Similarly, proteases, enzymes which modify or degrade possibly toxic aromatic or aliphatic compounds, and enzymes which promote the metabolism of inorganic compounds all serve to increase the viability of *C. glutamicum*. The proteases aid in the selective removal of misfolded or misregulated proteins, such as those that might occur under the relatively stressful environmental conditions encountered during large-scale fermentor culture. By altering these proteins, it may be possible to further enhance this activity and to improve the viability of *C. glutamicum* in culture. The aromatic/aliphatic modification or degradation proteins not only serve to detoxify these waste compounds (which may be encountered as impurities in culture medium or as waste products from cells themselves), but also to permit the cells to utilize alternate carbon sources if the optimal carbon source is limiting in the culture. By increasing their number and/or activity, the survival of *C. glutamicum* cells in culture may be enhanced. The inorganic metabolism proteins of the invention supply the cell with inorganic molecules required for all protein and nucleotide (among others) synthesis, and thus are critical for the overall viability of the cell. An increase in the number of viable cells producing one or more desired fine chemicals in large-scale culture should result in a concomitant increase in the yield, production, and/or efficiency of production of the fine chemical in the culture.

The invention provides novel nucleic acid molecules which encode proteins, referred to herein as HA proteins, which are capable of, for example, performing a function involved in the maintenance of homeostasis in *C. glutamicum*, or of participating in the ability of this microorganism to adapt to different environmental conditions. Nucleic acid molecules encoding an HA protein are referred to herein as HA nucleic acid molecules. In a preferred embodiment, an HA protein participates in *C. glutamicum* cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or possesses a *C. glutamicum* enzymatic or proteolytic activity. Examples of such proteins include those encoded by the genes set forth in Table 1.

- Accordingly, one aspect of the invention pertains to isolated nucleic acid molecules (e.g., cDNAs, DNAs, or RNAs) comprising a nucleotide sequence encoding an HA protein or biologically active portions thereof, as well as nucleic acid fragments suitable as primers or hybridization probes for the detection or amplification of HA-
- 5 encoding nucleic acids (e.g., DNA or mRNA). In particularly preferred embodiments, the isolated nucleic acid molecule comprises one of the nucleotide sequences set forth as the odd-numbered SEQ ID NOs in the Sequence Listing (e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7....), or the coding region or a complement thereof of one of these nucleotide sequences. In other particularly preferred embodiments, the
- 10 isolated nucleic acid molecule of the invention comprises a nucleotide sequence which hybridizes to or is at least about 50%, preferably at least about 60%, more preferably at least about 70%, 80% or 90%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more homologous to a nucleotide sequence set forth as an odd-numbered SEQ ID NO in the Sequence Listing (e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5,
- 15 SEQ ID NO:7....), or a portion thereof. In other preferred embodiments, the isolated nucleic acid molecule encodes one of the amino acid sequences set forth in as an even-numbered SEQ ID NO in the Sequence Listing (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8....). The preferred HA proteins of the present invention also preferably possess at least one of the HA activities described herein.
- 20 In another embodiment, the isolated nucleic acid molecule encodes a protein or portion thereof wherein the protein or portion thereof includes an amino acid sequence which is sufficiently homologous to an amino acid sequence of the invention (e.g., a sequence having an even-numbered SEQ ID NO: in the Sequence Listing), e.g., sufficiently homologous to an amino acid sequence of the invention such that the protein
- 25 or portion thereof maintains an HA activity. Preferably, the protein or portion thereof encoded by the nucleic acid molecule maintains the ability to participate in the maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions. In one embodiment, the protein encoded by the nucleic acid molecule is at least about 50%,
- 30 preferably at least about 60%, and more preferably at least about 70%, 80%, or 90% and most preferably at least about 95%, 96%, 97%, 98%, or 99% or more homologous to an amino acid sequence of the invention (e.g., an entire amino acid sequence selected from

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those having an even-numbered SEQ ID NO in the Sequence Listing). In another preferred embodiment, the protein is a full length *C. glutamicum* protein which is substantially homologous to an entire amino acid sequence of the invention (encoded by an open reading frame shown in the corresponding odd-numbered SEQ ID NOs in the

5 Sequence Listing (e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7....).

In another preferred embodiment, the isolated nucleic acid molecule is derived from *C. glutamicum* and encodes a protein (e.g., an HA fusion protein) which includes a biologically active domain which is at least about 50% or more homologous to one of the amino acid sequences of the invention (e.g., a sequence of one of the even-numbered

10 SEQ ID NOs in the Sequence Listing) and is able to participate in the repair or recombination of DNA, in the transposition of genetic material, in gene expression (i.e., the processes of transcription or translation), in protein folding, or in protein secretion in *Corynebacterium glutamicum*, or has one or more of the activities set forth in Table 1, and which also includes heterologous nucleic acid sequences encoding a heterologous

15 polypeptide or regulatory regions.

In another embodiment, the isolated nucleic acid molecule is at least 15 nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising a nucleotide sequence of the invention (e.g., a sequence of an odd-numbered SEQ ID NO in the Sequence Listing). Preferably, the isolated nucleic acid

20 molecule corresponds to a naturally-occurring nucleic acid molecule. More preferably, the isolated nucleic acid encodes a naturally-occurring *C. glutamicum* HA protein, or a biologically active portion thereof.

Another aspect of the invention pertains to vectors, e.g., recombinant expression vectors, containing the nucleic acid molecules of the invention, and host cells into which

25 such vectors have been introduced. In one embodiment, such a host cell is used to produce an HA protein by culturing the host cell in a suitable medium. The HA protein can be then isolated from the medium or the host cell.

Yet another aspect of the invention pertains to a genetically altered microorganism in which an HA gene has been introduced or altered. In one

30 embodiment, the genome of the microorganism has been altered by introduction of a nucleic acid molecule of the invention encoding wild-type or mutated HA sequence as a transgene. In another embodiment, an endogenous HA gene within the genome of the

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microorganism has been altered, *e.g.*, functionally disrupted, by homologous recombination with an altered HA gene. In another embodiment, an endogenous or introduced HA gene in a microorganism has been altered by one or more point mutations, deletions, or inversions, but still encodes a functional HA protein. In still
5 another embodiment, one or more of the regulatory regions (*e.g.*, a promoter, repressor, or inducer) of an HA gene in a microorganism has been altered (*e.g.*, by deletion, truncation, inversion, or point mutation) such that the expression of the HA gene is modulated. In a preferred embodiment, the microorganism belongs to the genus *Corynebacterium* or *Brevibacterium*, with *Corynebacterium glutamicum* being
10 particularly preferred. In a preferred embodiment, the microorganism is also utilized for the production of a desired compound, such as an amino acid, with lysine being particularly preferred.

In another aspect, the invention provides a method of identifying the presence or activity of *Corynebacterium diphtheriae* in a subject. This method includes detection of
15 one or more of the nucleic acid or amino acid sequences of the invention (*e.g.*, the sequences set forth in the Sequence Listing as SEQ ID NOs 1 through 440) in a subject, thereby detecting the presence or activity of *Corynebacterium diphtheriae* in the subject.

Still another aspect of the invention pertains to an isolated HA protein or a portion, *e.g.*, a biologically active portion, thereof. In a preferred embodiment, the
20 isolated HA protein or portion thereof can participate in the maintenance of homeostasis in *C. glutamicum*, or can perform a function involved in the adaptation of this microorganism to different environmental conditions. In another preferred embodiment, the isolated HA protein or portion thereof is sufficiently homologous to an amino acid
25 sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: in the Sequence Listing) such that the protein or portion thereof maintains the ability to participate in the maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions.

The invention also provides an isolated preparation of an HA protein. In preferred embodiments, the HA protein comprises an amino acid sequence of the
30 invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing). In another preferred embodiment, the invention pertains to an isolated full length protein which is substantially homologous to an entire amino acid sequence of the invention

(*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing) (encoded by an open reading frame set forth in a corresponding odd-numbered SEQ ID NO: of the Sequence Listing). In yet another embodiment, the protein is at least about 50%, preferably at least about 60%, and more preferably at least about 70%, 80%, or 90%,
5 and most preferably at least about 95%, 96%, 97%, 98%, or 99% or more homologous to an entire amino acid sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing). In other embodiments, the isolated HA protein comprises an amino acid sequence which is at least about 50% or more homologous to one of the amino acid sequences of the invention (*e.g.*, a sequence of an even-numbered
10 SEQ ID NO: of the Sequence Listing) and is able to participate in the maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions, or has one or more of the activities set forth in Table 1.

Alternatively, the isolated HA protein can comprise an amino acid sequence
15 which is encoded by a nucleotide sequence which hybridizes, *e.g.*, hybridizes under stringent conditions, or is at least about 50%, preferably at least about 60%, more preferably at least about 70%, 80%, or 90%, and even more preferably at least about 95%, 96%, 97%, 98%, or 99% or more homologous, to a nucleotide sequence of one of the even-numbered SEQ ID NOs set forth in the Sequence Listing. It is also preferred
20 that the preferred forms of HA proteins also have one or more of the HA bioactivities described herein.

The HA polypeptide, or a biologically active portion thereof, can be operatively linked to a non-HA polypeptide to form a fusion protein. In preferred embodiments, this fusion protein has an activity which differs from that of the HA protein alone. In other
25 preferred embodiments, this fusion protein participates in the maintenance of homeostasis in *C. glutamicum*, or performs a function involved in the adaptation of this microorganism to different environmental conditions. In particularly preferred embodiments, integration of this fusion protein into a host cell modulates production of a desired compound from the cell.

30 In another aspect, the invention provides methods for screening molecules which modulate the activity of an HA protein, either by interacting with the protein itself or a

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substrate or binding partner of the HA protein, or by modulating the transcription or translation of an HA nucleic acid molecule of the invention.

Another aspect of the invention pertains to a method for producing a fine chemical. This method involves the culturing of a cell containing a vector directing the expression of an HA nucleic acid molecule of the invention, such that a fine chemical is produced. In a preferred embodiment, this method further includes the step of obtaining a cell containing such a vector, in which a cell is transfected with a vector directing the expression of an HA nucleic acid. In another preferred embodiment, this method further includes the step of recovering the fine chemical from the culture. In a particularly preferred embodiment, the cell is from the genus *Corynebacterium* or *Brevibacterium*, or is selected from those strains set forth in Table 3.

Another aspect of the invention pertains to methods for modulating production of a molecule from a microorganism. Such methods include contacting the cell with an agent which modulates HA protein activity or HA nucleic acid expression such that a cell associated activity is altered relative to this same activity in the absence of the agent. In a preferred embodiment, the cell is modulated for one or more *C. glutamicum* processes involved in cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or enzymatic or proteolytic activities. The agent which modulates HA protein activity can be an agent which stimulates HA protein activity or HA nucleic acid expression. Examples of agents which stimulate HA protein activity or HA nucleic acid expression include small molecules, active HA proteins, and nucleic acids encoding HA proteins that have been introduced into the cell. Examples of agents which inhibit HA activity or expression include small molecules and antisense HA nucleic acid molecules.

Another aspect of the invention pertains to methods for modulating yields of a desired compound from a cell, involving the introduction of a wild-type or mutant HA gene into a cell, either maintained on a separate plasmid or integrated into the genome of the host cell. If integrated into the genome, such integration can be random, or it can take place by homologous recombination such that the native gene is replaced by the introduced copy, causing the production of the desired compound from the cell to be modulated. In a preferred embodiment, said yields are increased. In another preferred embodiment, said chemical is a fine chemical. In a particularly preferred embodiment,

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said fine chemical is an amino acid. In especially preferred embodiments, said amino acid is L-lysine.

Detailed Description of the Invention

5 The present invention provides HA nucleic acid and protein molecules which are involved in *C. glutamicum* cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or that have a *C. glutamicum* enzymatic or proteolytic activity. The molecules of the invention may be utilized in the modulation of production of fine chemicals from
10 microorganisms, such as *C. glutamicum*, either directly (e.g., where overexpression or optimization of activity of a protein involved in the production of a fine chemical (e.g., an enzyme) has a direct impact on the yield, production, and/or efficiency of production of a fine chemical from the modified *C. glutamicum*), or an indirect impact which nonetheless results in an increase of yield, production, and/or efficiency of production of
15 the desired compound (e.g., where modulation of the activity or number of copies of a *C. glutamicum* aromatic or aliphatic modification or degradation protein results in an increase in the viability of *C. glutamicum* cells, which in turn permits increased production in a large-scale culture setting). Aspects of the invention are further explicated below.

20

I. Fine Chemicals

 The term 'fine chemical' is art-recognized and includes molecules produced by an organism which have applications in various industries, such as, but not limited to, the pharmaceutical, agriculture, and cosmetics industries. Such compounds include
25 organic acids, such as tartaric acid, itaconic acid, and diaminopimelic acid, both proteinogenic and non-proteinogenic amino acids, purine and pyrimidine bases, nucleosides, and nucleotides (as described e.g. in Kuninaka, A. (1996) Nucleotides and related compounds, p. 561-612, in Biotechnology vol. 6, Rehm *et al.*, eds. VCH: Weinheim, and references contained therein), lipids, both saturated and unsaturated fatty
30 acids (e.g., arachidonic acid), diols (e.g., propane diol, and butane diol), carbohydrates (e.g., hyaluronic acid and trehalose), aromatic compounds (e.g., aromatic amines, vanillin, and indigo), vitamins and cofactors (as described in Ullmann's Encyclopedia of

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Industrial Chemistry, vol. A27, "Vitamins", p. 443-613 (1996) VCH: Weinheim and references therein; and Ong, A.S., Niki, E. & Packer, L. (1995) "Nutrition, Lipids, Health, and Disease" Proceedings of the UNESCO/Confederation of Scientific and Technological Associations in Malaysia, and the Society for Free Radical Research – Asia, held Sept. 1-3, 1994 at Penang, Malaysia, AOCS Press, (1995)), enzymes, polyketides (Cane *et al.* (1998) *Science* 282: 63-68), and all other chemicals described in Gutcho (1983) *Chemicals by Fermentation*, Noyes Data Corporation, ISBN: 0818805086 and references therein. The metabolism and uses of certain of these fine chemicals are further explicated below.

10

A. *Amino Acid Metabolism and Uses*

Amino acids comprise the basic structural units of all proteins, and as such are essential for normal cellular functioning in all organisms. The term "amino acid" is art-recognized. The proteinogenic amino acids, of which there are 20 species, serve as structural units for proteins, in which they are linked by peptide bonds, while the nonproteinogenic amino acids (hundreds of which are known) are not normally found in proteins (see Ulmann's *Encyclopedia of Industrial Chemistry*, vol. A2, p. 57-97 VCH: Weinheim (1985)). Amino acids may be in the D- or L- optical configuration, though L-amino acids are generally the only type found in naturally-occurring proteins.

Biosynthetic and degradative pathways of each of the 20 proteinogenic amino acids have been well characterized in both prokaryotic and eukaryotic cells (see, for example, Stryer, L. *Biochemistry*, 3rd edition, pages 578-590 (1988)). The 'essential' amino acids (histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, and valine), so named because they are generally a nutritional requirement due to the complexity of their biosyntheses, are readily converted by simple biosynthetic pathways to the remaining 11 'nonessential' amino acids (alanine, arginine, asparagine, aspartate, cysteine, glutamate, glutamine, glycine, proline, serine, and tyrosine). Higher animals do retain the ability to synthesize some of these amino acids, but the essential amino acids must be supplied from the diet in order for normal protein synthesis to occur.

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Aside from their function in protein biosynthesis, these amino acids are interesting chemicals in their own right, and many have been found to have various applications in the food, feed, chemical, cosmetics, agriculture, and pharmaceutical

- industries. Lysine is an important amino acid in the nutrition not only of humans, but also of monogastric animals such as poultry and swine. Glutamate is most commonly used as a flavor additive (mono-sodium glutamate, MSG) and is widely used throughout the food industry, as are aspartate, phenylalanine, glycine, and cysteine. Glycine, L-methionine and tryptophan are all utilized in the pharmaceutical industry. Glutamine, valine, leucine, isoleucine, histidine, arginine, proline, serine and alanine are of use in both the pharmaceutical and cosmetics industries. Threonine, tryptophan, and D/L-methionine are common feed additives. (Leuchtenberger, W. (1996) *Amino acids – technical production and use*, p. 466-502 in Rehm *et al.* (eds.) *Biotechnology* vol. 6, chapter 14a, VCH: Weinheim). Additionally, these amino acids have been found to be useful as precursors for the synthesis of synthetic amino acids and proteins, such as N-acetylcysteine, S-carboxymethyl-L-cysteine, (S)-5-hydroxytryptophan, and others described in Ulmann's Encyclopedia of Industrial Chemistry, vol. A2, p. 57-97, VCH: Weinheim, 1985.
- 15 The biosynthesis of these natural amino acids in organisms capable of producing them, such as bacteria, has been well characterized (for review of bacterial amino acid biosynthesis and regulation thereof, see Umbarger, H.E. (1978) *Ann. Rev. Biochem.* 47: 533-606). Glutamate is synthesized by the reductive amination of α -ketoglutarate, an intermediate in the citric acid cycle. Glutamine, proline, and arginine are each subsequently produced from glutamate. The biosynthesis of serine is a three-step process beginning with 3-phosphoglycerate (an intermediate in glycolysis), and resulting in this amino acid after oxidation, transamination, and hydrolysis steps. Both cysteine and glycine are produced from serine; the former by the condensation of homocysteine with serine, and the latter by the transfer of the side-chain β -carbon atom to tetrahydrofolate, in a reaction catalyzed by serine transhydroxymethylase. Phenylalanine, and tyrosine are synthesized from the glycolytic and pentose phosphate pathway precursors erythrose 4-phosphate and phosphoenolpyruvate in a 9-step biosynthetic pathway that differ only at the final two steps after synthesis of prephenate. Tryptophan is also produced from these two initial molecules, but its synthesis is an 11-step pathway. Tyrosine may also be synthesized from phenylalanine, in a reaction catalyzed by phenylalanine hydroxylase. Alanine, valine, and leucine are all biosynthetic products of pyruvate, the final product of glycolysis. Aspartate is formed

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from oxaloacetate, an intermediate of the citric acid cycle. Asparagine, methionine, threonine, and lysine are each produced by the conversion of aspartate. Isoleucine is formed from threonine. A complex 9-step pathway results in the production of histidine from 5-phosphoribosyl-1-pyrophosphate, an activated sugar.

- 5 Amino acids in excess of the protein synthesis needs of the cell cannot be stored, and are instead degraded to provide intermediates for the major metabolic pathways of the cell (for review see Stryer, L. *Biochemistry* 3rd ed. Ch. 21 "Amino Acid Degradation and the Urea Cycle" p. 495-516 (1988)). Although the cell is able to convert unwanted amino acids into useful metabolic intermediates, amino acid production is costly in
- 10 terms of energy, precursor molecules, and the enzymes necessary to synthesize them. Thus it is not surprising that amino acid biosynthesis is regulated by feedback inhibition, in which the presence of a particular amino acid serves to slow or entirely stop its own production (for overview of feedback mechanisms in amino acid biosynthetic pathways, see Stryer, L. *Biochemistry*, 3rd ed. Ch. 24: "Biosynthesis of Amino Acids and Heme" p.
- 15 575-600 (1988)). Thus, the output of any particular amino acid is limited by the amount of that amino acid present in the cell.

B. Vitamin, Cofactor, and Nutraceutical Metabolism and Uses

- Vitamins, cofactors, and nutraceuticals comprise another group of molecules
- 20 which the higher animals have lost the ability to synthesize and so must ingest, although they are readily synthesized by other organisms such as bacteria. These molecules are either bioactive substances themselves, or are precursors of biologically active substances which may serve as electron carriers or intermediates in a variety of metabolic pathways. Aside from their nutritive value, these compounds also have
- 25 significant industrial value as coloring agents, antioxidants, and catalysts or other processing aids. (For an overview of the structure, activity, and industrial applications of these compounds, see, for example, Ullman's *Encyclopedia of Industrial Chemistry*, "Vitamins" vol. A27, p. 443-613, VCH: Weinheim, 1996.) The term "vitamin" is art-
- 30 recognized, and includes nutrients which are required by an organism for normal functioning, but which that organism cannot synthesize by itself. The group of vitamins may encompass cofactors and nutraceutical compounds. The language "cofactor" includes nonproteinaceous compounds required for a normal enzymatic activity to

occur. Such compounds may be organic or inorganic; the cofactor molecules of the invention are preferably organic. The term "nutraceutical" includes dietary supplements having health benefits in plants and animals, particularly humans. Examples of such molecules are vitamins, antioxidants, and also certain lipids (e.g., polyunsaturated fatty acids).

The biosynthesis of these molecules in organisms capable of producing them, such as bacteria, has been largely characterized (Ullman's Encyclopedia of Industrial Chemistry, "Vitamins" vol. A27, p. 443-613, VCH: Weinheim, 1996; Michal, G. (1999) Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology, John Wiley & Sons; Ong, A.S., Niki, E. & Packer, L. (1995) "Nutrition, Lipids, Health, and Disease" Proceedings of the UNESCO/Confederation of Scientific and Technological Associations in Malaysia, and the Society for Free Radical Research – Asia, held Sept. 1-3, 1994 at Penang, Malaysia, AOCS Press: Champaign, IL X, 374 S).

Thiamin (vitamin B₁) is produced by the chemical coupling of pyrimidine and thiazole moieties. Riboflavin (vitamin B₂) is synthesized from guanosine-5'-triphosphate (GTP) and ribose-5'-phosphate. Riboflavin, in turn, is utilized for the synthesis of flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD). The family of compounds collectively termed 'vitamin B₆' (e.g., pyridoxine, pyridoxamine, pyridoxal-5'-phosphate, and the commercially used pyridoxin hydrochloride) are all derivatives of the common structural unit, 5-hydroxy-6-methylpyridine. Pantothenate (pantothenic acid, (R)-(+)-N-(2,4-dihydroxy-3,3-dimethyl-1-oxobutyl)-β-alanine) can be produced either by chemical synthesis or by fermentation. The final steps in pantothenate biosynthesis consist of the ATP-driven condensation of β-alanine and pantoic acid. The enzymes responsible for the biosynthesis steps for the conversion to pantoic acid, to β-alanine and for the condensation to pantothenic acid are known. The metabolically active form of pantothenate is Coenzyme A, for which the biosynthesis proceeds in 5 enzymatic steps. Pantothenate, pyridoxal-5'-phosphate, cysteine and ATP are the precursors of Coenzyme A. These enzymes not only catalyze the formation of pantothenate, but also the production of (R)-pantoic acid, (R)-pantolacton, (R)-panthanol (provitamin B₅), pantetheine (and its derivatives) and coenzyme A.

Biotin biosynthesis from the precursor molecule pimeloyl-CoA in microorganisms has been studied in detail and several of the genes involved have been

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identified. Many of the corresponding proteins have been found to also be involved in Fe-cluster synthesis and are members of the *nifS* class of proteins. Lipic acid is derived from octanoic acid, and serves as a coenzyme in energy metabolism, where it becomes part of the pyruvate dehydrogenase complex and the α -ketoglutarate

5 dehydrogenase complex. The folates are a group of substances which are all derivatives of folic acid, which in turn is derived from L-glutamic acid, p-amino-benzoic acid and 6-methylpterin. The biosynthesis of folic acid and its derivatives, starting from the metabolism intermediates guanosine-5'-triphosphate (GTP), L-glutamic acid and p-amino-benzoic acid has been studied in detail in certain microorganisms.

10 Corrinoids (such as the cobalamines and particularly vitamin B₁₂) and porphyrines belong to a group of chemicals characterized by a tetrapyrrole ring system. The biosynthesis of vitamin B₁₂ is sufficiently complex that it has not yet been completely characterized, but many of the enzymes and substrates involved are now known. Nicotinic acid (nicotinate), and nicotinamide are pyridine derivatives which are
15 also termed 'niacin'. Niacin is the precursor of the important coenzymes NAD (nicotinamide adenine dinucleotide) and NADP (nicotinamide adenine dinucleotide phosphate) and their reduced forms.

The large-scale production of these compounds has largely relied on cell-free chemical syntheses, though some of these chemicals have also been produced by large-
20 scale culture of microorganisms, such as riboflavin, Vitamin B₆, pantothenate, and biotin. Only Vitamin B₁₂ is produced solely by fermentation, due to the complexity of its synthesis. *In vitro* methodologies require significant inputs of materials and time, often at great cost.

25 C. Purine, Pyrimidine, Nucleoside and Nucleotide Metabolism and Uses

Purine and pyrimidine metabolism genes and their corresponding proteins are important targets for the therapy of tumor diseases and viral infections. The language "purine" or "pyrimidine" includes the nitrogenous bases which are constituents of nucleic acids, co-enzymes, and nucleotides. The term "nucleotide" includes the basic
30 structural units of nucleic acid molecules, which are comprised of a nitrogenous base, a pentose sugar (in the case of RNA, the sugar is ribose; in the case of DNA, the sugar is D-deoxyribose), and phosphoric acid. The language "nucleoside" includes molecules

which serve as precursors to nucleotides, but which are lacking the phosphoric acid moiety that nucleotides possess. By inhibiting the biosynthesis of these molecules, or their mobilization to form nucleic acid molecules, it is possible to inhibit RNA and DNA synthesis; by inhibiting this activity in a fashion targeted to cancerous cells, the ability of tumor cells to divide and replicate may be inhibited. Additionally, there are nucleotides which do not form nucleic acid molecules, but rather serve as energy stores (i.e., AMP) or as coenzymes (i.e., FAD and NAD).

Several publications have described the use of these chemicals for these medical indications, by influencing purine and/or pyrimidine metabolism (e.g. Christopherson, R.I. and Lyons, S.D. (1990) "Potent inhibitors of *de novo* pyrimidine and purine biosynthesis as chemotherapeutic agents." *Med. Res. Reviews* 10: 505-548). Studies of enzymes involved in purine and pyrimidine metabolism have been focused on the development of new drugs which can be used, for example, as immunosuppressants or anti-proliferants (Smith, J.L., (1995) "Enzymes in nucleotide synthesis." *Curr. Opin. Struct. Biol.* 5: 752-757; (1995) *Biochem Soc. Transact.* 23: 877-902). However, purine and pyrimidine bases, nucleosides and nucleotides have other utilities: as intermediates in the biosynthesis of several fine chemicals (e.g., thiamine, S-adenosyl-methionine, folates, or riboflavin), as energy carriers for the cell (e.g., ATP or GTP), and for chemicals themselves, commonly used as flavor enhancers (e.g., IMP or GMP) or for several medicinal applications (see, for example, Kuninaka, A. (1996) *Nucleotides and Related Compounds in Biotechnology* vol. 6, Rehm *et al.*, eds. VCH: Weinheim, p. 561-612). Also, enzymes involved in purine, pyrimidine, nucleoside, or nucleotide metabolism are increasingly serving as targets against which chemicals for crop protection, including fungicides, herbicides and insecticides, are developed.

The metabolism of these compounds in bacteria has been characterized (for reviews see, for example, Zalkin, H. and Dixon, J.E. (1992) "*de novo* purine nucleotide biosynthesis", in: *Progress in Nucleic Acid Research and Molecular Biology*, vol. 42, Academic Press, p. 259-287; and Michal, G. (1999) "Nucleotides and Nucleosides", Chapter 8 in: *Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology*, Wiley: New York). Purine metabolism has been the subject of intensive research, and is essential to the normal functioning of the cell. Impaired purine metabolism in higher animals can cause severe disease, such as gout. Purine nucleotides are synthesized from

ribose-5-phosphate, in a series of steps through the intermediate compound inosine-5'-phosphate (IMP), resulting in the production of guanosine-5'-monophosphate (GMP) or adenosine-5'-monophosphate (AMP), from which the triphosphate forms utilized as nucleotides are readily formed. These compounds are also utilized as energy stores, so
5 their degradation provides energy for many different biochemical processes in the cell. Pyrimidine biosynthesis proceeds by the formation of uridine-5'-monophosphate (UMP) from ribose-5-phosphate. UMP, in turn, is converted to cytidine-5'-triphosphate (CTP). The deoxy- forms of all of these nucleotides are produced in a one step reduction
10 reaction from the diphosphate ribose form of the nucleotide to the diphosphate deoxyribose form of the nucleotide. Upon phosphorylation, these molecules are able to participate in DNA synthesis.

D. Trehalose Metabolism and Uses

Trehalose consists of two glucose molecules, bound in α , α -1,1 linkage. It is
15 commonly used in the food industry as a sweetener, an additive for dried or frozen foods, and in beverages. However, it also has applications in the pharmaceutical, cosmetics and biotechnology industries (see, for example, Nishimoto *et al.*, (1998) U.S. Patent No. 5,759,610; Singer, M.A. and Lindquist, S. (1998) *Trends Biotech.* 16: 460-467; Paiva, C.L.A. and Panek, A.D. (1996) *Biotech. Ann. Rev.* 2: 293-314; and
20 Shiosaka, M. (1997) *J. Japan* 172: 97-102). Trehalose is produced by enzymes from many microorganisms and is naturally released into the surrounding medium, from which it can be collected using methods known in the art.

II. Maintenance of Homeostasis in *C. glutamicum* and Environmental Adaptation

25 The metabolic and other biochemical processes by which cells function are sensitive to environmental conditions such as temperature, pressure, solute concentration, and availability of oxygen. When one or more such environmental condition is perturbed or altered in a fashion that is incompatible with the normal functioning of these cellular processes, the cell must act to maintain an intracellular
30 environment which will permit them to occur despite the hostile extracellular environment. Gram positive bacterial cells, such as *C. glutamicum* cells, have a number of mechanisms by which internal homeostasis may be maintained despite unfavorable

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extracellular conditions. These include a cell wall, proteins which are able to degrade possibly toxic aromatic and aliphatic compounds, mechanisms of proteolysis whereby misfolded or misregulated proteins may be rapidly destroyed, and catalysts which permit intracellular reactions to occur which would not normally take place under the
5 conditions optimal for bacterial growth.

Aside from merely surviving in a hostile environment, bacterial cells (*e.g. C. glutamicum* cells) are also frequently able to adapt such that they are able to take advantage of such conditions. For example, cells in an environment lacking desired carbon sources may be able to adapt to growth on a less-suitable carbon source. Also,
10 cells may be able to utilize less desirable inorganic compounds when the commonly utilized ones are unavailable. *C. glutamicum* cells possess a number of genes which permit them to adapt to utilize inorganic and organic molecules which they would normally not encounter under optimal growth conditions as nutrients and precursors for metabolism. Aspects of cellular processes involved in homeostasis and adaptation are
15 further explicated below.

A. Modification and Degradation of Aromatic and Aliphatic Compounds

Bacterial cells are routinely exposed to a variety of aromatic and aliphatic compounds in nature. Aromatic compounds are organic molecules having a cyclic ring
20 structure, while aliphatic compounds are organic molecules having open chain structures rather than ring structures. Such compounds may arise as by-products of industrial processes (*e.g.*, benzene or toluene), but may also be produced by certain microorganisms (*e.g.*, alcohols). Many of these compounds are toxic to cells, particularly the aromatic compounds, which are highly reactive due to the high-energy ring structure. Thus, certain
25 bacteria have developed mechanisms by which they are able to modify or degrade these compounds such that they are no longer hazardous to the cell. Cells may possess enzymes that are able to, for example, hydroxylate, isomerize, or methylate aromatic or aliphatic compounds such that they are either rendered less toxic, or such that the modified form is able to be processed by standard cellular waste and degradation pathways. Also, cells may
30 possess enzymes which are able to specifically degrade one or more such potentially hazardous substance, thereby protecting the cell. Principles and examples of these types of modification and degradation processes in bacteria are described in several publications,

e.g., Sahm, H. (1999) "Procaryotes in Industrial Production" in Lengeler, J.W. *et al.*, eds. *Biology of the Procaryotes*, Thieme Verlag: Stuttgart; and Schlegel, H.G. (1992) *Allgemeine Mikrobiologie*, Thieme: Stuttgart).

Aside from simply inactivating hazardous aromatic or aliphatic compounds, many
5 bacteria have evolved to be able to utilize these compounds as carbon sources for continued
metabolism when the preferred carbon sources of the cell are not available. For example,
Pseudomonas strains able to utilize toluene, benzene, and 1,10-dichlorodecane as carbon
sources are known (Chang, B.V. *et al.* (1997) *Chemosphere* 35(12): 2807-2815; Wischnak,
C. *et al.* (1998) *Appl. Environ. Microbiol.* 64(9): 3507-3511; Churchill, S.A. *et al.* (1999)
10 *Appl. Environ. Microbiol.* 65(2): 549-552). There are similar examples from many other
bacterial species which are known in the art.

The ability of certain bacteria to modify or degrade aromatic and aliphatic
compounds has begun to be exploited. Petroleum is a complex mixture of chemicals which
includes aliphatic molecules and aromatic compounds. By applying bacteria having the
15 ability to degrade or modify these toxic compounds to an oil spill, for example, it is possible
to eliminate much of the environmental damage with high efficiency and low cost (see, for
example, Smith, M.R. (1990) "The biodegradation of aromatic hydrocarbons by bacteria"
Biodegradation 1(2-3): 191-206; and Suyama, T. *et al.* (1998) "Bacterial isolates degrading
aliphatic polycarbonates," *FEMS Microbiol. Lett.* 161(2): 255-261).

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B. Metabolism of Inorganic Compounds

Cells (e.g., bacterial cells) contain large quantities of different molecules, such as
water, inorganic ions, and organic substances (e.g., proteins, sugars, and other
macromolecules). The bulk of the mass of a typical cell consists of only 4 types of atoms:
25 carbon, oxygen, hydrogen, and nitrogen. Although they represent a smaller percentage of
the content of a cell, inorganic substances are equally as important to the proper functioning
of the cell. Such molecules include phosphorous, sulfur, calcium, magnesium, iron, zinc,
manganese, copper, molybdenum, tungsten, and cobalt. Many of these compounds are
critical for the construction of important molecules, such as nucleotides (phosphorous) and
30 amino acids (nitrogen and sulfur). Others of these inorganic ions serve as cofactors for
enzymic reactions or contribute to osmotic pressure. All such molecules must be taken up
by the bacterium from the surrounding environment.

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For each of these inorganic compounds it is desirable for the bacterium to take up the form which can be most readily used by the standard metabolic machinery of the cell. However, the bacterium may encounter environments in which these preferred forms are not readily available. In order to survive under these circumstances, it is important for bacteria

5 to have additional biochemical mechanisms which are able to convert less metabolically active but readily available forms of these inorganic compounds to ones which may be used in cellular metabolism. Bacteria frequently possess a number of genes encoding enzymes for this purpose, which are not expressed unless the desired inorganic species are not available. Thus, these genes for the metabolism of various inorganic compounds serve as

10 another tool which bacteria may use to adapt to suboptimal environmental conditions.

After carbon, the most important element in the cell is nitrogen. A typical bacterial cell contains between 12-15% nitrogen. It is a constituent of amino acids and nucleotides, as well as many other important molecules in the cell. Further, nitrogen may serve as a substitute for oxygen as a terminal electron acceptor in energy metabolism. Good sources

15 of nitrogen include many organic and inorganic compounds, such ammonia gas or ammonia salts (*e.g.*, NH_4Cl , $(\text{NH}_4)_2\text{SO}_4$, or NH_4OH), nitrates, urea, amino acids, or complex nitrogen sources like corn steep liquor, soy bean flour, soy bean protein, yeast extract, meat extract, etc. Ammonia nitrogen is fixed by the action of particular enzymes: glutamate dehydrogenase, glutamine synthase, and glutamine-2-oxoglutarate aminotransferase. The

20 transfer of amino-nitrogen from one organic molecule to another is accomplished by the aminotransferases, a class of enzymes which transfer one amino group from an alpha-amino acid to an alpha-keto acid. Nitrate may be reduced via nitrate reductase, nitrite reductase, and further redox enzymes until it is converted to molecular nitrogen or ammonia, which may be readily utilized by the cell in standard metabolic pathways.

25 Phosphorous is typically found intracellularly in both organic and inorganic forms, and may be taken up by the cell in either of these forms as well, though most microorganisms preferentially take up inorganic phosphate. The conversion of organic phosphate to a form which the cell can utilize requires the action of phosphatases (*e.g.*, phytases, which hydrolyze phyate-yielding phosphate and inositol derivatives). Phosphate

30 is a key element in the synthesis of nucleic acids, and also has a significant role in cellular energy metabolism (*e.g.*, in the synthesis of ATP, ADP, and AMP).

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Sulfur is a requirement for the synthesis of amino acids (e.g., methionine and cysteine), vitamins (e.g., thiamine, biotin, and lipoic acid) and iron sulfur proteins. Bacteria obtain sulfur primarily from inorganic sulfate, though thiosulfate, sulfite, and sulfide are also commonly utilized. Under conditions where these compounds may not be readily available, many bacteria express genes which enable them to utilize sulfonate compounds such as 2-aminosulfonate (taurine) (Kertesz, M.A. (1993) "Proteins induced by sulfate limitation in *Escherichia coli*, *Pseudomonas putida*, or *Staphylococcus aureus*." *J. Bacteriol.* 175: 1187-1190).

Other inorganic atoms, e.g., metal or calcium ions, are also critical for the viability of cells. Iron, for example, plays a key role in redox reactions and is a cofactor of iron-sulfur proteins, heme proteins, and cytochromes. The uptake of iron into bacterial cells may be accomplished by the action of siderophores, chelating agents which bind extracellular iron ions and translocate them to the interior of the cell. For reference on the metabolism of iron and other inorganic compounds, see: Lengeler *et al.* (1999) *Biology of Prokaryotes*, Thieme Verlag: Stuttgart; Neidhardt, F.C. *et al.*, eds. *Escherichia coli* and *Salmonella*. ASM Press: Washington, D.C.; Sonenshein, A.L. *et al.*, eds. (1997) *Bacillus subtilis* and Other Gram-Positive Bacteria, ASM Press: Washington, D.C.; Voet, D. and Voet, J.G. (1992) *Biochemie*, VCH: Weinheim; Brock, T.D. and Madigan, M.T. (1991) *Biology of Microorganisms*, 6th ed. Prentice Hall: Englewood Cliffs, p. 267-269; Rhodes, P.M. and Stanbury, P.F. *Applied Microbial Physiology – A Practical Approach*, Oxford Univ. Press: Oxford.

C. Enzymes and Proteolysis

The intracellular conditions for which bacteria such as *C. glutamicum* are optimized are frequently not conditions under which many biochemical reactions would normally take place. In order to make such reactions proceed under physiological conditions, cells utilize enzymes. Enzymes are proteinaceous biological catalysts, spatially orienting reacting molecules or providing a specialized environment such that the energy barrier to a biochemical reaction is lowered. Different enzymes catalyze different reactions, and each enzyme may be the subject of transcriptional, translational, or posttranslational regulation such that the reaction will only take place under appropriate conditions and at specified times. Enzymes may contribute to the

degradation (*e.g.*, the proteases), synthesis (*e.g.*, the synthases), or modification (*e.g.*, transferases or isomerases) of compounds, all of which enable the production of necessary compounds within the cell. This, in turn, contributes to the maintenance of cellular homeostasis.

5 However, the fact that enzymes are optimized for activity under the physiological conditions at which the bacterium is most viable means that when environmental conditions are perturbed, there is a significant possibility that enzyme activity will also be perturbed. For example, changes in temperature may result in aberrantly folded proteins, and the same is true for changes of pH – protein folding is
10 largely dependent on electrostatic and hydrophobic interactions of amino acids within the polypeptide chain, so any alteration to the charges on individual amino acids (as might be brought about by a change in cellular pH) may have a profound effect on the ability of the protein to correctly fold. Changes in temperature effectively change the amount of kinetic energy that the polypeptide molecule possesses, which affects the
15 ability of the polypeptide to settle into a correctly folded, energetically stable configuration. Misfolded proteins may be harmful to the cell for two reasons. First, the aberrantly folded protein may have a similarly aberrant activity, or no activity whatsoever. Second, misfolded proteins may lack the conformational regions necessary for proper regulation by other cellular systems and thus may continue to be active but in
20 an uncontrolled fashion.

 The cell has a mechanism by which misfolded enzymes and regulatory proteins may be rapidly destroyed before any damage occurs to the cell: proteolysis. Proteins such as those of the *la/lon* family and those of the *Clp* family specifically recognize and degrade misfolded proteins (see, *e.g.*, Sherman, M.Y., Goldberg, A.L. (1999) *EXS* 77:
25 57-78 and references therein and Porankiewicz J. (1999) *Molec. Microbiol.* 32(3): 449-58, and references therein; Neidhardt, F.C., *et al.* (1996) *E. coli* and *Salmonella*, ASM Press: Washington, D.C. and references therein; and Pritchard, G.G., and Coolbear, T. (1993) *FEMS Microbiol. Rev.* 12(1-3): 179-206 and references therein). These enzymes bind to misfolded or unfolded proteins and degrade them in an ATP-dependent manner.
30 Proteolysis thus serves as an important mechanism employed by the cell to prevent damage to normal cellular functions upon environmental changes, and it further permits

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cells to survive under conditions and in environments which would otherwise be toxic due to misregulated and/or aberrant enzyme or regulatory activity.

Proteolysis also has important functions in the cell under optimal environmental conditions. Within normal metabolic processes, proteases aid in the hydrolysis of peptide bonds, in the catabolism of complex molecules to provide necessary degradation products, and in protein modification. Secreted proteases play an important role in the catabolism of external nutrients even prior to the entry of these compounds into the cell. Further, proteolytic activity itself may serve regulatory functions; sporulation in *B. subtilis* and cell cycle progression in *Caulobacter* spp. are known to be regulated by key proteolytic events in each of these species (Gottesman, S. (1999) *Curr. Opin. Microbiol.* 2(2): 142-147). Thus, proteolytic processes are key for cellular survival under both suboptimal and optimal environmental conditions, and contribute to the overall maintenance of homeostasis in cells.

15 D. Cell Wall Production and Rearrangements

While the biochemical machinery of the cell may be able to readily adapt to different and possibly unfavorable environments, cells still require a general mechanism by which they may be protected from the environment. For many bacteria, the cell wall affords such protection, and also plays roles in adhesion, cell growth and division, and transport of desired solutes and waste materials.

In order to function, cells require intracellular concentrations of metabolites and other molecules that are substantially higher than those of the surrounding media. Since these metabolites are largely prevented from leaving the cell due to the presence of the hydrophobic membrane, the tendency of the system is for water molecules to enter the cell from the external medium such that the interior concentrations of solutes match the exterior concentrations. Water molecules are readily able to cross the cellular membrane, and this membrane is not able to withstand the resulting swelling and pressure, which may lead to osmotic lysis of the cell. The rigidity of the cell wall greatly improves the ability of the cell to tolerate these pressures, and offers a further barrier to the unwanted diffusion of these metabolites and desired solutes from the cell. Similarly, the cell wall also serves to prevent unwanted material from entering the cell.

The cell wall also participates in a number of other cellular processes, such as adhesion and cell growth and division. Due to the fact that the cell wall completely surrounds the cell, any interaction of the cell with its surroundings must be mediated by the cell wall. Thus, the cell wall must participate in any adherence of the cell to other cells and to desired surfaces. Further, the cell cannot grow or divide without concomitant changes in the cell wall. Since the protection that the wall affords requires its presence during growth, morphogenesis and multiplication, one of the key steps in cell division is cell wall synthesis within the cell such that a new cell divides from the old. Thus, frequently cell wall biosynthesis is regulated in tandem with cell growth and cell division (see, e.g., Sonenshein, A.L. et al, eds. (1993) *Bacillus subtilis* and Other Gram-Positive Bacteria, ASM: Washington, D.C.).

The structure of the cell wall varies between gram-positive and gram-negative bacteria. However, in both types, the fundamental structural unit of the wall remains similar: an overlapping lattice of two polysaccharides, N-acetyl glucosamine (NAG) and N-acetyl muramic acid (NAM) which are cross-linked by amino acids (most commonly L-alanine, D-glutamate, diaminopimelic acid, and D-alanine), termed 'peptidoglycan'. The processes involved in the synthesis of the cell wall are known (see, e.g., Michal, G., ed. (1999) *Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology*, Wiley: New York).

In gram-negative bacteria, the inner cellular membrane is coated by a single-layered peptidoglycan (approximately 10 nm thick), termed the murein-sacculus. This peptidoglycan structure is very rigid, and its structure determines the shape of the organism. The outer surface of the murein-sacculus is covered with an outer membrane, containing porins and other membrane proteins, phospholipids, and lipopolysaccharides. To maintain a tight association with the outer membrane, the gram-negative cell wall also has interspersed lipid molecules which serve to anchor it to the surrounding membrane.

In gram-positive bacteria, such as *Corynebacterium glutamicum*, the cytoplasmic membrane is covered by a multi-layered peptidoglycan, which ranges from 20-80 nm in thickness (see, e.g., Lengeler *et al.* (1999) *Biology of Prokaryotes* Thieme Verlag: Stuttgart, p. 913-918, p. 875-899, and p. 88-109 and references therein). The gram-positive cell wall also contains teichoic acid, a polymer of glycerol or ribitol linked through phosphate groups. Teichoic acid is also able to associate with amino acids, and forms covalent bonds with

muramic acid. Also present in the cell wall may be lipoteichoic acids and teichuronic acids. If present, cellular surface structures such as flagella or capsules will be anchored in this layer as well.

5 III. Elements and Methods of the Invention

The present invention is based, at least in part, on the discovery of novel molecules, referred to herein as HA nucleic acid and protein molecules, which participate in the maintenance of homeostasis in *C. glutamicum*, or which perform a function involved in the adaptation of this microorganism to different environmental
10 conditions. In one embodiment, the HA molecules participate in *C. glutamicum* cell wall biosynthesis or rearrangements, in the metabolism of inorganic compounds, in the modification or degradation of aromatic or aliphatic compounds, or have an enzymatic or proteolytic activity. In a preferred embodiment, the activity of the HA molecules of the present invention with regard to *C. glutamicum* cell wall biosynthesis or
15 rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or enzymatic or proteolytic activity has an impact on the production of a desired fine chemical by this organism. In a particularly preferred embodiment, the HA molecules of the invention are modulated in activity, such that the *C. glutamicum* cellular processes in which the HA molecules participate (*e.g.*, *C.*
20 *glutamicum* cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or enzymatic or proteolytic activity) are also altered in activity, resulting either directly or indirectly in a modulation of the yield, production, and/or efficiency of production of a desired fine chemical by *C. glutamicum*.

25 The language, "HA protein" or "HA polypeptide" includes proteins which participate in a number of cellular processes related to *C. glutamicum* homeostasis or the ability of *C. glutamicum* cells to adapt to unfavorable environmental conditions. For example, an HA protein may be involved in *C. glutamicum* cell wall biosynthesis or rearrangements, in the metabolism of inorganic compounds in *C. glutamicum*, in the
30 modification or degradation of aromatic or aliphatic compounds in *C. glutamicum*, or have a *C. glutamicum* enzymatic or proteolytic activity. Examples of HA proteins include those encoded by the HA genes set forth in Table 1 and by the odd-numbered

SEQ ID NOs. The terms "HA gene" or "HA nucleic acid sequence" include nucleic acid sequences encoding an HA protein, which consist of a coding region and also corresponding untranslated 5' and 3' sequence regions. Examples of HA genes include those set forth in Table 1. The terms "production" or "productivity" are art-recognized and include the concentration of the fermentation product (for example, the desired fine chemical) formed within a given time and a given fermentation volume (*e.g.*, kg product per hour per liter). The term "efficiency of production" includes the time required for a particular level of production to be achieved (for example, how long it takes for the cell to attain a particular rate of output of a fine chemical). The term "yield" or "product/carbon yield" is art-recognized and includes the efficiency of the conversion of the carbon source into the product (*i.e.*, fine chemical). This is generally written as, for example, kg product per kg carbon source. By increasing the yield or production of the compound, the quantity of recovered molecules, or of useful recovered molecules of that compound in a given amount of culture over a given amount of time is increased. The terms "biosynthesis" or a "biosynthetic pathway" are art-recognized and include the synthesis of a compound, preferably an organic compound, by a cell from intermediate compounds in what may be a multistep and highly regulated process. The terms "degradation" or a "degradation pathway" are art-recognized and include the breakdown of a compound, preferably an organic compound, by a cell to degradation products (generally speaking, smaller or less complex molecules) in what may be a multistep and highly regulated process. The language "metabolism" is art-recognized and includes the totality of the biochemical reactions that take place in an organism. The metabolism of a particular compound, then, (*e.g.*, the metabolism of an amino acid such as glycine) comprises the overall biosynthetic, modification, and degradation pathways in the cell related to this compound. The term "homeostasis" is art-recognized and includes all of the mechanisms utilized by a cell to maintain a constant intracellular environment despite the prevailing extracellular environmental conditions. A non-limiting example of such processes is the utilization of a cell wall to prevent osmotic lysis due to high intracellular solute concentrations. The term "adaptation" or "adaptation to an environmental condition" is art-recognized and includes mechanisms utilized by the cell to render the cell able to survive under nonpreferred environmental conditions (generally speaking, those environmental conditions in which one or more

5 favored nutrients are absent, or in which an environmental condition such as temperature, pH, osmolarity, oxygen percentage and the like fall outside of the optimal survival range of the cell). Many cells, including *C. glutamicum* cells, possess genes encoding proteins which are expressed under such environmental conditions and which permit continued growth in such suboptimal conditions.

In another embodiment, the HA molecules of the invention are capable of modulating the production of a desired molecule, such as a fine chemical, in a microorganism such as *C. glutamicum*. There are a number of mechanisms by which the alteration of an HA protein of the invention may directly affect the yield, production, and/or efficiency of production of a fine chemical from a *C. glutamicum* strain incorporating such an altered protein. For example, by engineering enzymes which modify or degrade aromatic or aliphatic compounds such that these enzymes are increased or decreased in activity or number, it may be possible to modulate the production of one or more fine chemicals which are the modification or degradation products of these compounds. Similarly, enzymes involved in the metabolism of inorganic compounds provide key molecules (e.g. phosphorous, sulfur, and nitrogen molecules) for the biosynthesis of such fine chemicals as amino acids, vitamins, and nucleic acids. By altering the activity or number of these enzymes in *C. glutamicum*, it may be possible to increase the conversion of these inorganic compounds (or to use alternate inorganic compounds) to thus permit improved rates of incorporation of inorganic atoms into these fine chemicals. Genetic engineering of *C. glutamicum* enzymes involved in general cellular processes may also directly improve fine chemical production, since many of these enzymes directly modify fine chemicals (e.g., amino acids) or the enzymes which are involved in fine chemical synthesis or secretion. Modulation of the activity or number of cellular proteases may also have a direct effect on fine chemical production, since many proteases may degrade fine chemicals or enzymes involved in fine chemical production or breakdown.

Further, the aforementioned enzymes which participate in aromatic/aliphatic compound modification or degradation, general biocatalysis, inorganic compound metabolism or proteolysis are each themselves fine chemicals, desirable for their activity in various *in vitro* industrial applications. By altering the number of copies of the gene for one or more of these enzymes in *C. glutamicum* it may be possible to increase the

number of these proteins produced by the cell, thereby increasing the potential yield or efficiency of production of these proteins from large-scale *C. glutamicum* or related bacterial cultures.

The alteration of an HA protein of the invention may also indirectly affect the
5 yield, production, and/or efficiency of production of a fine chemical from a *C. glutamicum* strain incorporating such an altered protein. For example, by modulating the activity and/or number of those proteins involved in the construction or rearrangement of the cell wall, it may be possible to modify the structure of the cell wall itself such that the cell is able to better withstand the mechanical and other stresses
10 present during large-scale fermentative culture. Also, large-scale growth of *C. glutamicum* requires significant cell wall production. Modulation of the activity or number of cell wall biosynthetic or degradative enzymes may allow more rapid rates of cell wall biosynthesis, which in turn may permit increased growth rates of this microorganism in culture and thereby increase the number of cells producing the desired
15 fine chemical.

By modifying the HA enzymes of the invention, one may also indirectly impact the yield, production, or efficiency of production of one or more fine chemicals from *C. glutamicum*. For example, many of the general enzymes in *C. glutamicum* may have a significant impact on global cellular processes (e.g., regulatory processes) which in turn
20 have a significant effect on fine chemical metabolism. Similarly, proteases, enzymes which modify or degrade possibly toxic aromatic or aliphatic compounds, and enzymes which promote the metabolism of inorganic compounds all serve to increase the viability of *C. glutamicum*. The proteases aid in the selective removal of misfolded or misregulated proteins, such as those that might occur under the relatively stressful
25 environmental conditions encountered during large-scale fermentor culture. By altering these proteins, it may be possible to further enhance this activity and to improve the viability of *C. glutamicum* in culture. The aromatic/aliphatic modification or degradation proteins not only serve to detoxify these waste compounds (which may be encountered as impurities in culture medium or as waste products from cells
30 themselves), but also to permit the cells to utilize alternate carbon sources if the optimal carbon source is limiting in the culture. By increasing their number and/or activity, the survival of *C. glutamicum* cells in culture may be enhanced. The inorganic metabolism

- proteins of the invention supply the cell with inorganic molecules required for all protein and nucleotide (among others) synthesis, and thus are critical for the overall viability of the cell. An increase in the number of viable cells producing one or more desired fine chemicals in large-scale culture should result in a concomitant increase in the yield,
- 5 production, and/or efficiency of production of the fine chemical in the culture.

- The isolated nucleic acid sequences of the invention are contained within the genome of a *Corynebacterium glutamicum* strain available through the American Type Culture Collection, given designation ATCC 13032. The nucleotide sequence of the isolated *C. glutamicum* HA DNAs and the predicted amino acid sequences of the *C.*
- 10 *glutamicum* HA proteins are shown in the Sequence Listing as odd-numbered SEQ ID NOs and even-numbered SEQ ID NOs, respectively, respectively. Computational analyses were performed which classified and/or identified these nucleotide sequences as sequences which encode proteins that participate in *C. glutamicum* cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or
- 15 degradation of aromatic or aliphatic compounds, or that have a *C. glutamicum* enzymatic or proteolytic activity.

- The present invention also pertains to proteins which have an amino acid sequence which is substantially homologous to an amino acid sequence of the invention (*e.g.*, the sequence of an even-numbered SEQ ID NO of the Sequence Listing). As used
- 20 herein, a protein which has an amino acid sequence which is substantially homologous to a selected amino acid sequence is least about 50% homologous to the selected amino acid sequence, *e.g.*, the entire selected amino acid sequence. A protein which has an amino acid sequence which is substantially homologous to a selected amino acid sequence can also be least about 50-60%, preferably at least about 60-70%, and more
- 25 preferably at least about 70-80%, 80-90%, or 90-95%, and most preferably at least about 96%, 97%, 98%, 99% or more homologous to the selected amino acid sequence.

- The HA protein or a biologically active portion or fragment thereof of the invention can participate in the maintenance of homeostasis in *C. glutamicum*, or can perform a function involved in the adaptation of this microorganism to different
- 30 environmental conditions, or have one or more of the activities set forth in Table 1.

Various aspects of the invention are described in further detail in the following subsections.

A. Isolated Nucleic Acid Molecules

One aspect of the invention pertains to isolated nucleic acid molecules that encode HA polypeptides or biologically active portions thereof, as well as nucleic acid fragments sufficient for use as hybridization probes or primers for the identification or amplification of HA-encoding nucleic acid (*e.g.*, HA DNA). As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (*e.g.*, cDNA or genomic DNA) and RNA molecules (*e.g.*, mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. This term also encompasses untranslated sequence located at both the 3' and 5' ends of the coding region of the gene: at least about 100 nucleotides of sequence upstream from the 5' end of the coding region and at least about 20 nucleotides of sequence downstream from the 3' end of the coding region of the gene. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA. An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated HA nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived (*e.g.*, a *C. glutamicum* cell). Moreover, an "isolated" nucleic acid molecule, such as a DNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, *e.g.*, a nucleic acid molecule having a nucleotide sequence of an odd-numbered SEQ ID NO of the Sequence Listing, or a portion thereof, can be isolated using standard molecular biology techniques and the sequence information provided herein. For example, a *C. glutamicum* HA DNA can be isolated from a *C. glutamicum* library using all or portion of one of the odd-numbered SEQ ID NO sequences of the Sequence Listing as a hybridization probe and standard hybridization techniques (*e.g.*, as described in Sambrook, J., Fritsh, E. F., and Maniatis,

T. *Molecular Cloning: A Laboratory Manual*. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

- Moreover, a nucleic acid molecule encompassing all or a portion of one of the nucleic acid sequences of the invention (e.g., an odd-numbered SEQ ID NO:) can be isolated by
- 5 the polymerase chain reaction using oligonucleotide primers designed based upon this sequence (e.g., a nucleic acid molecule encompassing all or a portion of one of the nucleic acid sequences of the invention (e.g., an odd-numbered SEQ ID NO of the Sequence Listing) can be isolated by the polymerase chain reaction using oligonucleotide primers designed based upon this same sequence). For example, mRNA
- 10 can be isolated from normal endothelial cells (e.g., by the guanidinium-thiocyanate extraction procedure of Chirgwin *et al.* (1979) *Biochemistry* 18: 5294-5299) and DNA can be prepared using reverse transcriptase (e.g., Moloney MLV reverse transcriptase, available from Gibco/BRL, Bethesda, MD; or AMV reverse transcriptase, available from Seikagaku America, Inc., St. Petersburg, FL). Synthetic oligonucleotide primers
- 15 for polymerase chain reaction amplification can be designed based upon one of the nucleotide sequences shown in the Sequence Listing. A nucleic acid of the invention can be amplified using cDNA or, alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and
- 20 characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to an HA nucleotide sequence can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

- In a preferred embodiment, an isolated nucleic acid molecule of the invention comprises one of the nucleotide sequences shown in the Sequence Listing. The nucleic
- 25 acid sequences of the invention, as set forth in the Sequence Listing, correspond to the *Corynebacterium glutamicum* HA DNAs of the invention. This DNA comprises sequences encoding HA proteins (i.e., the "coding region", indicated in each odd-numbered SEQ ID NO: sequence in the Sequence Listing), as well as 5' untranslated sequences and 3' untranslated sequences, also indicated in each odd-numbered SEQ ID
- 30 NO: in the Sequence Listing. Alternatively, the nucleic acid molecule can comprise only the coding region of any of the sequences in nucleic acid sequences of the Sequence Listing

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For the purposes of this application, it will be understood that each of the nucleic acid and amino acid sequences set forth in the Sequence Listing has an identifying RXA, RXN, RXS, or RXC number having the designation "RXA," "RXN," "RXS, or "RXC" followed by 5 digits (*i.e.*, RXA02458, RXN00249, RXS00153, or RXC00963).

- 5 Each of the nucleic acid sequences comprises up to three parts: a 5' upstream region, a coding region, and a downstream region. Each of these three regions is identified by the same RXA, RXN, RXS, or RXC designation to eliminate confusion. The recitation "one of the odd-numbered sequences in of the Sequence Listing", then, refers to any of the nucleic acid sequences in the Sequence Listing, which may also be distinguished by
- 10 their differing RXA, RXN, RXS, or RXC designations. The coding region of each of these sequences is translated into a corresponding amino acid sequence, which is also set forth in the Sequence Listing, as an even-numbered SEQ ID NO: immediately following the corresponding nucleic acid sequence. For example, the coding region for RXA02548 is set forth in SEQ ID NO:1, while the amino acid sequence which it
- 15 encodes is set forth as SEQ ID NO:2. The sequences of the nucleic acid molecules of the invention are identified by the same RXA, RXN, RXS, or RXC designations as the amino acid molecules which they encode, such that they can be readily correlated. For example, the amino acid sequences designated RXA02458, RXN00249, RXS00153, and RXC00963 are translations of the coding regions of the nucleotide sequences of nucleic
- 20 acid molecules RXA02458, RXN00249, RXS00153, and RXC00963, respectively. of the correspondence between the RXA, RXN, RXS, and RXC nucleotide and amino acid sequences of the invention and their assigned SEQ ID NOs is set forth in Table 1.

- Several of the genes of the invention are "F-designated genes". An F-designated gene includes those genes set forth in Table 1 which have an 'F' in front of the RXA,
- 25 RXN, RXS, or RXC designation. For example, SEQ ID NO:5, designated, as indicated on Table 1, as "F RXA00249", is an F-designated gene, as are SEQ ID NOs: 11, 15, and 33 (designated on Table 1 as "F RXA02264", "F RXA02274", and "F RXA00675", respectively).

- In one embodiment, the nucleic acid molecules of the present invention are not
- 30 intended to include those compiled in Table 2. In the case of the *dapD* gene, a sequence for this gene was published in Wehrmann, A., *et al.* (1998) *J. Bacteriol.* 180(12): 3159-3165. However, the sequence obtained by the inventors of the present application is

significantly longer than the published version. It is believed that the published version relied on an incorrect start codon, and thus represents only a fragment of the actual coding region.

In another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of one of the nucleotide sequences of the invention (*e.g.*, a sequence of an odd-numbered SEQ ID NO: of the Sequence Listing), or a portion thereof. A nucleic acid molecule which is complementary to one of the nucleotide sequences of the invention is one which is sufficiently complementary to one of the nucleotide sequences the Sequence Listing (*e.g.*, the sequence of an odd-numbered SEQ ID NO:) such that it can hybridize to one of the nucleotide sequences of the invention, thereby forming a stable duplex.

In still another preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleotide sequence which is at least about 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, or 60%, preferably at least about 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, or 70%, more preferably at least about 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, or 90%, or 91%, 92%, 93%, 94%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more homologous to a nucleotide sequence of the invention (*e.g.*, a sequence of an odd-numbered SEQ ID NO: of the Sequence Listing), or a portion thereof. Ranges and identity values intermediate to the above-recited ranges, (*e.g.*, 70-90% identical or 80-95% identical) are also intended to be encompassed by the present invention. For example, ranges of identity values using a combination of any of the above values recited as upper and/or lower limits are intended to be included. In an additional preferred embodiment, an isolated nucleic acid molecule of the invention comprises a nucleotide sequence which hybridizes, *e.g.*, hybridizes under stringent conditions, to one of the nucleotide sequences of the invention, or a portion thereof.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the coding region of the sequence of one of the odd-numbered SEQ ID NOs of the Sequence Listing, for example a fragment which can be used as a probe or primer or a fragment encoding a biologically active portion of an HA protein. The nucleotide sequences determined from the cloning of the HA genes from *C. glutamicum* allows for the generation of probes and primers designed for use in identifying and/or cloning HA

- homologues in other cell types and organisms, as well as HA homologues from other *Corynebacteria* or related species. The probe/primer typically comprises substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, preferably about
- 5 25, more preferably about 40, 50 or 75 consecutive nucleotides of a sense strand of one of the nucleotide sequences of the invention (*e.g.*, a sequence of one of the odd-numbered SEQ ID NOs of the Sequence Listing), an anti-sense sequence of one of these sequences, or naturally occurring mutants thereof. Primers based on a nucleotide sequence of the invention can be used in PCR reactions to clone HA homologues.
- 10 Probes based on the HA nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments, the probe further comprises a label group attached thereto, *e.g.* the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells
- 15 which misexpress an HA protein, such as by measuring a level of an HA-encoding nucleic acid in a sample of cells, *e.g.*, detecting HA mRNA levels or determining whether a genomic HA gene has been mutated or deleted.

- In one embodiment, the nucleic acid molecule of the invention encodes a protein or portion thereof which includes an amino acid sequence which is sufficiently
- 20 homologous to an amino acid sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO of the Sequence Listing) such that the protein or portion thereof maintains the ability to participate in the maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions. As used herein, the language "sufficiently homologous"
- 25 refers to proteins or portions thereof which have amino acid sequences which include a minimum number of identical or equivalent (*e.g.*, an amino acid residue which has a similar side chain as an amino acid residue in a sequence of one of the even-numbered SEQ ID NOs of the Sequence Listing) amino acid residues to an amino acid sequence of the invention such that the protein or portion thereof is able to participate in the
- 30 maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions. Proteins involved in *C. glutamicum* cell wall biosynthesis or rearrangements, metabolism of

inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or that have a *C. glutamicum* enzymatic or proteolytic activity, as described herein, may play a role in the production and secretion of one or more fine chemicals. Examples of such activities are also described herein. Thus, "the function of an HA protein"

- 5 contributes either directly or indirectly to the yield, production, and/or efficiency of production of one or more fine chemicals. Examples of HA protein activities are set forth in Table 1.

In another embodiment, the protein is at least about 50-60%, preferably at least about 60-70%, and more preferably at least about 70-80%, 80-90%, 90-95%, and most
10 preferably at least about 96%, 97%, 98%, 99% or more homologous to an entire amino acid sequence of the invention (e.g., a sequence of an even-numbered SEQ ID NO: of the Sequence Listing).

Portions of proteins encoded by the HA nucleic acid molecules of the invention are preferably biologically active portions of one of the HA proteins. As used herein,
15 the term "biologically active portion of an HA protein" is intended to include a portion, e.g., a domain/motif, of an HA protein that can participate in the maintenance of homeostasis in *C. glutamicum*, or that can perform a function involved in the adaptation of this microorganism to different environmental conditions, or has an activity as set forth in Table 1. To determine whether an HA protein or a biologically active portion
20 thereof can participate in *C. glutamicum* cell wall biosynthesis or rearrangements, metabolism of inorganic compounds, modification or degradation of aromatic or aliphatic compounds, or has a *C. glutamicum* enzymatic or proteolytic activity, an assay of enzymatic activity may be performed. Such assay methods are well known to those of ordinary skill in the art, as detailed in Example 8 of the Exemplification.

25 Additional nucleic acid fragments encoding biologically active portions of an HA protein can be prepared by isolating a portion of one of the amino acid sequences in of the invention (e.g., a sequence of an even-numbered SEQ ID NO: of the Sequence Listing), expressing the encoded portion of the HA protein or peptide (e.g., by recombinant expression *in vitro*) and assessing the activity of the encoded portion of the
30 HA protein or peptide.

The invention further encompasses nucleic acid molecules that differ from one of the nucleotide sequences of the invention (e.g., a sequence of an odd-numbered SEQ ID

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NO: of the Sequence Listing) (and portions thereof) due to degeneracy of the genetic code and thus encode the same HA protein as that encoded by the nucleotide sequences shown in of the invention. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in the Sequence Listing (e.g., an even-numbered SEQ ID NO:). In a still further embodiment, the nucleic acid molecule of the invention encodes a full length *C. glutamicum* protein which is substantially homologous to an amino acid sequence of the invention (encoded by an open reading frame shown in an odd-numbered SEQ ID NO: of the Sequence Listing).

10 It will be understood by one of ordinary skill in the art that in one embodiment the sequences of the invention are not meant to include the sequences of the prior art, such as those Genbank sequences set forth in Tables 2 or 4 which were available prior to the present invention. In one embodiment, the invention includes nucleotide and amino acid sequences having a percent identity to a nucleotide or amino acid sequence of the invention which is greater than that of a sequence of the prior art (e.g., a Genbank sequence (or the protein encoded by such a sequence) set forth in Tables 2 or 4). For example, the invention includes a nucleotide sequence which is greater than and/or at least 39% identical to the nucleotide sequence designated RXA00471 (SEQ ID NO:293), a nucleotide sequence which is greater than and/or at least 41% identical to the nucleotide sequence designated RXA00500 (SEQ ID NO:143), and a nucleotide sequence which is greater than and/or at least 35% identical to the nucleotide sequence designated RXA00502 (SEQ ID NO:147). One of ordinary skill in the art would be able to calculate the lower threshold of percent identity for any given sequence of the invention by examining the GAP-calculated percent identity scores set forth in Table 4 for each of the three top hits for the given sequence, and by subtracting the highest GAP-calculated percent identity from 100 percent. One of ordinary skill in the art will also appreciate that nucleic acid and amino acid sequences having percent identities greater than the lower threshold so calculated (e.g., at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, or 60%, preferably at least about 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, or 70%, more preferably at least about 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%,

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88%, 89%, or 90%, or 91%, 92%, 93%, 94%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more identical) are also encompassed by the invention.

In addition to the *C. glutamicum* HA nucleotide set forth in the Sequence Listing as odd-numbered SEQ ID NOs, it will be appreciated by those of ordinary skill in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of HA proteins may exist within a population (*e.g.*, the *C. glutamicum* population). Such genetic polymorphism in the HA gene may exist among individuals within a population due to natural variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding an HA protein, preferably a *C. glutamicum* HA protein. Such natural variations can typically result in 1-5% variance in the nucleotide sequence of the HA gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in HA that are the result of natural variation and that do not alter the functional activity of HA proteins are intended to be within the scope of the invention.

Nucleic acid molecules corresponding to natural variants and non-*C. glutamicum* homologues of the *C. glutamicum* HA DNA of the invention can be isolated based on their homology to the *C. glutamicum* HA nucleic acid disclosed herein using the *C. glutamicum* DNA, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15 nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising a nucleotide sequence of an odd-numbered SEQ ID NO: of the Sequence Listing. In other embodiments, the nucleic acid is at least 30, 50, 100, 250 or more nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% homologous to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 65%, more preferably at least about 70%, and even more preferably at least about 75% or more homologous to each other typically remain hybridized to each other. Such stringent conditions are known to those of ordinary skill in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are

hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50-65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to a nucleotide sequence of the invention corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein). In one embodiment, the nucleic acid encodes a natural *C. glutamicum* HA protein.

In addition to naturally-occurring variants of the HA sequence that may exist in the population, one of ordinary skill in the art will further appreciate that changes can be introduced by mutation into a nucleotide sequence of the invention, thereby leading to changes in the amino acid sequence of the encoded HA protein, without altering the functional ability of the HA protein. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in a nucleotide sequence of the invention. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of one of the HA proteins (e.g., an even-numbered SEQ ID NO: of the Sequence Listing) without altering the activity of said HA protein, whereas an "essential" amino acid residue is required for HA protein activity. Other amino acid residues, however, (e.g., those that are not conserved or only semi-conserved in the domain having HA activity) may not be essential for activity and thus are likely to be amenable to alteration without altering HA activity.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding HA proteins that contain changes in amino acid residues that are not essential for HA activity. Such HA proteins differ in amino acid sequence from a sequence of an even-numbered SEQ ID NO: of the Sequence Listing yet retain at least one of the HA activities described herein. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 50% homologous to an amino acid sequence of the invention and is capable of participating in the maintenance of homeostasis in *C. glutamicum*, or of performing a function involved in the adaptation of this microorganism to different environmental conditions, or has one or more of the activities set forth in Table 1. Preferably, the protein encoded by the nucleic acid

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molecule is at least about 50-60% homologous to the amino acid sequence of one of the odd-numbered SEQ ID NOs of the Sequence Listing, more preferably at least about 60-70% homologous to one of these sequences in, even more preferably at least about 70-80%, 80-90%, 90-95% homologous to one of these sequences, and most preferably at least about 96%, 97%, 98%, or 99% homologous to one of the amino acid sequences of the invention

To determine the percent homology of two amino acid sequences (*e.g.*, one of the amino acid sequences of the invention and a mutant form thereof) or of two nucleic acids, the sequences are aligned for optimal comparison purposes (*e.g.*, gaps can be introduced in the sequence of one protein or nucleic acid for optimal alignment with the other protein or nucleic acid). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in one sequence (*e.g.*, one of the amino acid sequences of the invention) is occupied by the same amino acid residue or nucleotide as the corresponding position in the other sequence (*e.g.*, a mutant form of the amino acid sequence), then the molecules are homologous at that position (*i.e.*, as used herein amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity"). The percent homology between the two sequences is a function of the number of identical positions shared by the sequences (*i.e.*, % homology = # of identical positions/total # of positions x 100).

An isolated nucleic acid molecule encoding an HA protein homologous to a protein sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing) can be created by introducing one or more nucleotide substitutions, additions or deletions into a nucleotide sequence of the invention such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into one of the nucleotide sequences of the invention by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic

- acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in an HA protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of an HA coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for an HA activity described herein to identify mutants that retain HA activity. Following mutagenesis of the nucleotide sequence of one of the odd-numbered SEQ ID NOs of the Sequence Listing, the encoded protein can be expressed recombinantly and the activity of the protein can be determined using, for example, assays described herein (see Example 8 of the Exemplification).
- In addition to the nucleic acid molecules encoding HA proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, *e.g.*, complementary to the coding strand of a double-stranded DNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire HA coding strand, or to only a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding an HA protein. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (*e.g.*, the entire coding region of SEQ ID NO. 3 (RXN00249) comprises nucleotides 1 to 957). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding HA. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (*i.e.*, also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding HA disclosed herein (*e.g.*, the sequences set forth as odd-numbered SEQ ID NOs in the Sequence Listing), antisense

nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of HA mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding or noncoding region of HA mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of HA mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a cell or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an HA protein to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The hybridization can be by
5 conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. The antisense molecule can be modified such that it specifically binds to a receptor or an antigen expressed on a selected cell surface, *e.g.*, by linking the antisense nucleic acid molecule to a peptide or
10 an antibody which binds to a cell surface receptor or antigen. The antisense nucleic acid molecule can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong prokaryotic, viral, or eukaryotic promoter are preferred.

15 In yet another embodiment, the antisense nucleic acid molecule of the invention is an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-
20 methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue *et al.* (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they
25 have a complementary region. Thus, ribozymes (*e.g.*, hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave HA mRNA transcripts to thereby inhibit translation of HA mRNA. A ribozyme having specificity for an HA-encoding nucleic acid can be designed based upon the nucleotide sequence of an HA DNA molecule disclosed herein (*i.e.*, SEQ ID
30 NO. 3 (RXN00249)). For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an HA-encoding mRNA. See, *e.g.*, Cech *et al.*

U.S. Patent No. 4,987,071 and Cech *et al.* U.S. Patent No. 5,116,742. Alternatively, HA mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, *e.g.*, Bartel, D. and Szostak, J.W. (1993) *Science* 261:1411-1418.

- 5 Alternatively, HA gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of an HA nucleotide sequence (*e.g.*, an HA promoter and/or enhancers) to form triple helical structures that prevent transcription of an HA gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. *et al.* (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and
- 10 Maher, L.J. (1992) *Bioassays* 14(12):807-15.

B. Recombinant Expression Vectors and Host Cells

- Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding an HA protein (or a portion thereof). As
- 15 used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of
- 20 autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to
- 25 which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors,
- 30 such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which are

5 operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (e.g., in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory

10 sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and

15 those which direct expression of the nucleotide sequence only in certain host cells. Preferred regulatory sequences are, for example, promoters such as cos-, tac-, trp-, tet-, trp-tet-, lpp-, lac-, lpp-lac-, lacI^q-, T7-, T5-, T3-, gal-, trc-, ara-, SP6-, army, SPO2, λ -P_R- or λ P_L, which are used preferably in bacteria. Additional regulatory sequences are, for example, promoters from yeasts and fungi, such as ADC1, MF α , AC, P-60, CYC1,

20 GAPDH, TEF, rp28, ADH, promoters from plants such as CaMV/35S, SSU, OCS, lib4, usp, STLS1, B33, nos or ubiquitin- or phaseolin-promoters. It is also possible to use artificial promoters. It will be appreciated by those of ordinary skill in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors

25 of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., HA proteins, mutant forms of HA proteins, fusion proteins, etc.).

The recombinant expression vectors of the invention can be designed for expression of HA proteins in prokaryotic or eukaryotic cells. For example, HA genes

30 can be expressed in bacterial cells such as *C. glutamicum*, insect cells (using baculovirus expression vectors), yeast and other fungal cells (see Romanos, M.A. *et al.* (1992) "Foreign gene expression in yeast: a review", *Yeast* 8: 423-488; van den Hondel,

- C.A.M.J.J. *et al.* (1991) "Heterologous gene expression in filamentous fungi" in: More Gene Manipulations in Fungi, J.W. Bennet & L.L. Lasure, eds., p. 396-428; Academic Press: San Diego; and van den Hondel, C.A.M.J.J. & Punt, P.J. (1991) "Gene transfer systems and vector development for filamentous fungi, in: Applied Molecular Genetics of Fungi, Peberdy, J.F. *et al.*, eds., p. 1-28, Cambridge University Press: Cambridge), algae and multicellular plant cells (see Schmidt, R. and Willmitzer, L. (1988) High efficiency *Agrobacterium tumefaciens* -mediated transformation of *Arabidopsis thaliana* leaf and cotyledon explants" *Plant Cell Rep.*: 583-586), or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein but also to the C-terminus or fused within suitable regions in the proteins. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

- Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith, D.B. and Johnson, K.S. (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein. In one embodiment, the coding sequence of the HA protein is cloned into a pGEX expression vector to create a vector encoding a fusion protein comprising, from the N-terminus to the C-terminus, GST-thrombin cleavage site-X protein. The fusion protein can be purified by affinity chromatography using glutathione-agarose resin.

Recombinant HA protein unfused to GST can be recovered by cleavage of the fusion protein with thrombin.

- Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann *et al.*, (1988) *Gene* 69:301-315) pLG338, pACYC184, pBR322, pUC18, pUC19, pKC30, pRep4, pHS1, pHS2, pPLc236, pMBL24, pLG200, pUR290, pIN-III113-B1, λ gt11, pBdCl, and pET 11d (Studier *et al.*, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89 ; and Pouwels *et al.*, eds. (1985) *Cloning Vectors*. Elsevier: New York ISBN 0 444 904018). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident λ prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter. For transformation of other varieties of bacteria, appropriate vectors may be selected. For example, the plasmids pIJ101, pIJ364, pIJ702 and pIJ361 are known to be useful in transforming *Streptomyces*, while plasmids pUB110, pC194, or pBD214 are suited for transformation of *Bacillus* species. Several plasmids of use in the transfer of genetic information into *Corynebacterium* include pHM1519, pBL1, pSA77, or pAJ667 (Pouwels *et al.*, eds. (1985) *Cloning Vectors*. Elsevier: New York ISBN 0 444 904018).

- One strategy to maximize recombinant protein expression is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in the bacterium chosen for expression, such as *C. glutamicum* (Wada *et al.* (1992) *Nucleic Acids Res.* 20: 2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

- In another embodiment, the HA protein expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari, *et al.*, (1987) *Embo J.* 6:229-234), 2 μ , pAG-1, Yep6, Yep13, pEMBL Ye23,

pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.*, (1987) *Gene* 54:113-123), and pYES2 (Invitrogen Corporation, San Diego, CA). Vectors and methods for the construction of vectors appropriate for use in other fungi, such as the filamentous fungi, include those detailed in: van den Hondel, C.A.M.J.J. & Punt, P.J. 5 (1991) "Gene transfer systems and vector development for filamentous fungi, in: Applied Molecular Genetics of Fungi, J.F. Peberdy, *et al.*, eds., p. 1-28, Cambridge University Press: Cambridge, and Pouwels *et al.*, eds. (1985) Cloning Vectors. Elsevier: New York (IBSN 0 444 904018).

Alternatively, the HA proteins of the invention can be expressed in insect cells 10 using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (*e.g.*, Sf 9 cells) include the pAc series (Smith *et al.* (1983) *Mol. Cell Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

In another embodiment, the HA proteins of the invention may be expressed in 15 unicellular plant cells (such as algae) or in plant cells from higher plants (*e.g.*, the spermatophytes, such as crop plants). Examples of plant expression vectors include those detailed in: Becker, D., Kemper, E., Schell, J. and Masterson, R. (1992) "New plant binary vectors with selectable markers located proximal to the left border", *Plant Mol. Biol.* 20: 1195-1197; and Bevan, M.W. (1984) "Binary *Agrobacterium* vectors for 20 plant transformation", *Nucl. Acid. Res.* 12: 8711-8721, and include pLGV23, pGHlac+, pBIN19, pAK2004, and pDH51 (Pouwels *et al.*, eds. (1985) Cloning Vectors. Elsevier: New York IBSN 0 444 904018).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian 25 expression vectors include pCDM8 (Seed, B. (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both 30 prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed.*, Cold Spring

Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.* (1987) *Genes Dev.* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv. Immunol.* 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) *EMBO J.* 8:729-733) and immunoglobulins (Banerji *et al.* (1983) *Cell* 33:729-740; Queen and Baltimore (1983) *Cell* 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) *PNAS* 86:5473-5477), pancreas-specific promoters (Edlund *et al.* (1985) *Science* 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally-regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) *Science* 249:374-379) and the α -fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev.* 3:537-546).

The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to HA mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene

expression using antisense genes see Weintraub, H. *et al.*, Antisense RNA as a molecular tool for genetic analysis, *Reviews - Trends in Genetics*, Vol. 1(1) (1986).

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and
5 "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used
10 herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, an HA protein can be expressed in bacterial cells such as *C. glutamicum*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those of ordinary skill in the art. Microorganisms related
15 to *Corynebacterium glutamicum* which may be conveniently used as host cells for the nucleic acid and protein molecules of the invention are set forth in Table 3.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection", "conjugation" and "transduction" are intended to
20 refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, linear DNA or RNA (*e.g.*, a linearized vector or a gene construct alone without a vector) or nucleic acid in the form of a vector (*e.g.*, a plasmid, phage, phasmid, phagemid, transposon or other DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, natural
25 competence, chemical-mediated transfer, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, *et al.* (*Molecular Cloning: A Laboratory Manual*, 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the
30 expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (*e.g.*, resistance to antibiotics) is

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generally introduced into the host cells along with the gene of interest. Preferred selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding an HA protein or can be
5 introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by, for example, drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

To create a homologous recombinant microorganism, a vector is prepared which contains at least a portion of an HA gene into which a deletion, addition or substitution
10 has been introduced to thereby alter, *e.g.*, functionally disrupt, the HA gene. Preferably, this HA gene is a *Corynebacterium glutamicum* HA gene, but it can be a homologue from a related bacterium or even from a mammalian, yeast, or insect source. In a preferred embodiment, the vector is designed such that, upon homologous recombination, the endogenous HA gene is functionally disrupted (*i.e.*, no longer
15 encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous HA gene is mutated or otherwise altered but still encodes functional protein (*e.g.*, the upstream regulatory region can be altered to thereby alter the expression of the endogenous HA protein). In the homologous recombination vector, the altered portion
20 of the HA gene is flanked at its 5' and 3' ends by additional nucleic acid of the HA gene to allow for homologous recombination to occur between the exogenous HA gene carried by the vector and an endogenous HA gene in a microorganism. The additional flanking HA nucleic acid is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA
25 (both at the 5' and 3' ends) are included in the vector (see *e.g.*, Thomas, K.R., and Capecchi, M.R. (1987) Cell 51: 503 for a description of homologous recombination vectors). The vector is introduced into a microorganism (*e.g.*, by electroporation) and cells in which the introduced HA gene has homologously recombined with the endogenous HA gene are selected, using art-known techniques.

30 In another embodiment, recombinant microorganisms can be produced which contain selected systems which allow for regulated expression of the introduced gene. For example, inclusion of an HA gene on a vector placing it under control of the lac

operon permits expression of the HA gene only in the presence of IPTG. Such regulatory systems are well known in the art.

In another embodiment, an endogenous HA gene in a host cell is disrupted (*e.g.*, by homologous recombination or other genetic means known in the art) such that
5 expression of its protein product does not occur. In another embodiment, an endogenous or introduced HA gene in a host cell has been altered by one or more point mutations, deletions, or inversions, but still encodes a functional HA protein. In still another embodiment, one or more of the regulatory regions (*e.g.*, a promoter, repressor, or inducer) of an HA gene in a microorganism has been altered (*e.g.*, by deletion,
10 truncation, inversion, or point mutation) such that the expression of the HA gene is modulated. One of ordinary skill in the art will appreciate that host cells containing more than one of the described HA gene and protein modifications may be readily produced using the methods of the invention, and are meant to be included in the present invention.

15 A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) an HA protein. Accordingly, the invention further provides methods for producing HA proteins using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding an HA protein has been
20 introduced, or into which genome has been introduced a gene encoding a wild-type or altered HA protein) in a suitable medium until HA protein is produced. In another embodiment, the method further comprises isolating HA proteins from the medium or the host cell.

25 C. Isolated HA Proteins

Another aspect of the invention pertains to isolated HA proteins, and biologically active portions thereof. An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material when produced by recombinant DNA techniques, or chemical precursors or other chemicals when chemically synthesized.
30 The language "substantially free of cellular material" includes preparations of HA protein in which the protein is separated from cellular components of the cells in which it is naturally or recombinantly produced. In one embodiment, the language

- "substantially free of cellular material" includes preparations of HA protein having less than about 30% (by dry weight) of non-HA protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-HA protein, still more preferably less than about 10% of non-HA protein, and most preferably less than about 5% non-HA protein. When the HA protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation. The language "substantially free of chemical precursors or other chemicals" includes
- 5 preparations of HA protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of HA protein having less than about 30% (by dry weight) of chemical
- 10 precursors or non-HA chemicals, more preferably less than about 20% chemical
- 15 precursors or non-HA chemicals, still more preferably less than about 10% chemical precursors or non-HA chemicals, and most preferably less than about 5% chemical precursors or non-HA chemicals. In preferred embodiments, isolated proteins or biologically active portions thereof lack contaminating proteins from the same organism from which the HA protein is derived. Typically, such proteins are produced by
- 20 recombinant expression of, for example, a *C. glutamicum* HA protein in a microorganism such as *C. glutamicum*.

An isolated HA protein or a portion thereof of the invention can participate in the repair or recombination of DNA, in the transposition of genetic material, in gene expression (*i.e.*, the processes of transcription or translation), in protein folding, or in

25 protein secretion in *Corynebacterium glutamicum*, or has one or more of the activities set forth in Table 1. In preferred embodiments, the protein or portion thereof comprises an amino acid sequence which is sufficiently homologous to an amino acid sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing) such that the protein or portion thereof maintains the ability to participate in the

30 maintenance of homeostasis in *C. glutamicum*, or to perform a function involved in the adaptation of this microorganism to different environmental conditions. The portion of the protein is preferably a biologically active portion as described herein. In another

preferred embodiment, an HA protein of the invention has an amino acid sequence set forth as an even-numbered SEQ ID NO: of the Sequence Listing. In yet another preferred embodiment, the HA protein has an amino acid sequence which is encoded by a nucleotide sequence which hybridizes, *e.g.*, hybridizes under stringent conditions, to a nucleotide sequence the invention (*e.g.*, a sequence of an odd-numbered SEQ ID NO: of the Sequence Listing). In still another preferred embodiment, the HA protein has an amino acid sequence which is encoded by a nucleotide sequence that is at least about 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, or 60%, preferably at least about 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, or 70%, more preferably at least about 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, or 90%, or 91%, 92%, 93%, 94%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more homologous to one of the nucleic acid sequences of the invention, or a portion thereof. Ranges and identity values intermediate to the above-recited values, (*e.g.*, 70-90% identical or 80-95% identical) are also intended to be encompassed by the present invention. For example, ranges of identity values using a combination of any of the above values recited as upper and/or lower limits are intended to be included. The preferred HA proteins of the present invention also preferably possess at least one of the HA activities described herein. For example, a preferred HA protein of the present invention includes an amino acid sequence encoded by a nucleotide sequence which hybridizes, *e.g.*, hybridizes under stringent conditions, to a nucleotide sequence of the invention, and which can participate in the maintenance of homeostasis in *C. glutamicum*, or can perform a function involved in the adaptation of this microorganism to different environmental conditions, or which has one or more of the activities set forth in Table 1.

In other embodiments, the HA protein is substantially homologous to an amino acid sequence of the invention (*e.g.*, a sequence of an even-numbered SEQ ID NO: of the Sequence Listing) and retains the functional activity of the protein of one of the amino acid sequences of the invention yet differs in amino acid sequence due to natural variation or mutagenesis, as described in detail in subsection I above. Accordingly, in another embodiment, the HA protein is a protein which comprises an amino acid sequence which is at least about 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, or 60%, preferably at least about 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%,

69%, or 70%, more preferably at least about 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, or 90%, or 91%, 92%, 93%, 94%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more homologous to an entire amino acid sequence of the invention and which

5 has at least one of the HA activities described herein. Ranges and identity values intermediate to the above-recited values, (e.g., 70-90% identical or 80-95% identical) are also intended to be encompassed by the present invention. For example, ranges of identity values using a combination of any of the above values recited as upper and/or lower limits are intended to be included. In another embodiment, the invention pertains

10 to a full length *C. glutamicum* protein which is substantially homologous to an entire amino acid sequence of the invention.

Biologically active portions of an HA protein include peptides comprising amino acid sequences derived from the amino acid sequence of an HA protein, e.g., the amino acid sequence of an even-numbered SEQ ID NO: of the Sequence Listing, the amino

15 acid sequence of a protein homologous to an HA protein, which include fewer amino acids than a full length HA protein or the full length protein which is homologous to an HA protein, and exhibit at least one activity of an HA protein. Typically, biologically active portions (peptides, e.g., peptides which are, for example, 5, 10, 15, 20, 30, 35, 36, 37, 38, 39, 40, 50, 100 or more amino acids in length) comprise a domain or motif with

20 at least one activity of an HA protein. Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the activities described herein. Preferably, the biologically active portions of an HA protein include one or more selected domains/motifs or portions thereof having biological activity.

25 HA proteins are preferably produced by recombinant DNA techniques. For example, a nucleic acid molecule encoding the protein is cloned into an expression vector (as described above), the expression vector is introduced into a host cell (as described above) and the HA protein is expressed in the host cell. The HA protein can then be isolated from the cells by an appropriate purification scheme using standard

30 protein purification techniques. Alternative to recombinant expression, an HA protein, polypeptide, or peptide can be synthesized chemically using standard peptide synthesis techniques. Moreover, native HA protein can be isolated from cells (e.g., endothelial

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cells), for example using an anti-HA antibody, which can be produced by standard techniques utilizing an HA protein or fragment thereof of this invention.

The invention also provides HA chimeric or fusion proteins. As used herein, an HA "chimeric protein" or "fusion protein" comprises an HA polypeptide operatively
5 linked to a non-HA polypeptide. An "HA polypeptide" refers to a polypeptide having an amino acid sequence corresponding to an HA protein, whereas a "non-HA polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the HA protein, e.g., a protein which is different from the HA protein and which is derived from the same or a different organism. Within the
10 fusion protein, the term "operatively linked" is intended to indicate that the HA polypeptide and the non-HA polypeptide are fused in-frame to each other. The non-HA polypeptide can be fused to the N-terminus or C-terminus of the HA polypeptide. For example, in one embodiment the fusion protein is a GST-HA fusion protein in which the HA sequences are fused to the C-terminus of the GST sequences. Such fusion proteins
15 can facilitate the purification of recombinant HA proteins. In another embodiment, the fusion protein is an HA protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of an HA protein can be increased through use of a heterologous signal sequence.

Preferably, an HA chimeric or fusion protein of the invention is produced by
20 standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid
25 undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric
30 gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (e.g., a GST polypeptide). An HA-

encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the HA protein.

Homologues of the HA protein can be generated by mutagenesis, *e.g.*, discrete point mutation or truncation of the HA protein. As used herein, the term "homologue" refers to a variant form of the HA protein which acts as an agonist or antagonist of the activity of the HA protein. An agonist of the HA protein can retain substantially the same, or a subset, of the biological activities of the HA protein. An antagonist of the HA protein can inhibit one or more of the activities of the naturally occurring form of the HA protein, by, for example, competitively binding to a downstream or upstream member of a biochemical cascade which includes the HA protein, by binding to a target molecule with which the HA protein interacts, such that no functional interaction is possible, or by binding directly to the HA protein and inhibiting its normal activity.

In an alternative embodiment, homologues of the HA protein can be identified by screening combinatorial libraries of mutants, *e.g.*, truncation mutants, of the HA protein for HA protein agonist or antagonist activity. In one embodiment, a variegated library of HA variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of HA variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential HA sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (*e.g.*, for phage display) containing the set of HA sequences therein. There are a variety of methods which can be used to produce libraries of potential HA homologues from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential HA sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, *e.g.*, Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura *et al.* (1984) *Annu. Rev. Biochem.* 53:323; Itakura *et al.* (1984) *Science* 198:1056; Ike *et al.* (1983) *Nucleic Acid Res.* 11:477).

In addition, libraries of fragments of the HA protein coding can be used to generate a variegated population of HA fragments for screening and subsequent

selection of homologues of an HA protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of an HA coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to
5 form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the HA protein.

10 Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of HA homologues. The most widely used techniques, which are amenable to high through-put
15 analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recursive ensemble mutagenesis (REM), a new technique which enhances the
20 frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify HA homologues (Arkin and Yourvan (1992) *PNAS* 89:7811-7815; Delgrave *et al.* (1993) *Protein Engineering* 6(3):327-331).

In another embodiment, cell based assays can be exploited to analyze a variegated HA library, using methods well known in the art.

25

D. Uses and Methods of the Invention

The nucleic acid molecules, proteins, protein homologues, fusion proteins, primers, vectors, and host cells described herein can be used in one or more of the following methods: identification of *C. glutamicum* and related organisms; mapping of
30 genomes of organisms related to *C. glutamicum*; identification and localization of *C. glutamicum* sequences of interest; evolutionary studies; determination of HA protein regions required for function; modulation of an HA protein activity; modulation of the

metabolism of one or more inorganic compounds; modulation of the modification or degradation of one or more aromatic or aliphatic compounds; modulation of cell wall synthesis or rearrangements; modulation of enzyme activity or proteolysis; and modulation of cellular production of a desired compound, such as a fine chemical.

- 5 The HA nucleic acid molecules of the invention have a variety of uses. First, they may be used to identify an organism as being *Corynebacterium glutamicum* or a close relative thereof. Also, they may be used to identify the presence of *C. glutamicum* or a relative thereof in a mixed population of microorganisms. The invention provides the nucleic acid sequences of a number of *C. glutamicum* genes; by probing the
- 10 extracted genomic DNA of a culture of a unique or mixed population of microorganisms under stringent conditions with a probe spanning a region of a *C. glutamicum* gene which is unique to this organism, one can ascertain whether this organism is present. Although *Corynebacterium glutamicum* itself is nonpathogenic, it is related to pathogenic species, such as *Corynebacterium diphtheriae*. *Corynebacterium diphtheriae*
- 15 is the causative agent of diphtheria, a rapidly developing, acute, febrile infection which involves both local and systemic pathology. In this disease, a local lesion develops in the upper respiratory tract and involves necrotic injury to epithelial cells; the bacilli secrete toxin which is disseminated through this lesion to distal susceptible tissues of the body. Degenerative changes brought about by the inhibition of protein synthesis in
- 20 these tissues, which include heart, muscle, peripheral nerves, adrenals, kidneys, liver and spleen, result in the systemic pathology of the disease. Diphtheria continues to have high incidence in many parts of the world, including Africa, Asia, Eastern Europe and the independent states of the former Soviet Union. An ongoing epidemic of diphtheria in the latter two regions has resulted in at least 5,000 deaths since 1990.
- 25 In one embodiment, the invention provides a method of identifying the presence or activity of *Corynebacterium diphtheriae* in a subject. This method includes detection of one or more of the nucleic acid or amino acid sequences of the invention (e.g., the sequences set forth in as odd-numbered or even-numbered SEQ ID NOs, respectively, in the Sequence Listing) in a subject, thereby detecting the presence or activity of
- 30 *Corynebacterium diphtheriae* in the subject. *C. glutamicum* and *C. diphtheriae* are related bacteria, and many of the nucleic acid and protein molecules in *C. glutamicum*

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are homologous to *C. diphtheriae* nucleic acid and protein molecules, and can therefore be used to detect *C. diphtheriae* in a subject.

The nucleic acid and protein molecules of the invention may also serve as markers for specific regions of the genome. This has utility not only in the mapping of the genome, but also for functional studies of *C. glutamicum* proteins. For example, to identify the region of the genome to which a particular *C. glutamicum* DNA-binding protein binds, the *C. glutamicum* genome could be digested, and the fragments incubated with the DNA-binding protein. Those which bind the protein may be additionally probed with the nucleic acid molecules of the invention, preferably with readily detectable labels; binding of such a nucleic acid molecule to the genome fragment enables the localization of the fragment to the genome map of *C. glutamicum*, and, when performed multiple times with different enzymes, facilitates a rapid determination of the nucleic acid sequence to which the protein binds. Further, the nucleic acid molecules of the invention may be sufficiently homologous to the sequences of related species such that these nucleic acid molecules may serve as markers for the construction of a genomic map in related bacteria, such as *Brevibacterium lactofermentum*.

The HA nucleic acid molecules of the invention are also useful for evolutionary and protein structural studies. The processes involved in adaptation and the maintenance of homeostasis in which the molecules of the invention participate are utilized by a wide variety of species; by comparing the sequences of the nucleic acid molecules of the present invention to those encoding similar enzymes from other organisms, the evolutionary relatedness of the organisms can be assessed. Similarly, such a comparison permits an assessment of which regions of the sequence are conserved and which are not, which may aid in determining those regions of the protein which are essential for the functioning of the enzyme. This type of determination is of value for protein engineering studies and may give an indication of what the protein can tolerate in terms of mutagenesis without losing function.

Manipulation of the HA nucleic acid molecules of the invention may result in the production of HA proteins having functional differences from the wild-type HA proteins. These proteins may be improved in efficiency or activity, may be present in greater numbers in the cell than is usual, or may be decreased in efficiency or activity.

The invention provides methods for screening molecules which modulate the activity of an HA protein, either by interacting with the protein itself or a substrate or binding partner of the HA protein, or by modulating the transcription or translation of an HA nucleic acid molecule of the invention. In such methods, a microorganism
5 expressing one or more HA proteins of the invention is contacted with one or more test compounds, and the effect of each test compound on the activity or level of expression of the HA protein is assessed.

The modulation of activity or number of HA proteins involved in cell wall biosynthesis or rearrangements may impact the production, yield, and/or efficiency of
10 production of one or more fine chemicals from *C. glutamicum* cells. For example, by altering the activity of these proteins, it may be possible to modulate the structure or thickness of the cell wall. The cell wall serves in large measure as a protective device against osmotic lysis and external sources of injury; by modifying the cell wall it may be possible to increase the ability of *C. glutamicum* to withstand the mechanical and shear
15 force stresses encountered by this microorganism during large-scale fermentor culture. Further, each *C. glutamicum* cell is surrounded by a thick cell wall, and thus, a significant portion of the biomass present in large scale culture consists of cell wall. By increasing the rate at which the cell wall is synthesized or by activating cell wall synthesis (through genetic engineering of the HA cell wall proteins of the invention) it
20 may be possible to improve the growth rate of the microorganism. Similarly, by decreasing the activity or number of proteins involved in the degradation of cell wall or by decreasing the repression of cell wall biosynthesis, an overall increase in cell wall production may be achieved. An increase in the number of viable *C. glutamicum* cells (as may be accomplished by any of the foregoing described protein alterations) should
25 result in increased numbers of cells producing the desired fine chemical in large-scale fermentor culture, which should permit increased yields or efficiency of production of these compounds from the culture.

The modulation of activity or number of *C. glutamicum* HA proteins that participate in the modification or degradation of aromatic or aliphatic compounds may
30 also have direct or indirect impacts on the production of one or more fine chemicals from these cells. Certain aromatic or aliphatic modification or degradation products are desirable fine chemicals (e.g., organic acids or modified aromatic and aliphatic

compounds); thus, by modifying the enzymes which perform these modifications (*e.g.*, hydroxylation, methylation, or isomerization) or degradation reactions, it may be possible to increase the yields of these desired compounds. Similarly, by decreasing the activity or number of proteins involved in pathways which further degrade the modified or breakdown products of the aforementioned reactions it may be possible to improve the yields of these fine chemicals from *C. glutamicum* cells in culture.

These aromatic and aliphatic modification and degradative enzymes are themselves fine chemicals. In purified form, these enzymes may be used to degrade aromatic and aliphatic compounds (*e.g.*, toxic chemicals such as petroleum products), either for the bioremediation of polluted sites, for the engineered decomposition of wastes, or for the large-scale and economically feasible production of desired modified aromatic or aliphatic compounds or their breakdown products, some of which may be conveniently used as carbon or energy sources for other fine chemical-producing compounds in culture (see, *e.g.*, Faber, K. (1995) *Biotransformations in Organic Chemistry*, Springer: Berlin and references therein; and Roberts, S.M., ed. (1992-1996) *Preparative Biotransformations*, Wiley: Chichester, and references therein). By genetically altering these proteins such that their regulation by other cellular mechanisms is lessened or abolished, it may be possible to increase the overall number or activity of these proteins, thereby improving not only the yield of these fine chemicals but also the activity of these harvested proteins.

The modification of these aromatic and aliphatic modifying and degradation enzymes may also have an indirect effect on the production of one or more fine chemical. Many aromatic and aliphatic compounds (such as those that may be encountered as impurities in culture media or as waste products from cellular metabolism) are toxic to cells; by modifying and/or degrading these compounds such that they may be readily removed or destroyed, cellular viability should be increased. Further, these enzymes may modify or degrade these compounds in such a manner that the resulting products may enter the normal carbon metabolism pathways of the cell, thus rendering the cell able to use these compounds as alternate carbon or energy sources. In large-scale culture situations, when there may be limiting amounts of optimal carbon sources, these enzymes provide a method by which cells may continue to grow and divide using aromatic or aliphatic compounds as nutrients. In either case, the

resulting increase in the number of *C. glutamicum* cells in the culture producing the desired fine chemical should in turn result in increased yields or efficiency of production of the fine chemical(s).

Modifications in activity or number of HA proteins involved in the metabolism of inorganic compounds may also directly or indirectly affect the production of one or more fine chemicals from *C. glutamicum* or related bacterial cultures. For example, many desirable fine chemicals, such as nucleic acids, amino acids, cofactors and vitamins (e.g., thiamine, biotin, and lipoic acid) cannot be synthesized without inorganic molecules such as phosphorous, nitrate, sulfate, and iron. The inorganic metabolism proteins of the invention permit the cell to obtain these molecules from a variety of inorganic compounds and to divert them into various fine chemical biosynthetic pathways. Therefore, by increasing the activity or number of enzymes involved in the metabolism of these inorganic compounds, it may be possible to increase the supply of these possibly limiting inorganic molecules, thereby directly increasing the production or efficiency of production of various fine chemicals from *C. glutamicum* cells containing such altered proteins. Modification of the activity or number of inorganic metabolism enzymes of the invention may also render *C. glutamicum* able to better utilize limited inorganic compound supplies, or to utilize nonoptimal inorganic compounds to synthesize amino acids, vitamins, cofactors, or nucleic acids, all of which are necessary for continued growth and replication of the cell. By improving the viability of these cells in large-scale culture, the number of *C. glutamicum* cells producing one or more fine chemicals in the culture may also be increased, in turn increasing the yields or efficiency of production of one or more fine chemicals.

C. glutamicum enzymes for general processes are themselves desirable fine chemicals. The specific properties of enzymes (i.e., regio- and stereospecificity, among others) make them useful catalysts for chemical reactions *in vitro*. Either whole *C. glutamicum* cells may be incubated with an appropriate substrate such that the desired product is produced by enzymes in the cell, or the desired enzymes may be overproduced and purified from *C. glutamicum* cultures (or those of a related bacterium) and subsequently utilized in *in vitro* reactions in an industrial setting (either in solution or immobilized on a suitable immobile phase). In either situation, the enzyme can either be a natural *C. glutamicum* protein, or it may be mutagenized to have an altered activity;

- typical industrial uses for such enzymes include as catalysts in the chemical industry (e.g., for synthetic organic chemistry) as food additives, as feed components, for fruit processing, for leather preparation, in detergents, in analysis and medicine, and in the textile industry (see, e.g., Yamada, H. (1993) "Microbial reactions for the production of useful organic compounds," *Chimica* 47: 5-10; Roberts, S.M. (1998) Preparative biotransformations: the employment of enzymes and whole-cells in synthetic chemistry," *J. Chem. Soc. Perkin Trans. 1*: 157-169; Zaks, A. and Dodds, D.R. (1997) "Application of biocatalysis and biotransformations to the synthesis of pharmaceuticals," *DDT* 2: 513-531; Roberts, S.M. and Williamson, N.M. (1997) "The use of enzymes for the preparation of biologically active natural products and analogues in optically active form," *Curr. Organ. Chemistry* 1: 1-20; Faber, K. (1995) Biotransformations in Organic Chemistry, Springer: Berlin; Roberts, S.M., ed. (1992-96) Preparative Biotransformations, Wiley: Chichester; Cheetham, P.S.J. (1995) "The applications of enzymes in industry" in : Handbook of Enzyme Biotechnology, 3rd ed., Wiseman, A., ed., Ellis Horwood, p. 419-552; and Ullmann's Encyclopedia of Industrial Chemistry (1987), vol. A9, Enzymes, p. 390-457). Thus, by increasing the activity or number of these enzymes, it may be possible to also increase the ability of the cell to convert supplied substrates to desired products, or to overproduce these enzymes for increased yields in large-scale culture. Further, by mutagenizing these proteins it may be possible to remove feedback inhibition or other repressive cellular regulatory controls such that greater numbers of these enzymes may be produced and activated by the cell, thereby leading to greater yields, production, or efficiency of production of these fine chemical proteins from large-scale cultures. Further, manipulation of these enzymes may alter the activity of one or more *C. glutamicum* metabolic pathways, such as those for the biosynthesis or secretion of one or more fine chemicals.

Mutagenesis of the proteolytic enzymes of the invention such that they are altered in activity or number may also directly or indirectly affect the yield, production, and/or efficiency of production of one or more fine chemicals from *C. glutamicum*. For example, by increasing the activity or number of these proteins, it may be possible to increase the ability of the bacterium to survive in large-scale culture, due to an increased ability of the cell to rapidly degrade proteins misfolded in response to the high temperatures, nonoptimal pH, and other stresses encountered during fermentor culture.

Increased numbers of cells in these cultures may result in increased yields or efficiency of production of one or more desired fine chemicals, due to the relatively larger number of cells producing these compounds in the culture. Also, *C. glutamicum* cells possess multiple cell-surface proteases which serve to break down external nutrients into
5 molecules which may be more readily incorporated by the cells as carbon/energy sources or nutrients of other kinds. An increase in activity or number of these enzymes may improve this turnover and increase the levels of available nutrients, thereby improving cell growth or production. Thus, modifications of the proteases of the invention may indirectly impact *C. glutamicum* fine chemical production.

10 A more direct impact on fine chemical production in response to the modification of one or more of the proteases of the invention may occur when these proteases are involved in the production or degradation of a desired fine chemical. By decreasing the activity of a protease which degrades a fine chemical or a protein involved in the synthesis of a fine chemical it may be possible to increase the levels of
15 that fine chemical (due to the decreased degradation or increased synthesis of the compound). Similarly, by increasing the activity of a protease which degrades a compound to result in a fine chemical or a protein involved in the degradation of a fine chemical, a similar result should be achieved: increased levels of the desired fine chemical from *C. glutamicum* cells containing these engineered proteins.

20 The aforementioned mutagenesis strategies for HA proteins to result in increased yields of a fine chemical from *C. glutamicum* are not meant to be limiting; variations on these strategies will be readily apparent to one of ordinary skill in the art. Using such strategies, and incorporating the mechanisms disclosed herein, the nucleic acid and protein molecules of the invention may be utilized to generate *C. glutamicum* or related
25 strains of bacteria expressing mutated HA nucleic acid and protein molecules such that the yield, production, and/or efficiency of production of a desired compound is improved. This desired compound may be any product produced by *C. glutamicum*, which includes the final products of biosynthesis pathways and intermediates of naturally-occurring metabolic pathways, as well as molecules which do not naturally
30 occur in the metabolism of *C. glutamicum*, but which are produced by a *C. glutamicum* strain of the invention.

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This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patent applications, patents, published patent applications, Tables, and the sequence listing cited throughout this application are hereby incorporated by reference.

Table 1: Genes in the Application

Nucleic Acid		Amino Acid		Identification Code	Contig.	NT Start		NT Stop		Function
SEQ ID NO		SEQ ID NO								
1		2								
3		4		RXA02548	GR00727	3	293			SULFATE ADENYLATE TRANSFERASE SUBUNIT 2 (EC 2.7.7.4)
5		6		RXN00249	VV0057	36825	35869			ADENYLYLSULFATE KINASE (EC 2.7.1.25)
7		8		F RXA02049	GR00037	8837	7884			ADENYLYLSULFATE KINASE (EC 2.7.1.25)
				RXA01073	GR00300	1274	2104			NH(3)-DEPENDENT NAD(+) SYNTHETASE (EC 6.3.5.1)
Urease										
Nucleic Acid		Amino Acid		Identification Code	Contig.	NT Start		NT Stop		Function
SEQ ID NO		SEQ ID NO								
9		10								
11		12		RXN02913	VV0020	8998	8513			UREASE BETA SUBUNIT (EC 3.5.1.5)
13		14		F RXA02264	GR00655	123	4			UREASE ALPHA SUBUNIT (EC 3.5.1.5)
15		16		RXN00274	VV0020	8609	6800			UREASE ALPHA SUBUNIT (EC 3.5.1.5)
17		18		F RXA02274	GR00656	3	1804			UREASE ALPHA SUBUNIT (EC 3.5.1.5)
19		20		RXA02265	GR00655	452	153			UREASE GAMMA SUBUNIT (EC 3.5.1.5)
21		22		RXA02278	GR00656	3420	4268			UREASE OPERON URED PROTEIN
23		24		RXA02275	GR00656	1632	2102			UREASE ACCESSORY PROTEIN UREE
25		26		RXA02276	GR00656	2105	2782			UREASE ACCESSORY PROTEIN UREF
27		28		RXA02277	GR00656	2802	3416			UREASE ACCESSORY PROTEIN UREG
29		30		RXA02603	GR00742	7742	8737			4-HYDROXYBENZOATE OCTAPRENYLTRANSFERASE (EC 2.5.1.-)
				RXA01385	GR00406	5320	3440			PHENOL 2 MONOOXYGENASE (EC 1.14.13.7)
Proteolysis										
Nucleic Acid		Amino Acid		Identification Code	Contig.	NT Start		NT Stop		Function
SEQ ID NO		SEQ ID NO								
31		32								
33		34		RXN00675	VV0005	33258	34049			METHIONINE AMINOPEPTIDASE (EC 3.4.11.18)
35		36		F RXA00675	GR00178	2	484			METHIONINE AMINOPEPTIDASE (EC 3.4.11.18)
37		38		RXA01609	GR00449	2740	3612			METHIONINE AMINOPEPTIDASE (EC 3.4.11.18)
39		40		RXA01358	GR00393	5337	6957			ATP-DEPENDENT PROTEASE LA (EC 3.4.21.53)
41		42		RXA01458	GR00420	3225	2176			ATP-DEPENDENT PROTEASE LA (EC 3.4.21.53)
43		44		RXA01654	GR00459	986	1981			(AL022121) putative alkaline serine protease [Mycobacterium tuberculosis]
45		46		RXN01868	VV0127	9980	11905			ZINC METALLOPROTEASE (EC 3.4.24.-)
47		48		F RXA01868	GR00534	1640	30			ZINC METALLOPROTEASE (EC 3.4.24.-)
49		50		RXA03028	GR00534	1954	1652			ZINC METALLOPROTEASE (EC 3.4.24.-)
51		52		F RXA02470	VV0008	41156	43930			ATP-DEPENDENT CLP PROTEASE ATP-BINDING SUBUNIT CLPA
					GR00715	2216	3196			ATP-DEPENDENT CLP PROTEASE ATP-BINDING SUBUNIT CLPA

Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
53	54	F RXA02471	GR00715	3159	4991	ATP-DEPENDENT CLP PROTEASE ATP-BINDING SUBUNIT CLPA
55	56	RXA02630	GR00748	2654	1332	(AL021999) putative serine protease [Mycobacterium tuberculosis]
57	58	RXA02834	GR000823	3	497	ATPases with chaperone activity, ATP-dependent protease subunit
59	60	RXA00112	GR00016	3687	2497	PROBABLE PERIPLASMIC SERINE PROTEASE DO-LIKE PRECURSOR
61	62	RXA00566	GR00152	742	137	ATP-DEPENDENT CLP PROTEASE PROTEOLYTIC SUBUNIT (EC 3.4.21.92)
63	64	RXA00567	GR00152	1388	798	ATP-DEPENDENT CLP PROTEASE PROTEOLYTIC SUBUNIT (EC 3.4.21.92)
65	66	RXN03094	WV0057	1794	43	CLPB PROTEIN
67	68	F RXA01668	GR00464	2205	3920	CLPB PROTEIN
69	70	RXN01120	WV0182	5678	4401	ATP-DEPENDENT CLP PROTEASE ATP-BINDING SUBUNIT CLPXP
71	72	F RXA01120	GR00310	2349	1072	ATP-DEPENDENT CLP PROTEASE ATP-BINDING SUBUNIT CLPXP
73	74	RXA00744	GR00202	10722	9781	Periplasmic serine proteases
75	76	RXA00844	GR00228	8620	4453	Hypothetical Secretory Serine Protease (EC 3.4.21.-)
77	78	RXA01151	GR00324	862	5	ATP-dependent Zn proteases
79	80	RXA02317	GR00665	9664	9053	PEPTIDASE E (EC 3.4.-.-)
81	82	RXA02844	GR00751	767	117	XAA-PRO DIPEPTIDASE (EC 3.4.13.9)
83	84	RXN02820	WV0131	4799	6109	GAMMA-GLUTAMYL TRANSPEPTIDASE (EC 2.3.2.2)
85	86	F RXA02820	GR00801	1	507	GAMMA-GLUTAMYL TRANSPEPTIDASE (EC 2.3.2.2)
87	88	F RXA02000	GR00589	3430	3933	GAMMA-GLUTAMYL TRANSPEPTIDASE (EC 2.3.2.2)
89	90	RXN03178	WV0334	921	121	PENICILLIN-BINDING PROTEIN 5' PRECURSOR (D-ALANYL-D-ALANINE CARBOXYPEPTIDASE) (EC 3.4.16.4)
91	92	F RXA02859	GR10005	846	121	PENICILLIN-BINDING PROTEIN 5' PRECURSOR (D-ALANYL-D-ALANINE CARBOXYPEPTIDASE) (EC 3.4.16.4)
93	94	RXA00137	GR00022	738	1826	XAA-PRO AMINOPEPTIDASE (EC 3.4.11.9)
95	96	RXN00499	WV0096	8158	9438	PROLINE IMINOPEPTIDASE (EC 3.4.11.5)
97	98	F RXA00499	GR00125	3	959	PROLINE IMINOPEPTIDASE
99	100	RXN00877	WV0099	2221	3885	PEPTIDYL-DIPEPTIDASE DCP (EC 3.4.15.5)
101	102	F RXA00877	GR00242	3	1067	PEPTIDYL-DIPEPTIDASE DCP (EC 3.4.15.5)
103	104	RXN01014	WV0209	13328	10728	AMINOPEPTIDASE N (EC 3.4.11.2)
105	106	F RXA01014	GR00289	3	1580	AMINOPEPTIDASE N (EC 3.4.11.2)
107	108	F RXA01018	GR00290	2289	3152	AMINOPEPTIDASE N (EC 3.4.11.2)
109	110	RXA001147	GR00323	1353	94	XAA-PRO AMINOPEPTIDASE I PRECURSOR (EC 3.4.11.1)
111	112	RXA001161	GR00329	1253	117	AMINOPEPTIDASE A/ (EC 3.4.11.1)
113	114	RXA01181	WV0065	1	957	AMINOPEPTIDASE
115	116	F RXA01181	GR00337	1	957	AMINOPEPTIDASE
117	118	RXN01277	WV0009	32155	34158	PROLYL ENDOPEPTIDASE (EC 3.4.21.26)
119	120	F RXA01277	GR00368	1738	50	PROLYL ENDOPEPTIDASE (EC 3.4.21.26)
121	122	RXA01914	GR00348	125	550	AMINOPEPTIDASE
123	124	RXA02048	GR00624	207	1580	AMINOPEPTIDASE N (EC 3.4.11.2)
125	126	RXN00621	WV0135	5853	5071	PROTEASE II (EC 3.4.21.83)
127	128	F RXA00621	GR00163	4075	4857	PTRB periplasmic protease
129	130	RXN00622	WV0135	5150	3735	PROTEASE II (EC 3.4.21.83)
131	132	F RXA00622	GR00163	4778	6193	PTRB periplasmic protease
133	134	RXN00982	WV0149	7596	6091	(L42756) proteinase [Streptomyces lividans]
135	136	F RXA00977	GR00275	1647	2660	(L42756) proteinase [Streptomyces lividans]
137	138	F RXA00982	GR00276	5194	4949	(L42756) proteinase [Streptomyces lividans]
139	140	RXA00152	GR00023	7175	5880	HFLC PROTEIN (EC 3.4.-.-)
141	142	RXA02558	GR00731	4938	3965	HFLC PROTEIN (EC 3.4.-.-)

Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
143	144	FXA00500	GR00125	969	1643	O-SIALOGLYCOPROTEIN ENDOPEPTIDASE (EC 3.4.24.57)
145	146	FXA00501	GR00125	1643	2149	O-SIALOGLYCOPROTEIN ENDOPEPTIDASE (EC 3.4.24.57)
147	148	FXA00502	GR00125	2156	3187	O-SIALOGLYCOPROTEIN ENDOPEPTIDASE (EC 3.4.24.57)
Enzymes in general						
Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
149	150	RXN002599	WV0098	16346	17110	Hypothetical Methyltransferase (EC 2.1.1.-)
151	152	F RXA02589	GR00741	13904	13040	Predicted S-adenosylmethionine-dependent methyltransferase
153	154	RXN00228	GR00032	26836	26072	SAM-dependent methyltransferases
155	156	RXN01885	WV0184	2004	2804	Hypothetical Methyltransferase (EC 2.1.1.-)
157	158	F RXA01886	GR00539	1589	2389	SAM-dependent methyltransferases
159	160	RXN02592	GR00741	18477	17707	SAM-dependent methyltransferases
161	162	RXN01795	WV0093	722	1318	MODIFICATION METHYLASE (EC 2.1.1.73)
163	164	F RXA01796	GR00507	706	1140	MODIFICATION METHYLASE (EC 2.1.1.73)
165	166	RXN01214	GR00351	1640	3130	LACCASE 1 PRECURSOR (EC 1.10.3.2)
167	168	RXN01250	GR00364	592	5	LACCASE 1 PRECURSOR (EC 1.10.3.2)
169	170	RXA02477	GR00715	10581	11201	CARBONIC ANHYDRASE (EC 4.2.1.1)
171	172	RXN00833	GR00225	374	6	THIOL PEROXIDASE (EC 1.11.1.-)
173	174	F RXA00833	GR00225	374	6	THIOL PEROXIDASE (EC 1.11.1.-)
175	176	RXA01224	GR00354	4186	5208	2-NITROPROPANE DIOXYGENASE (EC 1.13.11.32)
177	178	RXA01182	GR00337	1363	971	Hypothetical Oxidoreductase
179	180	RXN02531	GR00726	1226	1936	Hypothetical Oxidoreductase
181	182	RXN00989	WV0005	22416	20926	BETAINE-ALDEHYDE DEHYDROGENASE PRECURSOR (EC 1.2.1.8)
183	184	F RXA00689	GR00180	14001	775	BETAINE-ALDEHYDE DEHYDROGENASE PRECURSOR (EC 1.2.1.8)
185	186	RXN03128	WV0120	3	857	MORPHINE 6-DEHYDROGENASE (EC 1.1.1.218)
187	188	F RXA02192	GR00643	2	593	MORPHINE 6-DEHYDROGENASE (EC 1.1.1.218)
189	190	RXN02351	GR00679	132	1070	NITRILOTRACETATE MONOOXYGENASE COMPONENT A (EC 1.14.13.-)
191	192	RXN00905	WV0238	8075	8875	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
193	194	F RXA00905	GR00247	2	694	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
195	196	RXA00906	GR00247	630	1133	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
197	198	RXA00907	GR00247	1143	1265	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
199	200	RXA02101	GR00631	3104	1842	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
201	202	RXN02565	WV0154	14299	13034	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
203	204	F RXA02566	GR00733	1	342	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
205	206	F RXA02567	GR00734	3	740	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
207	208	RXN03077	WV0043	1729	2913	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
209	210	F RXA02855	GR10002	1693	2877	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
211	212	RXA000026	GR00003	3657	5042	Hypothetical Metal-Dependent Hydrolase
213	214	RXA01971	GR00569	963	133	Predicted hydrolases (HAD superfamily)
215	216	RXA01802	GR00509	3461	4291	Predicted Zn-dependent hydrolases
217	218	RXN00866	WV0258	3557	4522	Predicted Zn-dependent hydrolases
219	220	F RXA00866	GR00236	3585	4499	Predicted Zn-dependent hydrolases
221	222	RXA02410	GR00703	792	127	Predicted Zn-dependent hydrolases

Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
223	224	RXA00961	GR00267	2	433	SALICYLATE HYDROXYLASE (EC 1.14.13.1)
225	226	RXA00111	GR00016	930	1922	SOLUBLE EPOXIDE HYDROLASE (SEH) (EC 3.3.2.3)
227	228	RXA01932	GR00555	6479	5583	ACETYL-HYDROLASE (EC 3.1.1.-)
229	230	RXA02574	GR00739	833	1840	PUTATIVE SECRETED HYDROLASE
231	232	RXN00983	VW0231	1796	321	SALIDASE PRECURSOR (EC 3.2.1.18)
233	234	FRXA00983	GR00278	1200	4	SALIDASE PRECURSOR (EC 3.2.1.18)
235	236	RXA00984	GR00278	1716	1300	SALIDASE PRECURSOR (EC 3.2.1.18)
237	238	RXA02513	VW0193	737	6	SALIDASE PRECURSOR (EC 3.2.1.18)
239	240	FRXA02513	GR00722	93	824	SALIDASE PRECURSOR (EC 3.2.1.18)
241	242	RXA00903	GR00246	637	5	Putative epimerase
243	244	RXA01224	GR00354	4186	5208	2-NITROPROPANE DIOXYGENASE (EC 1.13.11.32)
245	246	RXA01571	GR00438	1360	1959	ALCOHOL DEHYDROGENASE (EC 1.1.1.1)
247	248	RXN02478	VW0119	7584	6350	SALIDASE PRECURSOR (EC 3.2.1.18)
249	250	RXN00343	VW0125	1118	6	SALIDASE PRECURSOR (EC 3.2.1.18)
251	252	RXN01555	VW0135	29820	29861	3-OXOSTEROID 1-DEHYDROGENASE (EC 1.3.99.4)
253	254	RXN01166	VW0117	18142	1787	3-OXOSTEROID 1-DEHYDROGENASE (EC 1.3.99.4)
255	256	RXN02001	VW0326	630	1787	EXTRACELLULAR LIPASE PRECURSOR (EC 3.1.1.3)
257	258	RXN03145	VW0142	7561	7115	N-ACYL-L-AMINO ACID AMIDOHYDROLASE (EC 3.5.1.14)
259	260	RXN01466	VW0019	7050	6091	4-OXALOGROTONATE TAUTOMERASE (EC 5.3.2.-)
261	262	RXN01145	VW0077	7538	6825	ARYLESTERASE (EC 3.1.1.2)
263	264	RXN03088	VW0032	3431	3817	KETOL-ACID REDUCTOISOMERASE (EC 1.1.1.86)
265	266	RXN02952	VW0320	1032	1947	Hypothetical Methyltransferase (EC 2.1.1.-)
267	268	RXN00513	VW0092	1573	653	PUTATIVE REDUCTASE CARBOXYVINYL-CARBOXYPHOSPHONATE PHOSPHORYLMUTASE (EC 2.7.8.23)
269	270	RXN01152	VW0136	1740	907	PROTEIN-L-ISOASPARTATE O-METHYLTRANSFERASE (EC 2.1.1.77)
271	272	RXN00787	VW0321	3736	5637	D-AMINO ACID DEHYDROGENASE LARGE SUBUNIT (EC 1.4.99.1)

N-metabolism

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
273	274	RXN01302	VW0148	2837	2385	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
275	276	FRXA01302	GR00376	370	5	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
277	278	RXN01308	VW0148	2406	4	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
279	280	FRXA01307	GR00377	686	6	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
281	282	FRXA01308	GR00378	1211	6	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
283	284	RXN01309	VW0158	1	801	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
285	286	FRXA01309	GR00379	719	51	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
287	288	RXA02017	GR00610	1731	1048	NITRATE REDUCTASE ALPHA CHAIN (EC 1.7.99.4)
289	290	RXA02018	GR00610	2788	1739	NITRATE REDUCTASE BETA CHAIN (EC 1.7.99.4)
291	292	RXA02016	GR00610	1036	280	NITRATE REDUCTASE BETA CHAIN (EC 1.7.99.4)
293	294	RXA00471	GR00119	2997	3886	NITRATE TENINITRILE RESPONSE REGULATOR PROTEIN NARL
295	296	RXA00133	GR00021	201	1013	NITRATE TENINITRILE RESPONSE REGULATOR PROTEIN NARP
297	298	FRXA00650	GR00169	4017	3382	NITRATE TENINITRILE RESPONSE REGULATOR PROTEIN NARP

Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Config.	NT Start	NT Stop	Function
299	300	RXA01189	GR00339	2545	1937	NITRATE/NITRITE RESPONSE REGULATOR PROTEIN NARP
301	302	RXA01607	GR00449	123	752	NITRATE/NITRITE RESPONSE REGULATOR PROTEIN NARP
303	304	RXN00470	WV0066	27401	28669	NITRATE/NITRITE SENSOR PROTEIN NARX (EC 2.7.3.-)
305	306	F RXA00470	GR00119	1752	2951	NITRATE/NITRITE SENSOR PROTEIN NARX (EC 2.7.3.-)
307	308	RXA00756	GR00203	2932	1937	N UTILIZATION SUBSTANCE PROTEIN A
309	310	RXA00139	GR00222	2514	3224	N UTILIZATION SUBSTANCE PROTEIN B
311	312	RXA001303	GR00376	1724	390	NITRITE EXTRUSION PROTEIN
313	314	RXA01412	GR00412	620	417	NITROGEN FIXATION PROTEIN FIXI (PROBABLE E1-E2 TYPE CATION ATPASE) (EC 3.6.1.-)
315	316	RXA00773	GR00205	3208	4350	NITROGEN REGULATION PROTEIN NIFR3
317	318	RXA02746	GR00764	1	267	NITROGEN REGULATORY PROTEIN P-II
319	320	RXA02745	GR00763	15350	14472	MODULATION ATP-BINDING PROTEIN I
321	322	RXN00820	WV0054	19455	19817	MODULATION PROTEIN N
323	324	F RXA00820	GR00221	1007	1369	MODULATION PROTEIN N
325	326	RXA001059	GR00296	6782	9390	OXYGEN-INSENSITIVE NAD(P)H NITROREDUCTASE (EC 1....-)
327	328	RXN01386	WV0008	39246	38317	NITRILASE REGULATOR
329	330	RXN00073	WV0154	2369	687	FERRDOXIN-NITRITE REDUCTASE (EC 1.7.7.1)
331	332	RXN03131	WV0127	276	4	RHIZOPNE CATABOLISM PROTEIN MOCC
333	334	RXS00153	WV0157	4195	4620	MODULATION PROTEIN

Urease

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO
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Identification Code	Config.	NT Start	NT Stop	Function
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Phosphate and Phosphonate metabolism

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO
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Identification Code	Config.	NT Start	NT Stop	Function
RXN01716	WV0319	3259	2774	EXOPOLYPHOSPHATASE (EC 3.6.1.11)
RXN02972	WV0319	2763	2353	EXOPOLYPHOSPHATASE (EC 3.6.1.11)
RXN00663	WV0142	10120	11493	PHOH PROTEIN HOMOLOG
RXN00778	WV0103	18126	19250	PHOSPHATE-BINDING PERIPLASMIC PROTEIN PRECURSOR
RXN00250	WV0189	286	1032	DEDA PROTEIN - ALKALINE PHOSPHATASE LIKE PROTEIN

Sulfate metabolism

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO
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Identification Code	Config.	NT Start	NT Stop	Function
RXA00072	GR00012	446	6	PHOSPHOADENOSINE PHOSPHOSULFATE REDUCTASE (EC 1.8.99.4)
RXA00793	GR00211	1469	2644	SULFATE STARVATION-INDUCED PROTEIN 6
RXA01192	GR00342	161	733	SULFATE STARVATION-INDUCED PROTEIN 6

Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
351	352	RXA0715	GR00188	2120	2914	THIOSULFATE SULFURTRANSFERASE (EC 2.8.1.1)
353	354	RXA01664	GR00463	1306	485	THIOSULFATE SULFURTRANSFERASE (EC 2.8.1.1)
355	356	RXN02334	W0141	7939	7217	THIOSULFATE SULFURTRANSFERASE (EC 2.8.1.1)
357	358	F RXA02334	GR00672	2	335	THIOSULFATE SULFURTRANSFERASE (EC 2.8.1.1)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
359	360	RXN01499	W0008	7034	3213	ENTEROBACTIN SYNTHETASE COMPONENT F
361	362	RXN01997	W0084	33308	33793	FERRITIN

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
363	364	RXA01848	GR00524	1532	789	MAGNESIUM-CHELATASE SUBUNIT CHLI
365	366	RXN01849	W0139	16415	17515	MAGNESIUM-CHELATASE SUBUNIT CHLI
367	368	F RXA01849	GR00524	2004	1395	MAGNESIUM-CHELATASE SUBUNIT CHLI
369	370	F RXA01691	GR00474	370	4	MAGNESIUM-CHELATASE SUBUNIT CHLI
371	372	RXN00665	W0252	135	635	MG2+/CITRATE COMPLEX SECONDARY TRANSPORTER

Fe-Metabolism

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
359	360	RXN01499	W0008	7034	3213	ENTEROBACTIN SYNTHETASE COMPONENT F
361	362	RXN01997	W0084	33308	33793	FERRITIN

Mg Metabolism

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
363	364	RXA01848	GR00524	1532	789	MAGNESIUM-CHELATASE SUBUNIT CHLI
365	366	RXN01849	W0139	16415	17515	MAGNESIUM-CHELATASE SUBUNIT CHLI
367	368	F RXA01849	GR00524	2004	1395	MAGNESIUM-CHELATASE SUBUNIT CHLI
369	370	F RXA01691	GR00474	370	4	MAGNESIUM-CHELATASE SUBUNIT CHLI
371	372	RXN00665	W0252	135	635	MG2+/CITRATE COMPLEX SECONDARY TRANSPORTER

Modification and degradation of aromatic compounds

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
373	374	RXN02026	W0007	28635	28901	3-DEHYDROQUINATE DEHYDRATASE (EC 4.2.1.10)
375	376	RXN02308	W0025	8507	8247	O-SUCCINYL-BENZOIC ACID-COA LIGASE (EC 6.2.1.26)
377	378	RXN03000	W0025	570	4	SALICYLATE HYDROXYLASE (EC 1.14.13.1)
379	380	RXN03036	W0014	671	6	PROTocatechuate 3,4-dioxygenase BETA CHAIN (EC 1.13.11.3)
381	382	RXN02374	W00229	12631	12437	4-NITROPHENYLPHOSPHATASE (EC 3.1.3.41)
383	384	RXN00393	W0025	7241	6348	1,4-DIHYDROXY-2-NAPHTHOATE OCTAPRENYLTRANSFERASE (EC 2.5.-)
385	386	RXN00948	W0107	4266	5384	12-oxophthalate reductase (EC 1.3.1.42)
387	388	RXN01923	W0020	3384	4133	2-HYDROXY-6-OXO-6-PHENYLHEXA-2,4-DIENOATE HYDROLASE (EC 3.7.1.-)
389	390	RXN00398	W0025	14633	13984	2-PYRONE-4,6-DICARBOXYLATE LACTONASE (EC 3.1.1.57)
391	392	RXN02813	W0128	13120	14118	3-CARBOXY-CIS-CIS-MUCONATE CYCLOISOMERASE HOMOLOG (EC 5.5.1.2)
393	394	RXN0136	W0134	13373	14467	3-DEHYDROQUINATE SYNTHASE (EC 4.6.1.3)
395	396	RXN02508	W0007	26733	28586	3-DEHYDROQUINATE DEHYDRATASE (EC 4.2.1.-)
397	398	RXN02839	W00362	3	449	4-HYDROXYBENZOATE OCTAPRENYLTRANSFERASE (EC 2.5.1.-)
399	400	RXN00539	W0128	7858	8712	CATECHOL 1,2-DIOXYGENASE (EC 1.13.11.1)
401	402	RXN02530	W0057	5469	6125	DIMETHYLANILINE MONOOXYGENASE (EC 1.6.5.5)
403	404	RXN00434	W0112	12078	11212	QUINONE OXIDOREDUCTASE (EC 1.6.5.5)
405	406	RXN01616	W0050	24649	23875	QUINONE OXIDOREDUCTASE (EC 1.6.5.5)
407	408	RXN01842	W0234	1615	2532	QUINONE OXIDOREDUCTASE (EC 1.6.5.5)

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Table 1 (continued)

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
409	410	RXN00541	VW0128	7440	5950	TOLUATE 1,2-DIOXYGENASE ALPHA SUBUNIT (EC 1.14.12.-)
411	412	RXN01993	VW0182	16	1143	VANILLATE DEMETHYLASE (EC 1.14.-)
413	414	RXN00558	VW0083	15705	16397	PHENOL 2-MONOXYGENASE (EC 1.14.13.7)
415	416	RXN00178	VW0174	14670	15554	hydroxyquinol 1,2-dioxygenase (EC 1.13.11.37)
417	418	RXN01461	VW0128	12414	13025	PROTOCATECHUATE 3,4-DIOXYGENASE ALPHA CHAIN (EC 1.13.11.3)
419	420	RXN01653	VW0321	12867	11407	DIBENZOTHIOPHENE DESULFURIZATION ENZYME A
421	422	RXN02053	VW0009	39448	40026	DRGA PROTEIN
423	424	RXN001777	VW0174	13589	14656	MALEYLACETATE REDUCTASE (EC 1.3.1.32)
425	426	RXC005953	VW0249	1816	2652	PROTEIN involved in degradation of aromatic compounds

Modification and degradation of aliphatic compounds

Nucleic Acid SEQ ID NO	Amino Acid SEQ ID NO	Identification Code	Contig.	NT Start	NT Stop	Function
427	428	RXN00259	VW0176	43379	42402	ALKANAL MONOOXYGENASE ALPHA CHAIN (EC 1.14.14.3)
429	430	F RXA00299	GR00048	7376	6633	ALKANAL MONOOXYGENASE ALPHA CHAIN (EC 1.14.14.3)
431	432	RXA00332	GR00057	16086	15385	ALKANAL MONOOXYGENASE ALPHA CHAIN (EC 1.14.14.3)
433	434	RXA01838	GR00519	2	820	ALKANAL MONOOXYGENASE ALPHA CHAIN (EC 1.14.14.3)
435	436	RXA02643	GR00750	1603	560	ALKANAL MONOOXYGENASE ALPHA CHAIN (EC 1.14.14.3)
437	438	RXA019333	GR00555	6590	7192	2-HALOALKANOIC ACID DEHALOGENASE I (EC 3.8.1.2)
439	440	RXA02351	GR00679	132	1070	NITRILOTRIACETATE MONOOXYGENASE COMPONENT A (EC 1.14.13.-)

TABLE 2 - Excluded Genes

GenBank™ Accession No.	Gene Name	Gene Function	Reference
A09073	peg	Phosphoenol pyruvate carboxylase	Bachmann, B. et al. "DNA fragment coding for phosphoenolpyruvate carboxylase, recombinant DNA carrying said fragment, strains carrying the recombinant DNA and method for producing L-amino acids using said strains," Patent: EP 0358940-A 3 03/21/90
A45579, A45581, A45583, A45585 A45587		Threonine dehydratase	Moockel, B. et al. "Production of L-isoleucine by means of recombinant micro-organisms with deregulated threonine dehydratase," Patent: WO 9519442-A 5 07/20/95
AB003132	murC; fisQ; fisZ		Kobayashi, M. et al. "Cloning, sequencing, and characterization of the fisZ gene from coryneform bacteria," <i>Biochem. Biophys. Res. Commun.</i> , 236(2):383-388 (1997)
AB015023	murC; fisQ		Wachi, M. et al. "A murC gene from Coryneform bacteria," <i>Appl. Microbiol. Biotechnol.</i> , 51(2):223-228 (1999)
AB018530	disR		Kimura, E. et al. "Molecular cloning of a novel gene, disR, which rescues the detergent sensitivity of a mutant derived from <i>Brevibacterium lactofermentum</i> ," <i>Bioc. Biotechnol. Biochem.</i> , 60(10):1565-1570 (1996)
AB018531	disR1; disR2		
AB020624	murI	D-glutamate racemase	
AB023377	tkt	transketolase	
AB024708	glbB; glbD	Glutamine 2-oxoglutarate aminotransferase large and small subunits	
AB025424	acn	aconitase	
AB027714	rep	Replication protein	
AB027715	rep; aad	Replication protein; aminoglycoside adenylyltransferase	
AF005242	argC	N-acetylglutamate-5-semialdehyde dehydrogenase	
AF005635	ghlA	Glutamine synthetase	
AF030405	hisF	cyclase	
AF030520	argG	Argininosuccinate synthetase	
AF031518	argF	Ornithine carbamoyltransferase	
AF036932	aroD	3-dehydroquinate dehydratase	
AF038548	pyc	Pyruvate carboxylase	

Table 2 (continued)

	dcIAE; apt; rel	Dipeptide-binding protein; adenine phosphoribosyltransferase; GTP pyrophosphokinase	Wehmeier, L. et al. "The role of the <i>Corynebacterium glutamicum</i> rel gene in (p)ppGpp metabolism," <i>Microbiology</i> , 144:1853-1862 (1998)
AF038651			
AF041436	argR	Arginine repressor	
AF045998	inpA	Inositol monophosphate phosphatase	
AF048764	argH	Argininosuccinate lyase	
AF049897	argC; argJ; argB; argD; argF; argR; argG; argH	N-acetylglutanylphosphate reductase; ornithine acetyltransferase; N-acetylglutamate kinase; acetylornithine transaminase; ornithine carbamoyltransferase; arginine repressor; argininosuccinate synthase; argininosuccinate lyase	
AF050109	inhA	Enoyl-acyl carrier protein reductase	
AF050166	hisG	ATP phosphoribosyltransferase	
AF051846	hisA	Phosphoribosylformimino-5-amino-1-phosphoribosyl-4-imidazolecarboxamide isomerase	
AF052652	metA	Homoserine O-acetyltransferase	Park, S. et al. "Isolation and analysis of metA, a methionine biosynthetic gene encoding homoserine acetyltransferase in <i>Corynebacterium glutamicum</i> ," <i>Mol. Cells</i> , 8(3):286-294 (1998)
AF053071	aroB	Dehydroquininate synthetase	
AF060558	hisH	Glutamine amidotransferase	
AF06704	hisE	Phosphoribosyl-ATP-pyrophosphotransferase	
AF114233	aroA	5-enolpyruvylshikimate 3-phosphate synthase	
AF116184	panD	L-aspartate-alpha-decarboxylase precursor	Dusch, N. et al. "Expression of the <i>Corynebacterium glutamicum</i> panD gene encoding L-aspartate-alpha-decarboxylase leads to pantothenate overproduction in <i>Escherichia coli</i> ," <i>Appl. Environ. Microbiol.</i> , 65(4):1530-1539 (1999)
AF124518	aroD; aroE	3-dehydroquinase; shikimate dehydrogenase	
AF124600	aroC; aroK; aroB; pppQ	Chorismate synthase; shikimate kinase; 3-dehydroquinase synthase; putative cytoplasmic peptidase	
AF145897	inhA		
AF145898	inhA		

Table 2 (continued)

A1001436	ectP	Transport of ectoine, glycine betaine, proline	Peter, H. et al. "Corynebacterium glutamicum is equipped with four secondary carriers for compatible solutes: Identification, sequencing, and characterization of the proline/ectoine uptake system, ProP, and the ectoine/proline/glycine betaine carrier, EctP." <i>J. Bacteriol.</i> , 180(22):6005-6012 (1998)
A1004934	dapD	Tetrahydrodipicolinate succinylase (incomplete)	Wehrmann, A. et al. "Different modes of diaminopimelate synthesis and their role in cell wall integrity: A study with <i>Corynebacterium glutamicum</i> ." <i>J. Bacteriol.</i> , 180(12):3159-3165 (1998)
A1007732	ppc; secG; amt; ood; sox A	Phosphoenolpyruvate-carboxylase; ?; high affinity ammonium uptake protein; putative ornithine-cyclodecarboxylase; sarcosine oxidase	
A1010319	ftsY, glnB, glnD; srp; amtP	Involved in cell division; PII protein; uridylyltransferase (uridylyl-removing enzyme); signal recognition particle; low affinity ammonium uptake protein	Jakoby, M. et al. "Nitrogen regulation in <i>Corynebacterium glutamicum</i> : Isolation of genes involved in biochemical characterization of corresponding proteins," <i>FEBS Microbiol.</i> , 173(2):303-310 (1999)
A1132968	cat	Chloramphenicol acetyl transferase	
A1224946	mqo	L-malate: quinone oxidoreductase	Molenaar, D. et al. "Biochemical and genetic characterization of the membrane-associated malate dehydrogenase (acceptor) from <i>Corynebacterium glutamicum</i> ," <i>Eur. J. Biochem.</i> , 254(2):395-403 (1998)
A1238250	ndh	NADH dehydrogenase	
A1238703	porA	Porin	Lichtinger, T. et al. "Biochemical and biophysical characterization of the cell wall porin of <i>Corynebacterium glutamicum</i> : The channel is formed by a low molecular mass polypeptide," <i>Biochemistry</i> , 37(43):15024-15032 (1998)
D17429		Transposable element IS31831	Vertes et al. "Isolation and characterization of IS31831, a transposable element from <i>Corynebacterium glutamicum</i> ," <i>Mol. Microbiol.</i> , 11(4):739-746 (1994)
D84102	odhA	2-oxoglutarate dehydrogenase	Usuda, Y. et al. "Molecular cloning of the <i>Corynebacterium glutamicum</i> (Brevibacterium lactofermentum AJ12036) odhA gene encoding a novel type of 2-oxoglutarate dehydrogenase," <i>Microbiology</i> , 142:3347-3354 (1996)
E01358	hdh; hk	Homoserine dehydrogenase; homoserine kinase	Katsumata, R. et al. "Production of L-threonine and L-isoleucine," Patent: JP 1987232392-A 1 10/12/87
E01359		Upstream of the start codon of homoserine kinase gene	Katsumata, R. et al. "Production of L-threonine and L-isoleucine," Patent: JP 1987232392-A 2 10/12/87
E01375		Tryptophan operon	
E01376	trpL; trpE	Leader peptide; anthranilate synthase	Matsui, K. et al. "Tryptophan operon, peptide and protein coded thereby, utilization of tryptophan operon gene expression and production of tryptophan," Patent: JP 1987244382-A 1 10/24/87

Table 2 (continued)		
E01377	Promoter and operator regions of tryptophan operon	Matsui, K. et al. "Tryptophan operon, peptide and protein coded thereby, utilization of tryptophan operon gene expression and production of tryptophan," Patent: JP 1987244382-A 1 10/24/87
E03937	Biotin-synthase	Hatakeyama, K. et al. "DNA fragment containing gene capable of coding biotin synthetase and its utilization," Patent: JP 1992278088-A 1 10/02/92
E04040	Diamino pelargonic acid aminotransferase	Kohama, K. et al. "Gene coding diamino pelargonic acid aminotransferase and deshydrobiotin synthetase and its utilization," Patent: JP 1992330284-A 1 11/18/92
E04041	Deshydrobiotinsynthetase	Kohama, K. et al. "Gene coding diamino pelargonic acid aminotransferase and deshydrobiotin synthetase and its utilization," Patent: JP 1992330284-A 1 11/18/92
E04307	Flavum aspartase	Kurusu, Y. et al. "Gene DNA coding aspartase and utilization thereof," Patent: JP 1993030977-A 1 02/09/93
E04376	Isoctiric acid lyase	Katsumata, R. et al. "Gene manifestation controlling DNA," Patent: JP 1993056782-A 3 03/09/93
E04377	Isoctiric acid lyase N-terminal fragment	Katsumata, R. et al. "Gene manifestation controlling DNA," Patent: JP 1993056782-A 3 03/09/93
E04484	Prephenate dehydratase	Sotouchi, N. et al. "Production of L-phenylalanine by fermentation," Patent: JP 1993076352-A 2 03/30/93
E05108	Aspartokinase	Fugono, N. et al. "Gene DNA coding Aspartokinase and its use," Patent: JP 1993184366-A 1 07/27/93
E05112	Dihydro-dipichorinate synthetase	Hatakeyama, K. et al. "Gene DNA coding dihydrodipicolinic acid synthetase and its use," Patent: JP 1993184371-A 1 07/27/93
E05776	Diaminopimelic acid dehydrogenase	Kobayashi, M. et al. "Gene DNA coding Diaminopimelic acid dehydrogenase and its use," Patent: JP 1993284970-A 1 11/02/93
E05779	Threonine synthase	Kohama, K. et al. "Gene DNA coding threonine synthase and its use," Patent: JP 1993284972-A 1 11/02/93
E06110	Prephenate dehydratase	Kikuchi, T. et al. "Production of L-phenylalanine by fermentation method," Patent: JP 1993344881-A 1 12/27/93
E06111	Mutated Prephenate dehydratase	Kikuchi, T. et al. "Production of L-phenylalanine by fermentation method," Patent: JP 1993344881-A 1 12/27/93
E06146	Acetohydroxy acid synthetase	Inui, M. et al. "Gene capable of coding Acetohydroxy acid synthetase and its use," Patent: JP 1993344893-A 1 12/27/93
E06825	Aspartokinase	Sugimoto, M. et al. "Mutant aspartokinase gene," patent: JP 1994062866-A 1 03/08/94
E06826	Mutated aspartokinase alpha subunit	Sugimoto, M. et al. "Mutant aspartokinase gene," patent: JP 1994062866-A 1 03/08/94

Table 2 (continued)

E06827		Mutated aspartokinase alpha subunit	Sugimoto, M. et al. "Mutant aspartokinase gene," patent: JP 1994062866-A 1 03/08/94
E07701	sec Y		Homo, N. et al. "Gene DNA participating in integration of membrane protein to membrane," Patent: JP 1994169780-A 1 06/21/94
E08177		Aspartokinase	Sato, Y. et al. "Genetic DNA capable of coding Aspartokinase released from feedback inhibition and its utilization," Patent: JP 1994261766-A 1 09/20/94
E08178, E08179, E08180, E08181, E08182		Feedback inhibition-released Aspartokinase	Sato, Y. et al. "Genetic DNA capable of coding Aspartokinase released from feedback inhibition and its utilization," Patent: JP 1994261766-A 1 09/20/94
E08232		Acetohydroxy-acid isomeroreductase	Inui, M. et al. "Gene DNA coding acetohydroxy acid isomeroreductase," Patent: JP 1994277067-A 1 10/04/94
E08234	secE		Asai, Y. et al. "Gene DNA coding for translocation machinery of protein," Patent: JP 1994277073-A 1 10/04/94
E08643		FT aminotransferase and decthiobiotin synthetase promoter region	Hatakeyama, K. et al. "DNA fragment having promoter function in coryneform bacterium," Patent: JP 1995031476-A 1 02/03/95
E08646		Biotin synthetase	Hatakeyama, K. et al. "DNA fragment having promoter function in coryneform bacterium," Patent: JP 1995031476-A 1 02/03/95
E08649		Aspartase	Kohama, K. et al. "DNA fragment having promoter function in coryneform bacterium," Patent: JP 1995031478-A 1 02/03/95
E08900		Dihydrodipicolinate reductase	Madori, M. et al. "DNA fragment containing gene coding Dihydrodipicolinate acid reductase and utilization thereof," Patent: JP 1995075578-A 1 03/20/95
E08901		Diaminopimelic acid decarboxylase	Madori, M. et al. "DNA fragment containing gene coding Diaminopimelic acid decarboxylase and utilization thereof," Patent: JP 1995075579-A 1 03/20/95
E12594		Serine hydroxymethyltransferase	Hatakeyama, K. et al. "Production of L-tryptophan," Patent: JP 1997028391-A 1 02/04/97
E12760, E12759, E12758		transposase	Moriya, M. et al. "Amplification of gene using artificial transposon," Patent: JP 1997070291-A 03/18/97
E12764		Arginyl-tRNA synthetase; diaminopimelic acid decarboxylase	Moriya, M. et al. "Amplification of gene using artificial transposon," Patent: JP 1997070291-A 03/18/97
E12767		Dihydrodipicolinic acid synthetase	Moriya, M. et al. "Amplification of gene using artificial transposon," Patent: JP 1997070291-A 03/18/97
E12770		aspartokinase	Moriya, M. et al. "Amplification of gene using artificial transposon," Patent: JP 1997070291-A 03/18/97
E12773		Dihydrodipicolinic acid reductase	Moriya, M. et al. "Amplification of gene using artificial transposon," Patent: JP 1997070291-A 03/18/97

Table 2 (continued)

E13655		Glucose-6-phosphate dehydrogenase	Harakeyama, K. et al. "Glucose-6-phosphate dehydrogenase and DNA capable of coding the same," Patent: JP 199724661-A. 1 09/02/97
L01508	IlvA	Threonine dehydratase	Moockel, B. et al. "Functional and structural analysis of the threonine dehydratase of Corynebacterium glutamicum," <i>J. Bacteriol.</i> , 174:8065-8072 (1992)
L07603	BC 4.2.1.15	3-deoxy-D-arabinoheptulosonate-7-phosphate synthase	Chen, C. et al. "The cloning and nucleotide sequence of Corynebacterium glutamicum 3-deoxy-D-arabinoheptulosonate-7-phosphate synthase gene," <i>FEMS Microbiol. Lett.</i> , 107:223-230 (1993)
L09232	IlvB; ilvN; ilvC	Acetylhydroxy acid synthase large subunit; Acetylhydroxy acid synthase small subunit; Acetylhydroxy acid isomeroreductase	Keilhauer, C. et al. "Isolation and synthesis in Corynebacterium glutamicum: molecular analysis of the ilvB-ilvN-ilvC operon," <i>J. Bacteriol.</i> , 175(17):5595-5603 (1993)
L18874	PtsM	Phosphoenolpyruvate sugar phosphotransferase	Fouet, A. et al. "Bacillus subtilis sucrose-specific enzyme II of the phosphotransferase system: expression in Escherichia coli and homology to enzymes II from enteric bacteria," <i>PNAS USA</i> , 84(24):8773-8777 (1987); Lee, J.K. et al. "Nucleotide sequence of the gene encoding the Corynebacterium glutamicum mannose enzyme II and analyses of the deduced protein sequence," <i>FEMS Microbiol. Lett.</i> , 119(1-2):137-145 (1994)
L27123	aceB	Malate synthase	Lee, H.-S. et al. "Molecular characterization of aceB, a gene encoding malate synthase in Corynebacterium glutamicum," <i>J. Microbiol. Biotechnol.</i> , 4(4):256-263 (1994)
L27126		Pyruvate kinase	Jetten, M. S. et al. "Structural and functional analysis of pyruvate kinase from Corynebacterium glutamicum," <i>Appl. Environ. Microbiol.</i> , 60(7):2501-2507 (1994)
L28760	aceA	Isocitrate lyase	Oguiza, J.A. et al. "Molecular cloning, DNA sequence analysis, and characterization of the Corynebacterium diptheriae dxtR from Brevibacterium lactofermentum," <i>J. Bacteriol.</i> , 177(2):465-467 (1995)
L35906	dxtR	Diphtheria toxin repressor	Follettie, M.T. et al. "Molecular cloning and nucleotide sequence of the Corynebacterium glutamicum pheA gene," <i>J. Bacteriol.</i> , 167:695-702 (1986)
M13774		Prephenate dehydratase	Park, Y.-H. et al. "Phylogenetic analysis of the coryneform bacteria by 5S rRNA sequences," <i>J. Bacteriol.</i> , 169:1801-1806 (1987)
M16175	5S rRNA		Sano, K. et al. "Structure and function of the trp operon control regions of Brevibacterium lactofermentum, a glutamic-acid-producing bacterium," <i>Gene</i> , 52:191-200 (1987)
M16663	trpE	Anthrax toxin synthase, 5' end	
M16664	trpA	Tryptophan synthase, 3' end	Sano, K. et al. "Structure and function of the trp operon control regions of Brevibacterium lactofermentum, a glutamic-acid-producing bacterium," <i>Gene</i> , 52:191-200 (1987)

Table 2 (continued)

M25819		Phosphoenolpyruvate carboxylase	O'Regan, M. et al. "Cloning and nucleotide sequence of the Phosphoenolpyruvate carboxylase-coding gene of <i>Corynebacterium glutamicum</i> ATCC 13032," <i>Gene</i> 77(2):237-251 (1989)
M85106		23S rRNA gene insertion sequence	Roller, C. et al. "Gram-positive bacteria with a high DNA G+C content are characterized by a common insertion within their 23S rRNA genes," <i>J. Gen. Microbiol.</i> , 138:1167-1175 (1992)
M85107, M85108		23S rRNA gene insertion sequence	Roller, C. et al. "Gram-positive bacteria with a high DNA G+C content are characterized by a common insertion within their 23S rRNA genes," <i>J. Gen. Microbiol.</i> , 138:1167-1175 (1992)
M89931	acdD; bmq; ybbw	Beta C-S lyase; branched-chain amino acid uptake carrier; hypothetical protein ybbw	Rosol, I. et al. "The <i>Corynebacterium glutamicum</i> acdD gene encodes a C-S lyase with alpha, beta-elimination activity that degrades aminothioleysteine," <i>J. Bacteriol.</i> 174(9):2968-2977 (1992); Tauch, A. et al. "Isolucine uptake in <i>Corynebacterium glutamicum</i> ATCC 13032 is directed by the bmq gene product," <i>Arch. Microbiol.</i> , 169(4):303-312 (1998)
S59299	trp	Leader gene (promoter)	Herry, D.M. et al. "Cloning of the trp gene cluster from a tryptophan-producing strain of <i>Corynebacterium glutamicum</i> : identification of a mutation in the trp leader sequence," <i>Appl. Environ. Microbiol.</i> , 59(3):791-799 (1993)
U11545	trpD	Anthranilate phosphoribosyltransferase	O'Gara, J.P. and Dunican, L.K. (1994) Complete nucleotide sequence of the <i>Corynebacterium glutamicum</i> ATCC 21850 trpD gene," Thesis, Microbiology Department, University College Galway, Ireland.
U13922	cgIIIM; cgIIR; elgIIR	Putative type II 5-cytosine methyltransferase; putative type II restriction endonuclease; putative type I or type III restriction endonuclease	Schafer, A. et al. "Cloning and characterization of a DNA region encoding a stress-sensitive restriction system from <i>Corynebacterium glutamicum</i> ATCC 13032 and analysis of its role in intergeneric conjugation with <i>Escherichia coli</i> ," <i>J. Bacteriol.</i> , 176(23):7305-7319 (1994); Schafer, A. et al. "The <i>Corynebacterium glutamicum</i> cgIIIM gene encoding a 5-cytosine in an McrBC-deficient <i>Escherichia coli</i> strain," <i>Gene</i> , 203(2):95-101 (1997)
U14965	recA		Ankri, S. et al. "Mutations in the <i>Corynebacterium glutamicum</i> proline biosynthetic pathway: A natural bypass of the proA step," <i>J. Bacteriol.</i> , 178(15):4412-4419 (1996)
U31224	ppx		Ankri, S. et al. "Mutations in the <i>Corynebacterium glutamicum</i> proline biosynthetic pathway: A natural bypass of the proA step," <i>J. Bacteriol.</i> , 178(15):4412-4419 (1996)
U31225	proC	L-proline: NADP+ 5-oxidoreductase	Ankri, S. et al. "Mutations in the <i>Corynebacterium glutamicum</i> proline biosynthetic pathway: A natural bypass of the proA step," <i>J. Bacteriol.</i> , 178(15):4412-4419 (1996)
U31230	obg; proB; unkdh	?gamma glutamyl kinase; similar to D-isomer specific 2-hydroxyacid dehydrogenases	Ankri, S. et al. "Mutations in the <i>Corynebacterium glutamicum</i> proline biosynthetic pathway: A natural bypass of the proA step," <i>J. Bacteriol.</i> , 178(15):4412-4419 (1996)

Table 2 (continued)

U31281	bioB	Biotin synthase	Serebriskii, I.G., "Two new members of the bio B superfamily: Cloning, sequencing and expression of bio B genes of <i>Methylobacillus flagellatum</i> and <i>Corynebacterium glutamicum</i> ," <i>Gene</i> , 175:15-22 (1996)
U35023	tnrR, accBC	Thiosulfate sulfurtransferase; acyl CoA carboxylase	Jager, W. et al. "A <i>Corynebacterium glutamicum</i> gene encoding a two-domain protein similar to biotin carboxylases and biotin-carboxyl-carrier proteins," <i>Arch. Microbiol.</i> , 166(2):76-82 (1996)
U45535	cmr	Multidrug resistance protein	Jager, W. et al. "A <i>Corynebacterium glutamicum</i> gene conferring multidrug resistance in the heterologous host <i>Escherichia coli</i> ," <i>J. Bacteriol.</i> , 179(7):2449-2451 (1997)
U45536	clpB	Heat shock ATP-binding protein	
U55587	aphA-3	3',5'-aminoglycoside phosphotransferase	
U89648		<i>Corynebacterium glutamicum</i> unidentified sequence involved in histidine biosynthesis; partial sequence	
X04960	trpA, trpB; trpC; trpD; trpE; trpG; trpL	Tryptophan operon	Matsui, K. et al. "Complete nucleotide and deduced amino acid sequences of the <i>Brevibacterium lactofermentum</i> tryptophan operon," <i>Nucleic Acids Res.</i> , 14(24):10113-10114 (1986)
X07563	lys A	DAP decarboxylase (meso-diaminopimelate decarboxylase, EC 4.1.1.20)	Yeh, P. et al. "Nucleic sequence of the lysA gene of <i>Corynebacterium glutamicum</i> and possible mechanisms for modulation of its expression," <i>Mol. Gen. Genet.</i> , 212(1):112-119 (1988)
X14234	EC 4.1.1.31	Phosphoenolpyruvate carboxylase	Eikmanns, B.J. et al. "The Phosphoenolpyruvate carboxylase gene of <i>Corynebacterium glutamicum</i> : Molecular cloning, nucleotide sequence, and expression," <i>Mol. Gen. Genet.</i> , 218(2):330-339 (1989); Lepiniec, L. et al. "Sorghum Phosphoenolpyruvate carboxylase gene family: structure, function and molecular evolution," <i>Plant. Mol. Biol.</i> , 21 (3):487-502 (1993)
X17313	fdx	Fructose-bisphosphate aldolase	Von der Osten, C.H. et al. "Molecular cloning, nucleotide sequence and fine-structural analysis of the <i>Corynebacterium glutamicum</i> fdx gene: structural comparison of <i>C. glutamicum</i> fructose-1, 6-bisphosphate aldolase to class I and class II aldolases," <i>Mol. Microbiol.</i>
X53993	dapA	L-2, 3-dihydrodipicolinate synthetase (EC 4.2.1.52)	Bonnasse, S. et al. "Nucleic sequence of the dapA gene from <i>Corynebacterium glutamicum</i> ," <i>Nucleic Acids Res.</i> , 18(21):6421 (1990)
X54223		AIB-related site	Cianciotto, N. et al. "DNA sequence homology between att B-related sites of <i>Corynebacterium diphtheriae</i> , <i>Corynebacterium ulcerans</i> , <i>Corynebacterium glutamicum</i> , and the attP site of <i>lambdacorynephage</i> ," <i>FEBS. Microbiol. Lett.</i> , 66:299-302 (1990)
X54740	argS, lysA	Arginyl-tRNA synthetase; Diaminopimelate decarboxylase	Marcel, T. et al. "Nucleotide sequence and organization of the upstream region of the <i>Corynebacterium glutamicum</i> lysA gene," <i>Mol. Microbiol.</i> , 4(11):1819-1830 (1990)

Table 2 (continued)

X55994	trpL, trpE	Purative leader peptide; anthranilate synthase component 1	Heery, D.M. et al. "Nucleotide sequence of the <i>Corynebacterium glutamicum</i> trpE gene," <i>Nucleic Acids Res.</i> , 18(23):7138 (1990)
X56037	thrC	Threonine synthase	Han, K.S. et al. "The molecular structure of the <i>Corynebacterium glutamicum</i> threonine synthase gene," <i>Mol. Microbiol.</i> , 4(10):1693-1702 (1990)
X56075	attB-related site	Attachment site	Cianciotto, N. et al. "DNA sequence homology between att B-related sites of <i>Corynebacterium diptheriae</i> , <i>Corynebacterium ulcerans</i> , <i>Corynebacterium glutamicum</i> , and the attP site of <i>lambdacorynephage</i> ," <i>FEMS. Microbiol. Lett.</i> , 66:299-302 (1990)
X57226	lysC-alpha; lysC-beta; asd	Aspartokinase-alpha subunit; Aspartokinase-beta subunit; aspartate beta semialdehyde dehydrogenase	Kalinowski, J. et al. "Genetic and biochemical analysis of the Aspartokinase from <i>Corynebacterium glutamicum</i> ," <i>Mol. Microbiol.</i> , 5(5):1197-1204 (1991); Kalinowski, J. et al. "Aspartokinase genes lysC alpha and lysC beta overlap and are adjacent to the aspartate beta-semialdehyde dehydrogenase gene asd in <i>Corynebacterium glutamicum</i> ," <i>Mol. Gen. Genet.</i> , 224(3):317-324 (1990)
X59403	gap/pgk; tpi	Glyceraldehyde-3-phosphate; phosphoglycerate kinase; triosephosphate isomerase	Eikmanns, B.J. "Identification, sequence analysis, and expression of a <i>Corynebacterium glutamicum</i> gene cluster encoding the three glycolytic enzymes glyceraldehyde-3-phosphate dehydrogenase, 3-phosphoglycerate kinase, and triosephosphate isomerase," <i>J. Bacteriol.</i> , 174(19):6076-6086 (1992)
X59404	gdh	Glutamate dehydrogenase	Bormann, E.R. et al. "Molecular analysis of the <i>Corynebacterium glutamicum</i> gdh gene encoding glutamate dehydrogenase," <i>Mol. Microbiol.</i> , 6(3):317-326 (1992)
X60312	lysI	L-lysine permease	Siepe-Feldhaus, A.H. et al. "Molecular analysis of the <i>Corynebacterium glutamicum</i> lysI gene involved in lysine uptake," <i>Mol. Microbiol.</i> , 5(12):2995-3003 (1991)
X66078	copI	PSI protein	Jolliff, G. et al. "Cloning and nucleotide sequence of the copI gene encoding PSI, one of the two major secreted proteins of <i>Corynebacterium glutamicum</i> : The deduced N-terminal region of PSI is similar to the Mycobacterium antigen 85 complex," <i>Mol. Microbiol.</i> , 6(16):2349-2362 (1992)
X66112	glt	Citrate synthase	Eikmanns, B.J. et al. "Cloning sequence, expression and transcriptional analysis of the <i>Corynebacterium glutamicum</i> gltA gene encoding citrate synthase," <i>Microbiol.</i> , 140:1817-1828 (1994)
X67737	dapB	Dihydrodipicolinate reductase	
X69103	csp2	Surface layer protein PS2	Peyret, J.L. et al. "Characterization of the cspB gene encoding PS2, an ordered surface-layer protein in <i>Corynebacterium glutamicum</i> ," <i>Mol. Microbiol.</i> , 9(1):97-109 (1993)
X69104		IS3 related insertion element	Bonamy, C. et al. "Identification of ISI206, a <i>Corynebacterium glutamicum</i> IS3-related insertion sequence and phylogenetic analysis," <i>Mol. Microbiol.</i> , 14(3):571-581 (1994)

Table 2 (continued)

X70959	leuA	Isopropylmalate synthase	Patek, M. et al. "Leucine synthesis in Corynebacterium glutamicum: enzyme activities, structure of leuA, and effect of leuA inactivation on lysine synthesis," <i>Appl. Environ. Microbiol.</i> , 60(1):133-140 (1994)
X71489	icd	Isocitrate dehydrogenase (NADP+)	Eikmanns, B.J. et al. "Cloning sequence analysis, expression, and inactivation of the Corynebacterium glutamicum icd gene encoding isocitrate dehydrogenase and biochemical characterization of the enzyme," <i>J. Bacteriol.</i> , 177(3):774-782 (1995)
X72835	GDHA	Glutamate dehydrogenase (NADP+)	
X75083, X70584	mttA	S-methyltryptophan resistance	Heery, D.M. et al. "A sequence from a tryptophan-hyperproducing strain of Corynebacterium glutamicum encoding resistance to S-methyltryptophan," <i>Biochem. Biophys. Res. Commun.</i> , 201(3):1255-1262 (1994)
X75085	recA		Fitzpatrick, R. et al. "Construction and characterization of recA mutant strains of Corynebacterium glutamicum and Brevibacterium lactofermentum," <i>Appl. Microbiol. Biotechnol.</i> , 42(4):575-580 (1994)
X75504	aceA, thiX	Partial isocitrate lyase; ?	Reinscheid, D.J. et al. "Characterization of the isocitrate lyase gene from Corynebacterium glutamicum and biochemical analysis of the enzyme," <i>J. Bacteriol.</i> , 176(12):3474-3483 (1994)
X76875		ATPase beta-subunit	Ludwig, W. et al. "Phylogenetic relationships of bacteria based on comparative sequence analysis of elongation factor Tu and ATP-synthase beta-subunit genes," <i>Antonie Van Leeuwenhoek</i> , 64:285-303 (1993)
X77034	tuf	Elongation factor Tu	Ludwig, W. et al. "Phylogenetic relationships of bacteria based on comparative sequence analysis of elongation factor Tu and ATP-synthase beta-subunit genes," <i>Antonie Van Leeuwenhoek</i> , 64:285-303 (1993)
X77384	recA		Billman-Jacobe, H. "Nucleotide sequence of a recA gene from Corynebacterium glutamicum," <i>DNA Seq.</i> , 4(6):403-404 (1994)
X78491	aceB	Malate synthase	Reinscheid, D.J. et al. "Malate synthase from Corynebacterium glutamicum pta-ack operon encoding phosphotransacetylase: sequence analysis," <i>Microbiology</i> , 140:3099-3108 (1994)
X80629	16S rDNA	16S ribosomal RNA	Rainey, F.A. et al. "Phylogenetic analysis of the genera Rhodococcus and Norcardia and evidence for the evolutionary origin of the genus Norcardia from within the radiation of Rhodococcus species," <i>Microbiol.</i> , 141:523-528 (1995)
X81191	gluA; gluB; gluC; gluD	Glutamate uptake system	Kronmeyer, W. et al. "Structure of the gluABCD cluster encoding the glutamate uptake system of Corynebacterium glutamicum," <i>J. Bacteriol.</i> , 177(5):1152-1158 (1995)
X81379	dapE	Succinyl/diaminopimelate desuccinylase	Wehrmann, A. et al. "Analysis of different DNA fragments of Corynebacterium glutamicum complementing dapE of Escherichia coli," <i>Microbiology</i> , 40:3349-56 (1994)

Table 2 (continued)

X82061	16S rDNA	16S ribosomal RNA	Ruimy, R. et al. "Phylogeny of the genus <i>Corynebacterium</i> deduced from analyses of small-subunit ribosomal DNA sequences," <i>Int. J. Syst. Bacteriol.</i> , 45(4):740-746 (1995)
X82928	asd; lysC	Aspartate-semialdehyde dehydrogenase; ?	Serebrjiski, I. et al. "Multicopy suppression by asd gene and osmotic stress-dependent complementation by heterologous proA in proA mutants," <i>J. Bacteriol.</i> , 177(24):7255-7260 (1995)
X82929	proA	Gamma-glutamyl phosphate reductase	Serebrjiski, I. et al. "Multicopy suppression by asd gene and osmotic stress-dependent complementation by heterologous proA in proA mutants," <i>J. Bacteriol.</i> , 177(24):7255-7260 (1995)
X84237	16S rDNA	16S ribosomal RNA	Pascual, C. et al. "Phylogenetic analysis of the genus <i>Corynebacterium</i> based on 16S rRNA gene sequences," <i>Int. J. Syst. Bacteriol.</i> , 45(4):724-728 (1995)
X85965	aroF, dapE	Aromatic amino acid permease; ?	Wehrmann et al. "Functional analysis of sequences adjacent to dapE of <i>C. glutamicum</i> proline reveals the presence of aroP, which encodes the aromatic amino acid transporter," <i>J. Bacteriol.</i> , 177(20):5991-5993 (1995)
X86157	argB; argC; argJ; argF; argI	Acetylglutamate kinase; N-acetyl-gamma-glutamyl-phosphate reductase; acetylornithine aminotransferase; ornithine carbamoyltransferase; glutamate N-acetyltransferase	Sakanyan, V. et al. "Genes and enzymes of the acetyl cycle of arginine biosynthesis in <i>Corynebacterium glutamicum</i> : enzyme evolution in the early steps of the arginine pathway," <i>Microbiology</i> , 142:99-108 (1996)
X89084	pia; ackA	Phosphate acetyltransferase; acetate kinase	Reinscheid, D.J. et al. "Cloning, sequence analysis, expression and inactivation of the <i>Corynebacterium glutamicum</i> pia-ack operon encoding phosphotransacetylase and acetate kinase," <i>Microbiology</i> , 145:503-513 (1999)
X89850	attB	Attachment site	Le Marrec, C. et al. "Genetic characterization of site-specific integration functions of phi AAU2 infecting 'Arthrobacter aureus C70,'" <i>J. Bacteriol.</i> , 178(7):1996-2004 (1996)
X90356		Promoter fragment F1	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90357		Promoter fragment F2	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90358		Promoter fragment F10	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90359		Promoter fragment F13	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)

Table 2 (continued)

X90360	Promoter fragment F22	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90361	Promoter fragment F34	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90362	Promoter fragment F37	Patek, M. et al. "Promoters from <i>C. glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90363	Promoter fragment F45	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90364	Promoter fragment F64	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90365	Promoter fragment F75	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90366	Promoter fragment PF101	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90367	Promoter fragment PF104	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X90368	Promoter fragment PF109	Patek, M. et al. "Promoters from <i>Corynebacterium glutamicum</i> : cloning, molecular analysis and search for a consensus motif," <i>Microbiology</i> , 142:1297-1309 (1996)
X93513	amt	Siewe, R.M. et al. "Functional and genetic characterization of the (methyl) ammonium uptake carrier of <i>Corynebacterium glutamicum</i> ," <i>J. Biol. Chem.</i> , 271(10):5398-5403 (1996)
X93514	betP	Peter, H. et al. "Isolation, characterization, and expression of the <i>Corynebacterium glutamicum</i> betP gene, encoding the transport system for the compatible solute glycine betaine," <i>J. Bacteriol.</i> , 178(17):5229-5234 (1996)
X95649	orf4	Patek, M. et al. "Identification and transcriptional analysis of the dapB-ORF2-dapA-ORF4 operon of <i>Corynebacterium glutamicum</i> , encoding two enzymes involved in L-lysine synthesis," <i>Biotechnol. Lett.</i> , 19:1113-1117 (1997)
X96471	lysE, lysG	Vrijic, M. et al. "A new type of transporter with a new type of cellular function: L-lysine export from <i>Corynebacterium glutamicum</i> ," <i>Mol. Microbiol.</i> , 22(5):815-826 (1996)

Table 2 (continued)

X96580	panB; panC; xylB	3-methyl-2-oxobutanoate hydroxymethyltransferase; pantoate-beta-alanine ligase; xylulokinase	Sahn, H. et al. "D-pantothenate synthesis in <i>Corynebacterium glutamicum</i> and use of panBC and genes encoding L-valine synthesis for D-pantothenate overproduction," <i>Appl. Environ. Microbiol.</i> , 65(5):1973-1979 (1999)
X96962		Insertion sequence IS1207 and transposase	Ramos, A. et al. "Cloning, sequencing and expression of the gene encoding elongation factor P in the amino-acid producer <i>Brevibacterium lactofermentum</i> (<i>Corynebacterium glutamicum</i> ATCC 13869)," <i>Gene</i> , 198:217-222 (1997)
X99289		Elongation factor P	
Y00140	thrB	Homoserine kinase	Mateos, L.M. et al. "Nucleotide sequence of the homoserine kinase (thrB) gene of the <i>Brevibacterium lactofermentum</i> ," <i>Nucleic Acids Res.</i> , 15(9):3922 (1987)
Y00151	ddh	Meso-diaminopimelate D-dehydrogenase (EC 1.4.1.16)	Ishino, S. et al. "Nucleotide sequence of the meso-diaminopimelate D-dehydrogenase gene from <i>Corynebacterium glutamicum</i> ," <i>Nucleic Acids Res.</i> , 15(9):3917 (1987)
Y00476	thrA	Homoserine dehydrogenase	Mateos, L.M. et al. "Nucleotide sequence of the homoserine dehydrogenase (thrA) gene of the <i>Brevibacterium lactofermentum</i> ," <i>Nucleic Acids Res.</i> , 15(24):10598 (1987)
Y00546	hom; thrB	Homoserine dehydrogenase; homoserine kinase	Peoples, O.P. et al. "Nucleotide sequence and fine structural analysis of the <i>Corynebacterium glutamicum</i> hom-thrB operon," <i>Mol. Microbiol.</i> , 2(1):63-72 (1988)
Y08964	murC; fsq/divD; fsz	UPD-N-acetylmuramate-alanine ligase; division initiation protein or cell division protein; cell division protein	Honrath, M.P. et al. "Identification, characterization, and chromosomal organization of the fsz gene from <i>Brevibacterium lactofermentum</i> ," <i>Mol. Gen. Genet.</i> , 259(1):97-104 (1998)
Y09163	putP	High affinity proline transport system	Peter, H. et al. "Isolation of the putP gene of <i>Corynebacterium glutamicum</i> and characterization of a low-affinity uptake system for compatible solutes," <i>Arch. Microbiol.</i> , 168(2):143-151 (1997)
Y09548	pyc	Pyruvate carboxylase	Peters-Wendisch, P.G. et al. "Pyruvate carboxylase from <i>Corynebacterium glutamicum</i> : characterization, expression and inactivation of the pyc gene," <i>Microbiology</i> , 144:915-927 (1998)
Y09578	leuB	3-isopropylmalate dehydrogenase	Pajek, M. et al. "Analysis of the leuB gene from <i>Corynebacterium glutamicum</i> ," <i>Appl. Microbiol. Biotechnol.</i> , 50(1):42-47 (1998)
Y12472		Attachment site bacteriophage Phi-16	Moreau, S. et al. "Site-specific integration of coryneophage Phi-16: The construction of an integration vector," <i>Microbiol.</i> , 145:539-548 (1999)
Y12537	proP	Proline/ectoine uptake system protein	Peter, H. et al. " <i>Corynebacterium glutamicum</i> is equipped with four secondary carriers for compatible solutes: Identification, sequencing, and characterization of the proline/ectoine uptake system, ProP, and the ectoine/proline/glycine betaine carrier, EcP," <i>J. Bacteriol.</i> , 180(22):6005-6012 (1998)

Table 2 (continued)

Y13221	glnA	Glutamine synthetase I	Jakoby, M. et al. "Isolation of <i>Corynebacterium glutamicum</i> glnA gene encoding glutamine synthetase I," <i>FEMS Microbiol. Lett.</i> , 154(1):81-88 (1997)
Y16642	lpd	Dihydrolipoamide dehydrogenase	
Y18059		Attachment site Corynephage 304L	Moreau, S. et al. "Analysis of the integration functions of φ304L: An integrase module among corynephages," <i>Virology</i> , 255(1):150-159 (1999)
Z21501	argS; lysA	Arginyl-tRNA synthetase; diaminiopimelate decarboxylase (partial)	Oguiza, J.A. et al. "A gene encoding arginyl-tRNA synthetase is located in the upstream region of the lysA gene in <i>Brevibacterium lactofermentum</i> : Regulation of argS-lysA cluster expression by arginine," <i>J. Bacteriol.</i> , 175(22):7356-7362 (1993)
Z21502	dapA; dapB	Dihydrodipicolinate synthase; dihydrodipicolinate reductase	Pisabarro, A. et al. "A cluster of three genes (dapA, orf2, and dapB) of <i>Brevibacterium lactofermentum</i> encodes dihydrodipicolinate reductase, and a third polypeptide of unknown function," <i>J. Bacteriol.</i> , 175(9):2743-2749 (1993)
Z29563	thrC	Threonine synthase	Malumbres, M. et al. "Analysis and expression of the thrC gene of the encoded threonine synthase," <i>Appl. Environ. Microbiol.</i> , 60(7):2209-2219 (1994)
Z46753	16S rDNA	Gene for 16S ribosomal RNA	
Z49822	sigA	SigA sigma factor	Oguiza, J.A. et al. "Multiple sigma factor genes in <i>Brevibacterium lactofermentum</i> : Characterization of sigA and sigB," <i>J. Bacteriol.</i> , 178(2):550-553 (1996)
Z49823	galE; dtxR	Catalytic activity UDP-galactose 4-epimerase; diphtheria toxin regulatory protein	Oguiza, J.A. et al. "The galE gene encoding the UDP-galactose 4-epimerase of <i>Brevibacterium lactofermentum</i> is coupled transcriptionally to the dmdR gene," <i>Gene</i> , 177:103-107 (1996)
Z49824	orf1; sigB	?; SigB sigma factor	Oguiza, J.A. et al. "Multiple sigma factor genes in <i>Brevibacterium lactofermentum</i> : Characterization of sigA and sigB," <i>J. Bacteriol.</i> , 178(2):550-553 (1996)
Z66534		Transposase	Correia, A. et al. "Cloning and characterization of an IS-like element present in the genome of <i>Brevibacterium lactofermentum</i> ATCC 13869," <i>Gene</i> , 170(1):91-94 (1996)

* A sequence for this gene was published in the indicated reference. However, the sequence obtained by the inventors of the present application is significantly longer than the published version. It is believed that the published version relied on an incorrect start codon, and thus represents only a fragment of the actual coding region.

TABLE 3: *Corynebacterium* and *Brevibacterium* Strains Which May be Used in the Practice of the Invention

Genus	species	ATCC	FERM	NRRL	CECT	NCIMB	CBS	NCTC	DSMZ
<i>Brevibacterium</i>	<i>ammoniagenes</i>	21054							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19350							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19351							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19352							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19353							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19354							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19355							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	19356							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	21055							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	21077							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	21553							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	21580							
<i>Brevibacterium</i>	<i>ammoniagenes</i>	39101							
<i>Brevibacterium</i>	<i>butanicum</i>	21196							
<i>Brevibacterium</i>	<i>divaricatum</i>	21792	P928						
<i>Brevibacterium</i>	<i>flavum</i>	21474							
<i>Brevibacterium</i>	<i>flavum</i>	21129							
<i>Brevibacterium</i>	<i>flavum</i>	21518							
<i>Brevibacterium</i>	<i>flavum</i>			B11474					
<i>Brevibacterium</i>	<i>flavum</i>			B11472					
<i>Brevibacterium</i>	<i>flavum</i>	21127							
<i>Brevibacterium</i>	<i>flavum</i>	21128							
<i>Brevibacterium</i>	<i>flavum</i>	21427							
<i>Brevibacterium</i>	<i>flavum</i>	21475							
<i>Brevibacterium</i>	<i>flavum</i>	21517							
<i>Brevibacterium</i>	<i>flavum</i>	21528							
<i>Brevibacterium</i>	<i>flavum</i>	21529							
<i>Brevibacterium</i>	<i>flavum</i>			B11477					
<i>Brevibacterium</i>	<i>flavum</i>			B11478					
<i>Brevibacterium</i>	<i>flavum</i>	21127							
<i>Brevibacterium</i>	<i>flavum</i>			B11474					
<i>Brevibacterium</i>	<i>healii</i>	15527							
<i>Brevibacterium</i>	<i>ketoglutamicum</i>	21004							
<i>Brevibacterium</i>	<i>ketoglutamicum</i>	21089							
<i>Brevibacterium</i>	<i>ketosoreductum</i>	21914							
<i>Brevibacterium</i>	<i>lactofermentum</i>				70				
<i>Brevibacterium</i>	<i>lactofermentum</i>				74				
<i>Brevibacterium</i>	<i>lactofermentum</i>				77				
<i>Brevibacterium</i>	<i>lactofermentum</i>	21798							
<i>Brevibacterium</i>	<i>lactofermentum</i>	21799							
<i>Brevibacterium</i>	<i>lactofermentum</i>	21800							
<i>Brevibacterium</i>	<i>lactofermentum</i>	21801							
<i>Brevibacterium</i>	<i>lactofermentum</i>			B11470					
<i>Brevibacterium</i>	<i>lactofermentum</i>			B11471					

Genus	species	ATCC	DSMZ	NRRL	CECT	NCIMB	CBS	NCTC	DSMZ
Brevibacterium	lactofermentum	21086							
Brevibacterium	lactofermentum	21420							
Brevibacterium	lactofermentum	21086							
Brevibacterium	lactofermentum	31269							
Brevibacterium	linens	9174							
Brevibacterium	linens	19391							
Brevibacterium	linens	8377							
Brevibacterium	paraffinolyticum					11160			
Brevibacterium	spec.						717.73		
Brevibacterium	spec.						717.73		
Brevibacterium	spec.	14604							
Brevibacterium	spec.	21860							
Brevibacterium	spec.	21864							
Brevibacterium	spec.	21865							
Brevibacterium	spec.	21866							
Brevibacterium	spec.	19240							
Corynebacterium	acetoacidophilum	21476							
Corynebacterium	acetoacidophilum	13870							
Corynebacterium	acetoglutamicum			B11473					
Corynebacterium	acetoglutamicum			B11475					
Corynebacterium	acetoglutamicum	15806							
Corynebacterium	acetoglutamicum	21491							
Corynebacterium	acetoglutamicum	31270							
Corynebacterium	acetophilum			B3671					
Corynebacterium	ammoniogenes	6872						2399	
Corynebacterium	ammoniogenes	15511							
Corynebacterium	fujikense	21496							
Corynebacterium	glutamicum	14067							
Corynebacterium	glutamicum	39137							
Corynebacterium	glutamicum	21254							
Corynebacterium	glutamicum	21255							
Corynebacterium	glutamicum	31830							
Corynebacterium	glutamicum	13032							
Corynebacterium	glutamicum	14305							
Corynebacterium	glutamicum	15455							
Corynebacterium	glutamicum	13058							
Corynebacterium	glutamicum	13059							
Corynebacterium	glutamicum	13060							
Corynebacterium	glutamicum	21492							
Corynebacterium	glutamicum	21513							
Corynebacterium	glutamicum	21526							
Corynebacterium	glutamicum	21543							
Corynebacterium	glutamicum	13287							
Corynebacterium	glutamicum	21851							
Corynebacterium	glutamicum	21253							
Corynebacterium	glutamicum	21514							
Corynebacterium	glutamicum	21516							
Corynebacterium	glutamicum	21299							

Genus	species	ATCC	FERM	NRRL	CECT	NCIMB	GBS	NCTC	DSMZ
Corynebacterium	glutamicum	21300							
Corynebacterium	glutamicum	39684							
Corynebacterium	glutamicum	21488							
Corynebacterium	glutamicum	21649							
Corynebacterium	glutamicum	21650							
Corynebacterium	glutamicum	19223							
Corynebacterium	glutamicum	13869							
Corynebacterium	glutamicum	21157							
Corynebacterium	glutamicum	21158							
Corynebacterium	glutamicum	21159							
Corynebacterium	glutamicum	21355							
Corynebacterium	glutamicum	31808							
Corynebacterium	glutamicum	21674							
Corynebacterium	glutamicum	21562							
Corynebacterium	glutamicum	21563							
Corynebacterium	glutamicum	21564							
Corynebacterium	glutamicum	21565							
Corynebacterium	glutamicum	21566							
Corynebacterium	glutamicum	21567							
Corynebacterium	glutamicum	21568							
Corynebacterium	glutamicum	21569							
Corynebacterium	glutamicum	21570							
Corynebacterium	glutamicum	21571							
Corynebacterium	glutamicum	21572							
Corynebacterium	glutamicum	21573							
Corynebacterium	glutamicum	21579							
Corynebacterium	glutamicum	19049							
Corynebacterium	glutamicum	19050							
Corynebacterium	glutamicum	19051							
Corynebacterium	glutamicum	19052							
Corynebacterium	glutamicum	19053							
Corynebacterium	glutamicum	19054							
Corynebacterium	glutamicum	19055							
Corynebacterium	glutamicum	19056							
Corynebacterium	glutamicum	19057							
Corynebacterium	glutamicum	19058							
Corynebacterium	glutamicum	19059							
Corynebacterium	glutamicum	19060							
Corynebacterium	glutamicum	19185							
Corynebacterium	glutamicum	13286							
Corynebacterium	glutamicum	21515							
Corynebacterium	glutamicum	21527							
Corynebacterium	glutamicum	21544							
Corynebacterium	glutamicum	21492							
Corynebacterium	glutamicum			B8183					
Corynebacterium	glutamicum			B8182					
Corynebacterium	glutamicum			B12416					
Corynebacterium	glutamicum			B12417					

Genus	species	ATCC	FERM	NRRL	CECT	NCIMB	CBS	NCTC	DSMZ
Corynebacterium	glutamicum			B12418					
Corynebacterium	glutamicum			B11476					
Corynebacterium	glutamicum	21608							
Corynebacterium	lilium		P973						
Corynebacterium	nitrilophilus	21419				11594			
Corynebacterium	spec.		P4445						
Corynebacterium	spec.		P4446						
Corynebacterium	spec.	31088							
Corynebacterium	spec.	31089							
Corynebacterium	spec.	31090							
Corynebacterium	spec.	31090							
Corynebacterium	spec.	31090							
Corynebacterium	spec.	15954							20145
Corynebacterium	spec.	21857							
Corynebacterium	spec.	21862							
Corynebacterium	spec.	21863							

ATCC: American Type Culture Collection, Rockville, MD, USA

FERM: Fermentation Research Institute, Chiba, Japan

NRRL: ARS Culture Collection, Northern Regional Research Laboratory, Peoria, IL, USA

CECT: Coleccion Espanola de Cultivos Tipo, Valencia, Spain

NCIMB: National Collection of Industrial and Marine Bacteria Ltd., Aberdeen, UK

CBS: Centraalbureau voor Schimmelcultures, Baarn, NL

NCTC: National Collection of Type Cultures, London, UK

DSMZ: Deutsche Sammlung von Mikroorganismen und Zellkulturen, Braunschweig, Germany

For reference see Sugawara, H. et al. (1993) World directory of collections of cultures of microorganisms: Bacteria, fungi and yeasts (4th edn), World federation for culture collections world data center on microorganisms, Saimata, Japan.

Table 4: Alignment Results

ID #	length (nt)	Genbank Hit	Length	Accession	Name of Genbank Hit	Source of Genbank Hit	% homology (GAP)	Date of Deposit
na00026	1508	GB_RCMMHC310M6	158405	AF109906	Mus musculus MHC class III region RD gene, partial cds; B1, C2, G9A, NG22, G8, HSP70, HSP70, HSC70, and smRNP genes, complete cds; G7A gene, partial cds; and unknown genes.	Mus musculus	38,003	10-DEC-1998
		GB_HTG2:AC007029	119007	AC007029	Homo sapiens clone DJ0855F16, *** SEQUENCING IN PROGRESS	Homo sapiens	37,943	7-Apr-99
		GB_HTG2:AC007029	119007	AC007029	Homo sapiens clone DJ0855F16, *** SEQUENCING IN PROGRESS	Homo sapiens	37,943	7-Apr-99
na00072					***, 1 unordered pieces.			
					***, 1 unordered pieces.			
na00111	1116	GB_BA1:SAUSIG8	2748	M94370	Stigmatella aurantiaca Sigma factor (sigA) gene, complete cds.	Stigmatella aurantiaca	40,435	16-Aug-94
		GB_BA1:SC588	28500	AL022374	Streptomyces coelicolor cosmid 588.	Streptomyces coelicolor	40,090	22-Apr-98
		GB_BA2:AE001767	9086	AE001767	Thermotoga maritima section 79 of 136 of the complete genome.	Thermotoga maritima	35,091	2-Jun-99
na00112	1314	GB_EST35:AU075536	418	AU075536	ALU075536 Rice shoot Oryza sativa cDNA clone S0028_2Z, mRNA sequence.	Oryza sativa	39,423	7-Jul-99
		GB_GSS9:AO157585	847	AO157585	nbx00009B16r CUGI Rice BAC Library Oryza sativa genomic clone nbx00009B16r, genomic survey sequence.	Oryza sativa	40,867	12-Sep-98
		GB_GSS14:AO510314	542	AO510314	nbx00095C05f CUGI Rice BAC Library Oryza sativa genomic clone nbx00095C05f, genomic survey sequence.	Oryza sativa	39,372	04-MAY-1999
na00133	936	GB_BA1:SC2G5	38404	AL035478	Streptomyces coelicolor cosmid 2G5.	Streptomyces coelicolor	41,170	11-Jun-99
		GB_EST7:W64291	515	W64291	md98h12.r1 Soares mouse embryo NbME13.5 14.5 Mus musculus cDNA clone IMAGE:386087 5' similar to gbl26528 Mus musculus Rab11b mRNA, complete cds (MOUSE); mRNA sequence.	Mus musculus	35,306	10-Jun-96
		GB_PR3:AC005624	39594	AC005624	Homo sapiens chromosome 19, cosmid R30017, complete sequence.	Homo sapiens	39,054	6-Sep-98
na00137	1212	GB_BA2:AF124600	4115	AF124600	Corynebacterium glutamicum chorismate synthase (aroC), shikimate kinase (aroK), and 3-dehydroquinate synthase (aroB) genes, complete cds; and putative cytoplasmic peptidase (pepQ) gene, partial cds.	Corynebacterium glutamicum	99,867	04-MAY-1999
		GB_BA1:MTCY159	33818	Z63863	Mycobacterium tuberculosis H37Rv complete genome; segment 111/162.	Mycobacterium tuberculosis	40,959	17-Jun-98
		GB_BA1:MT3DEHQ	3437	X59509	M.tuberculosis, genes for 3-dehydroquinate synthase and 3-dehydroquinase.	Mycobacterium tuberculosis	52,583	30-Jun-93
na00139	834	GB_BA1:BLELONP	738	X59289	B.lactofermentum gene encoding elongation factor P.	Corynebacterium glutamicum	100,000	1-Nov-97
		GB_PL1:SPAC24C9	38596	Z98601	S.pombe chromosome I cosmid c24C9.	Schizosaccharomyces pombe	35,230	24-Feb-99
		GB_HTG1:CEY102A5_1110000	739711	Z39711	Caenorhabditis elegans chromosome V clone Y102A5, *** SEQUENCING IN PROGRESS ***; in unordered pieces.	Caenorhabditis elegans	37,775	Z39711

Table 4 (continued)

rx00152	1419	GB_BA1:MTCV277	38300	Z79701	Myco bacterium tuberculosis H37Rv complete genome; segment 65/162.	Myco bacterium tuberculosis	58,500	17-Jun-98
		GB_BA1:MSGY456	37316	AD000001	Myco bacterium tuberculosis sequence from clone y456.	Myco bacterium tuberculosis	38,913	03-DEC-1996
		GB_BA2:AF002133	15437	AF002133	Myco bacterium avium strain G1R10 transcriptional regulator (mav81) gene, partial cds, aconitase (act), invasin 1 (inv1), invasin 2 (inv2), transcriptional regulator (moxR), ketoacyl-reductase (fabG), enoyl-reductase (inhA) and ferrochelatase (mav272) genes, complete cds.	Myco bacterium avium	64,009	26-MAR-1998
rx000226	948	GB_PR3:AC005755	43299	AC005755	Homo sapiens chromosome 19, fosmid 35347, complete sequence.	Homo sapiens	36,209	02-OCT-1998
		GB_GSS5:AQ818463	413	AQ818463	HS_5250_A2_B08_SP6E RPCH-11 Human Male BAC Library Homo sapiens genomic clone Plate=826 Col=16 Row=C, genomic survey sequence.	Homo sapiens	37,288	26-Aug-99
		GB_GSS5:AQ782337	832	AQ782337	HS_3184_B1_H12_T7C CIT Approved Human Genomic Sperm Library D Homo sapiens genomic clone Plate=3184 Col=23 Row=P, genomic survey sequence.	Homo sapiens	35,917	2-Aug-99
rx000249	980	GB_BA2:AF035608	3614	AF035608	Pseudomonas aeruginosa ATP sulfurylase small subunit (cysD) and ATP sulfurylase GTP-binding subunit/APS kinase (cysN) genes, complete cds.	Pseudomonas aeruginosa	50,205	1-Jun-98
		GB_BA1:AB017641	17101	AB017641	Micromonospora griseorubida gene for polyketide synthase, complete cds.	Micromonospora griseorubida	40,266	2-Apr-99
		GB_BA2:AF002133	15437	AF002133	Myco bacterium avium strain G1R10 transcriptional regulator (mav81) gene, partial cds, aconitase (act), invasin 1 (inv1), invasin 2 (inv2), transcriptional regulator (moxR), ketoacyl-reductase (fabG), enoyl-reductase (inhA) and ferrochelatase (mav272) genes, complete cds.	Myco bacterium avium	38,429	26-MAR-1998
rx000299	1101	GB_BA2:CORCSLYS	2821	M89931	Corynebacterium glutamicum beta C-S lyase (aecD) and branched-chain amino acid uptake carrier (brnQ) genes, complete cds, and hypothetical protein YhbW (yhbW) gene, partial cds.	Corynebacterium glutamicum	100,000	4-Jun-98
		GB_BA1:CGECTP	2719	AJ001436	Corynebacterium glutamicum ecp gene.	Corynebacterium glutamicum	41,143	20-Nov-98
		GB_BA2:AF181035	5922	AF181035	Rhodobacter sphaeroides glycolysis utilization operon, complete sequence.	Rhodobacter sphaeroides	36,701	7-Sep-99
rx000332	825	GB_BA1:CGTHRC	3120	X56037	Corynebacterium glutamicum thrC gene for threonine synthase (EC 4.2.99.2).	Corynebacterium glutamicum	37,730	17-Jun-97
		GB_PAT:109078	3146	109078	Sequence 4 from Patent WO 8809819.	Unknown.	38,700	02-DEC-1994
		GB_PR3:HS1333B15	73666	AL109954	Human DNA sequence from clone 333B15 on chromosome 20, complete sequence.	Homo sapiens	37,203	23-Nov-99
rx000470	1392	GB_PL2:DCPCNAM	865	X62977	D.carota mRNA for proliferating cell nuclear antigen (PCNA).	Daucus carota	37,914	30-Sep-99

Table 4 (continued)

GB_PL2:AC006267	101644	AC006267	Arabidopsis thaliana BAC F5M13 from chromosome IV near 21.5 cM, Arabidopsis thaliana complete sequence.	36,158	27-Apr-99
GB_BA1:TT10SARNA	721	Y15063	Thermus thermophilus 10Sa RNA gene.	39,494	18-Aug-98
GB_BA1:SERERYAA	11219	M63676	S erythraea first ORF of eryA gene, complete cds.	38,781	26-Apr-93
GB_PAT:AR049367	11219	AR049367	Sequence 1 from patient US 6524513.	38,781	29-Sep-99
GB_BA1:SERERYAA	11219	M63676	S erythraea first ORF of eryA gene, complete cds.	38,205	26-Apr-93
GB_PR4:AC007206	42732	AC007206	Homo sapiens chromosome 19, cosmid R27370, complete sequence. Homo sapiens erythraea	34,982	4-Apr-99
GB_EST26:AI344735	462	AI344735	qp05a10x1 NCL_CGAP_Kid5 Homo sapiens cDNA clone IMAGE:1917114 3' similar to gb:M13600 1-LYMPHOCYTE MATURATION-ASSOCIATED PROTEIN (HUMAN), mRNA sequence.	42,675	2-Feb-99
GB_PR4:AC006479	161837	AC006479	Homo sapiens clone DJ105J04, complete sequence.	38,462	11-Nov-99
GB_PR4:AC006111	190825	AC006111	Homo sapiens chromosome 16 clone RPC1-11_461A9, complete sequence.	40,736	3-Jul-99
GB_HTG2:AF128834	196589	AF128834	Homo sapiens chromosome 8 clone BAC 57G24 map 8p12, *** SEQUENCING IN PROGRESS ***, in unordered pieces.	34,062	28-Feb-99
GB_HTG2:AF128834	196589	AF128834	Homo sapiens chromosome 8 clone BAC 57G24 map 8p12, *** SEQUENCING IN PROGRESS ***, in unordered pieces.	34,062	28-Feb-99
GB_BA1:D86429	5925	D86429	Saccharopolyspora rectivirgula gene for beta-galactosidase, complete cds.	53,871	09-DEC-1998
GB_HTG1:HS1099D15	1301	AL035456	Homo sapiens chromosome 20 clone RP5-1099D15, *** SEQUENCING IN PROGRESS ***, in unordered pieces.	33,546	23-Nov-99
GB_HTG1:HS1099D15	1301	AL035456	Homo sapiens chromosome 20 clone RP5-1099D15, *** SEQUENCING IN PROGRESS ***, in unordered pieces.	33,546	23-Nov-99
GB_BA2:U00015	42325	U00015	Mycobacterium leprae cosmid B1620.	34,783	01-MAR-1994
GB_BA1:U00020	36947	U00020	Mycobacterium leprae cosmid B229.	34,900	01-MAR-1994
GB_HTG1:HS179115	210672	Z84464	Homo sapiens chromosome 13 clone 179115, *** SEQUENCING IN PROGRESS ***, in unordered pieces.	32,898	22-Jan-97
GB_BA1:MTV008	63033	AL021246	Mycobacterium tuberculosis H37Rv complete genome, segment 108/162.	37,011	17-Jun-98
GB_BA2:AF071885	2188	AF071885	Streptomyces coelicolor ATP-dependent Clp protease proteolytic subunit 1 (clpP1) and ATP-dependent Clp protease proteolytic subunit 2 (clpP2) genes, complete cds; and ATP-dependent Clp protease ATP-binding subunit ClpX (clpX) gene, partial cds.	62,963	29-Jun-99
GB_BA2:AF013216	15742	AF013216	Myxococcus xanthus Dog (dog), isocitrate lyase (icl), Mle (mle), Ufo (ufo), fumarate hydratase (fhy), and proteasome major subunit (clpP) genes, complete cds; and acyl-CoA oxidase (aco) gene, partial cds.	54,683	28-Jan-98

Table 4 (continued)

xa00587	714	GB_BA1:MTV008	63033	AL021246	Mycobacterium tuberculosis H37Rv complete genome; segment 108/162.	Mycobacterium tuberculosis	42,090	17-Jun-98
		GB_BA1:CGBPH16	962	Y12472	C glutamicum DNA, attachment site bacteriophage Phi-16.	Corynebacterium glutamicum	40,000	05-MAR-1999
		GB_BA1:ECOC1PPA	1236	J05534	Escherichia coli ATP-dependent ctp protease proteolytic component (glpP) gene, complete cds.	Escherichia coli	52,119	26-Apr-93
xa00621	906	GB_EST1:D36491	360	D36491	CELK03GYF Yuji Kohara unpublished cDNA Caenorhabditis elegans cDNA clone yk3g11.5; mRNA sequence.	Caenorhabditis elegans	40,390	8-Aug-94
		GB_IN2:CELCT6A3	34968	U41534	Caenorhabditis elegans cosmid C16A3.	Caenorhabditis elegans	35,477	19-MAY-1999
		GB_HTG3:AC009311	160198	AC009311	Homo sapiens clone NH0311L03. *** SEQUENCING IN PROGRESS	Homo sapiens	38,636	13-Aug-99
					***, 3 unordered pieces.			
xa00622	1539	GB_BA1:AB004795	3039	AB004795	Pseudomonas sp. gene for dipeptidyl aminopeptidase, complete cds.	Pseudomonas sp.	54,721	5-Feb-99
		GB_BA1:MBOP11	2392	D38405	Moraxella lacunata gene for protease II, complete cds.	Moraxella lacunata	50,167	8-Feb-99
		GB_IN2:AF078916	2960	AF078916	Trypanosoma brucei oligopeptidase B (opb) gene, complete cds.	Trypanosoma brucei	48,076	08-OCT-1999
xa00650	759	GB_BA2:AF161327	2021	AF161327	Corynebacterium diphtheriae histidine kinase CnrS (chrS) and response regulator ChrA (chrA) genes, complete cds.	Corynebacterium diphtheriae	51,319	9-Sep-99
		GB_PL2:ATAC006533	99188	AC006533	Arabidopsis thaliana chromosome II BAC F20M17 genomic sequence, complete sequence.	Arabidopsis thaliana	38,051	26-MAY-1999
		GB_PL2:ATAC006533	99188	AC006533	Arabidopsis thaliana chromosome II BAC F20M17 genomic sequence, complete sequence.	Arabidopsis thaliana	35,403	26-MAY-1999
xa00675	915	GB_BA1:SC3C8	33095	AL023861	Streptomyces coelicolor cosmid 3C8.	Streptomyces coelicolor	36,836	15-Jan-99
		GB_PR3:AC005736	215441	AC005736	Homo sapiens chromosome 16, BAC clone 462G18 (LANL), complete sequence.	Homo sapiens	42,027	01-OCT-1998
		GB_IN2:AC005719	188357	AC005719	Drosophila melanogaster, chromosome 2L, region 38A5-38B4, BAC clone BACR48M05, complete sequence.	Drosophila melanogaster	35,531	27-OCT-1999
xa00689	1614	GB_PAT1:E07294	2975	E07294	genomic DNA encoding dehydrogenase of Bacillus stearothermophilus.	Bacillus stearothermophilus	45,677	29-Sep-97
		GB_BA1:BACALDHT	1975	D13846	B. stearothermophilus aldH T gene for aldehyde dehydrogenase, complete cds.	Bacillus stearothermophilus	45,677	20-Feb-99
		GB_BA2:PPU6338	5276	U96338	Pseudomonas putida NCIMB 9866 plasmid pRA4000 p-cresol degradative pathway genes, p-hydroxybenzaldehyde dehydrogenase (pohA), p-cresol methylhydroxylase, cytochrome subunit precursor (pohC), unknown (pohX) and p-cresol methylhydroxylase, flavoprotein subunit (pohF) genes, complete cds.	Pseudomonas putida	44,317	13-MAY-1999
xa00715	918	GB_EST30:AI647104	218	AI647104	vn15cd1 y1 Stragene mouse heart (#937316) Mus musculus cDNA clone IMAGE:1021248 5', mRNA sequence.	Mus musculus	58,511	29-Apr-99
		GB_EST17:AA638159	447	AA638159	vn15cd1 r1 Stragene mouse heart (#937316) Mus musculus cDNA clone IMAGE:1021248 5', mRNA sequence.	Mus musculus	41,195	22-OCT-1997

Table 4 (continued)

GB_EST10:AA184468	583	AA184468	mt52h05.r1 Stratagene mouse embryonic carcinoma (#9373717) Mus musculus cDNA clone IMAGE:533561 5' similar to gb:D10918 Mouse mRNA for ubiquitin like protein, partial sequence (MOUSE);, mRNA sequence.	40,426	12-Feb-97
GB_HTG3:AC009855	167592	AC009855	Homo sapiens clone 1_C_5, *** SEQUENCING IN PROGRESS ***	36,673	3-Sep-99
GB_HTG3:AC009855	167592	AC009855	13 unordered pieces.	36,673	3-Sep-99
GB_HTG3:AC009855	167592	AC009855	Homo sapiens clone 1_C_5, *** SEQUENCING IN PROGRESS ***	36,673	3-Sep-99
GB_PRA4:AC005082	166739	AC005082	13 unordered pieces.	39,557	8-Sep-99
GB_BAT1:MLCB596	38426	AL035472	Homo sapiens clone RG271G13, complete sequence.	54,562	27-Aug-99
GB_GSS12:AC368028	652	AC368028	Mycobacterium leprae cosmid B596.	42,657	5-Feb-99
GB_HTG3:AC008067	151242	AC008067	tox0001N11r CUGI Tomato BAC Library Lycopersicon esculentum genomic clone tox0001N11r; genomic survey sequence.	37,239	8-Sep-99
GB_BAT1:MLU15182	40123	U15182	Homo sapiens clone NH0303104, *** SEQUENCING IN PROGRESS ***	36,616	09-MAR-1995
GB_BAT1:MSGL6110S	37769	L78622	2 unordered pieces.	35,714	15-Jun-96
GB_GSS14:AC578181	728	AC578181	Mycobacterium leprae cosmid B2265.	39,246	2-Jun-99
GB_GSS5:ACQ769737	519	AQ769737	Mycobacterium leprae cosmid L611 DNA sequence.	37,765	28-Jul-99
GB_BAT1:RTU08434	2400	U08434	nbxb0083P08r; genomic survey sequence.	40,700	16-Apr-97
GB_EST31:F33810	243	F33810	HS_3160_A2_G04_T7C CIT Approved Human Genomic Spem Library D Homo sapiens genomic clone Plate=3160 Col=8 Row=M; genomic survey sequence.	41,564	13-MAY-1999
GB_PRA4:AC005868	96180	AC005868	Rhizobium trifolii orotate phosphoribosyltransferase (pyrE) and fructokinase (frk) genes, complete cds.	32,298	27-Feb-99
GB_EST8:A4000903	396	A4000903	HSPD27491 HM3 Homo sapiens cDNA clone s3000041E12, mRNA sequence.	42,045	18-Jul-96
GB_EST25:A1317789	696	A1317789	Homo sapiens 12q24.2 PAC RPC15-944M2 (Roswell Park Cancer Institute Human PAC Library) complete sequence.	38,557	17-DEC-1998
GB_PH1:BPHE589	41489	AJ006589	mq38b04.r1 Soares mouse embryo NkME13.5 14.5 Mus musculus cDNA clone IMAGE:426031 5', mRNA sequence.	41,806	29-Apr-99
GB_HTG3:AC006887	215801	AC006887	uj2009.y1 Sugano mouse embryo mewa Mus musculus cDNA clone IMAGE:1920544 5' similar to WP-C13C4.5 CE08130 SUGAR TRANSPORTER ;, mRNA sequence.	35,798	24-Feb-99
GB_HTG3:AC006887	215801	AC006887	Bacteriophage phi-C31 complete genome.	35,798	24-Feb-99
GB_GSS15:AQ605195	459	AQ605195	Caenorhabditis elegans clone Y59H11, *** SEQUENCING IN PROGRESS ***	38,074	10-Jun-99
			PROGRESS ***		
			Caenorhabditis elegans clone Y59H11, *** SEQUENCING IN PROGRESS ***		
			3 unordered pieces.		
			Caenorhabditis elegans clone Y59H11, *** SEQUENCING IN PROGRESS ***		
			3 unordered pieces.		
			HS_2136_B1_C12_T7C CIT Approved Human Genomic Spem Library D Homo sapiens genomic clone Plate=2136 Col=23 Row=F; genomic survey sequence.		

Table 4 (continued)

na00866	1066	GB_BA1:CGORT-4GEN	2398	X95649	Homo sapiens chromosome 14 clone R-108987, *** SEQUENCING IN PROGRESS ***, in ordered pieces. Homo sapiens chromosome 14 clone R-108987, *** SEQUENCING IN PROGRESS ***, in ordered pieces. C.gilanicum ORF4 gene.	Homo sapiens	38,120	15-OCT-1999
		GB_HTG1:CNS00M8S	214599	AL079302		Homo sapiens	38,120	15-OCT-1999
		GB_HTG1:CNS00M8S	214599	AL079302		Homo sapiens	38,120	15-OCT-1999
		GB_BA1:BLDAPAB	3572	Z21502	B.lactofementum dapA and dapB genes for dihydrodipicolinate synthase and dihydrodipicolinate reductase. DNA encoding Brevibacterium dihydrodipicolinic acid reductase.	Corynebacterium glutamicum Corynebacterium glutamicum Corynebacterium glutamicum	99,273 99,301 99,659	10-MAR-1998 16-AUG-93 28-JUL-99
		GB_PAT:E14517	1411	E14517		Unknown.	62,767	01-DEC-1998
		GB_PAT:I92050	567	I92050	Sequence 17 from patent US 5726299.	Unknown.	62,767	3-APR-98
		GB_PAT:I78760	567	I78760	Sequence 16 from patent US 5693781.	Unknown.	36,456	12-NOV-98
		GB_BA2:AE000426	10240	AE000426	Escherichia coli K-12 MG1655 section 316 of 400 of the complete genome.	Escherichia coli	32,762	08-MAR-1999
		GB_BA2:AE001598	11136	AE001598	Chlamydia pneumoniae section 14 of 103 of the complete genome.	Chlamydia pneumoniae	35,849	4-AUG-99
		GB_PL2:AF079370	2897	AF079370	Kluyveromyces fragilis invertase (INV1) gene, complete cds.	Kluyveromyces fragilis	40,138	08-MAR-1999
		GB_BA2:AE001598	11136	AE001598	Chlamydia pneumoniae section 14 of 103 of the complete genome.	Chlamydia pneumoniae	35,076	28-SEP-99
		GB_PR2:HSQ15C24	73192	AJ239325	Homo sapiens chromosome 21 from cosmid LLNLc116-1C16 and LLNLc116-15C24 map 21q22.3 region D21S171-LA161, complete sequence.	Homo sapiens	33,500	6-JUL-98
		GB_G8S4:AQ691923	446	AQ691923	HS_5400_B2_G04_SP8E RPCL-11 Human Male BAC Library Homo sapiens genomic clone Plate=976 Col=8 Row=N, genomic survey sequence.	Homo sapiens	41,127	24-AUG-99
		GB_EST37:Al967802	479	Al967802	Ljnpes12-930-46 Ljnp Lambda HybridZap two-hybrid library Lotus japonicus cDNA clone LP330-12-46 5' similar to 60S ribosomal protein L7A, mRNA sequence.	Lotus japonicus	97,071	3-APR-98
		GB_PAT:I78750	588	I78750	Sequence 6 from patent US 5693781.	Unknown.	97,071	01-DEC-1998
		GB_PAT:I92039	588	I92039	Sequence 6 from patent US 5726299.	Unknown.	39,016	23-NOV-99
		GB_PR3:HS929C8	139190	AL020994	Human DNA sequence from clone 929C8 on chromosome 22q12.1-12.3 Contains CA repeat, GSS, STS, complete sequence.	Homo sapiens	97,561	3-APR-98
		GB_PAT:I78750	588	I78750	Sequence 6 from patent US 5693781.	Unknown.	97,561	01-DEC-1998
		GB_PAT:I92039	588	I92039	Sequence 6 from patent US 5726299.	Unknown.	37,222	3-APR-98
		GB_PAT:I78750	588	I78750	Sequence 6 from patent US 5693781.	Chromobacterium violaceum	39,868	02-OCT-1999
		GB_BA1:AB032799	9077	AB032799	Chromobacterium violaceum violacin biosynthetic gene cluster (vio A, vio B, vio C, vio D), complete cds.	Chromobacterium violaceum	42,760	30-AUG-99
		GB_BA2:AF-172851	10094	AF-172851	Chromobacterium violaceum violacin biosynthetic gene cluster, complete sequence.	Chromobacterium violaceum	39,551	02-OCT-1999
		GB_BA1:AB032799	9077	AB032799	Chromobacterium violaceum violacin biosynthetic gene cluster (vio A, vio B, vio C, vio D), complete cds.	Chromobacterium violaceum		

Table 4 (continued)

ra00992	1629	GB_BA1:BLARGS	2501	Z21501	B.lactofermentum argS and lysA genes for arginyl-tRNA synthetase and diaminopimelate decarboxylase (partial).	Corynebacterium glutamicum	39,003	28-DEC-1993
		GB_BA1:CGLYSA	2344	X54740	Corynebacterium glutamicum argS-lysA operon gene for the upstream region of the arginyl-tRNA synthetase and diaminopimelate decarboxylase (EC 4.1.1.20).	Corynebacterium glutamicum	41,435	30-Jun-93
		GB_PAT:E14508	3579	E14508	DNA encoding Brevibacterium diaminopimelic acid decarboxylase and arginyl-tRNA synthetase.	Corynebacterium glutamicum	40,566	28-Jul-99
ra00983	1599	GB_HTG2:AC008152	24000	AC008152	Leishmania major chromosome 35 clone L7936 strain Friedlin, *** SEQUENCING IN PROGRESS *** 4 unordered pieces.	Leishmania major	38,658	28-Jul-99
		GB_HTG2:AC008152	24000	AC008152	Leishmania major chromosome 35 clone L7936 strain Friedlin, *** SEQUENCING IN PROGRESS *** 4 unordered pieces.	Leishmania major	38,658	28-Jul-99
		GB_HTG3:AC008648	87249	AC008648	Homo sapiens chromosome 5 clone C1978SKB.166E14, *** SEQUENCING IN PROGRESS *** 22 unordered pieces.	Homo sapiens	36,102	3-Aug-99
ra00984	440	GB_BA1:MWINED	3098	D01045	Micromonospora viridifaciens DNA for nrdR protein and neuraminidase, complete cds.	Micromonospora viridifaciens	59,226	2-Feb-99
		GB_PAT:E02375	1881	E02375	Neuraminidase gene.	Micromonospora viridifaciens	59,226	29-Sep-97
		GB_PR4:HUAC004513	101311	AC004513	Homo sapiens Chromosome 16 BAC clone C1987SK-A-926E7, complete sequence.	Homo sapiens	41,204	23-Nov-99
ra01014	2724	GB_BA1:MTV008	63033	AL021246	Mycobacterium tuberculosis H37Rv complete genome; segment 1081162.	Mycobacterium tuberculosis	56,167	17-Jun-98
		GB_BA1:STMAMPEPN	2849	L23172	Streptomyces lividans aminopeptidase N gene, complete cds.	Streptomyces lividans	57,067	18-MAY-1994
		GB_BA1:SC7H2	42655	AL109732	Streptomyces coelicolor cosmid 7H2.	Streptomyces coelicolor A3(2)	37,551	2-Aug-99
ra01059	732	GB_HTG3:AC008154	172241	AC008154	Homo sapiens chromosome 7, *** SEQUENCING IN PROGRESS *** 26 unordered pieces.	Homo sapiens	39,499	8-Sep-99
		GB_HTG3:AC008154	172241	AC008154	Homo sapiens chromosome 7, *** SEQUENCING IN PROGRESS *** 26 unordered pieces.	Homo sapiens	39,499	8-Sep-99
		GB_EST32:A1756574	299	A1756574	ead210.y1 Eimeria M5-6 Macrozolite stage Eimeria tenella cDNA 5', mRNA sequence.	Eimeria tenella	37,793	23-Jun-99
ra01073	954	GB_BA1:BACOUTB	1004	M15811	Bacillus subtilis ouB gene encoding a sporulation protein, complete cds.	Bacillus subtilis	53,723	26-Apr-93
		GB_PR4:AC007938	167237	AC007938	Homo sapiens clone UWGC-djs201 from 7q31, complete sequence.	Homo sapiens	34,322	1-Jul-99
		GB_PL2:ATAC006282	92577	AC006282	Arabidopsis thaliana chromosome II BAC F13K3 genomic sequence, complete sequence.	Arabidopsis thaliana	36,181	13-MAR-1999
ra01120	1401	GB_BA1:MTV008	63033	AL021246	Mycobacterium tuberculosis H37Rv complete genome; segment 1081162.	Mycobacterium tuberculosis	36,715	17-Jun-98
		GB_BA1:CA110321	6710	AJ010321	Caulobacter crescentus partial lig gene and cfpA, cfpX, ion genes.	Caulobacter crescentus	63,311	01-OCT-1998

Table 4 (continued)

GB_BA2:AF150957	4440	AF150957	Azospirillum brasilense trigger factor (tig), heat-shock protein ClpP (clpP), and heat-shock protein ClpX (clpX) genes, complete cds; and Lon protease (lon) gene, partial cds.	Azospirillum brasilense	60,613	7-Jun-99
GB_PR3:HS408N23	97916	Z98048	Human DNA sequence from PAC 408N23 on chromosome 22q13. Contains HIP, HSC70-INTERACTING PROTEIN (PROGESTERONE RECEPTOR-ASSOCIATED P48 PROTEIN), ESTs and STS.	Homo sapiens	34,567	23-Nov-99
GB_BA2:AE01227	26849	AE01227	Trypanosoma pallidum section 43 of 87 of the complete genome.	Trypanosoma pallidum	37,564	16-Jul-98
GB_PR3:HS408N23	97916	Z98048	Human DNA sequence from PAC 408N23 on chromosome 22q13. Contains HIP, HSC70-INTERACTING PROTEIN (PROGESTERONE RECEPTOR-ASSOCIATED P48 PROTEIN), ESTs and STS.	Homo sapiens	34,911	23-Nov-99
GB_BA1:MTCY261	27322	Z97559	Mycobacterium tuberculosis H37Rv complete genome, segment 95/162.	Mycobacterium tuberculosis	38,789	17-Jun-98
GB_HTG4:AC009849	114933	AC009849	Drosophila melanogaster chromosome 2 clone BACR07H03 (D864) RPC1-98.07.H.8 map 31B-31C strain y; on bw sp. *** SEQUENCING IN PROGRESS ***. 55 unordered pieces.	Drosophila melanogaster	39,213	25-OCT-1999
GB_HTG4:AC009849	114933	AC009849	Drosophila melanogaster chromosome 2 clone BACR07H03 (D864) RPC1-98.07.H.8 map 31B-31C strain y; on bw sp. *** SEQUENCING IN PROGRESS ***. 55 unordered pieces.	Drosophila melanogaster	39,213	25-OCT-1999
GB_BA2:AF176799	2943	AF176799	Lactobacillus pentosus PcpQ (pepQ) and catabolite control protein A (ccpA) genes, complete cds.	Lactobacillus pentosus	37,043	5-Sep-99
GB_BA2:AF012084	3082	AF012084	Lactobacillus helveticus prodigiosus (pepQ) gene, complete cds.	Lactobacillus helveticus	45,796	1-Jul-98
GB_EST32:AI728955	611	AI728955	BNLGH12114 Six-day Cotton fiber Gossypium hirsutum cDNA 5' similar to (AC004481) putative permealase (Arabidopsis thaliana), mRNA sequence.	Gossypium hirsutum	37,647	11-Jun-99
GB_BA1:MLCB22	40281	Z98741	Mycobacterium leprae cosmid B22.	Mycobacterium leprae	61,570	22-Aug-97
GB_BA1:MTCY190	34150	Z70283	Mycobacterium tuberculosis H37Rv complete genome, segment 98/162.	Mycobacterium tuberculosis	60,434	17-Jun-98
GB_BA1:SC5F7	40024	AL099872	Streptomyces coelicolor cosmid 5F7.	Streptomyces coelicolor A3(2)	57,011	22-Jul-99
GB_HTG1:CEY116A8	2110000	Z98858	Caenorhabditis elegans chromosome IV clone Y116A8. *** SEQUENCING IN PROGRESS ***. in unordered pieces.	Caenorhabditis elegans	34,843	26-Oct-99
GB_HTG1:CEY116A8	2110000	Z98858	Caenorhabditis elegans chromosome IV clone Y116A8. *** SEQUENCING IN PROGRESS ***. in unordered pieces.	Caenorhabditis elegans	34,843	26-Oct-99
GB_INT:CEY116A8C	260341	AL117204	Caenorhabditis elegans cosmid Y116A8C, complete sequence.	Caenorhabditis elegans	34,843	19-Nov-99
GB_BA1:D90915	130001	D90915	Synechocystis sp. PCC6803 complete genome, 1727, 2137/259-2267259.	Synechocystis sp.	36,538	7-Feb-99
GB_BA1:D90915	130001	D90915	Synechocystis sp. PCC6803 complete genome, 1727, 2137/259-2267259.	Synechocystis sp.	34,512	7-Feb-99

Table 4 (continued)

GB_HTG3C3AC010515	41038	AC010515	Homo sapiens chromosome 19 clone LLNL-R_249H9, *** SEQUENCING IN PROGRESS ***, 31 unordered pieces.	Homo sapiens	33,564	15-Sep-99
GB_OMCFP180RRC	5425	X87224	Canis familiaris mRNA for ribosome receptor, p180.	Canis familiaris	41,229	22-Jan-99
GB_OMCFP180RRC	5425	X87224	Canis familiaris mRNA for ribosome receptor, p180.	Canis familiaris	38,187	22-Jan-99
GB_IN1:CEV47D3A	198814	AL117202	Caenorhabditis elegans cosmid Y47D3A, complete sequence.	Caenorhabditis elegans	36,504	19-Nov-99
GB_PRR4AC006039	176257	AC006039	Homo sapiens clone NH0319F03, complete sequence.	Homo sapiens	34,984	05-MAY-1999
GB_PRR4AC006039	176257	AC006039	Homo sapiens clone NH0319F03, complete sequence.	Homo sapiens	35,951	05-MAY-1999
GB_EST22-A070047	479	A070047	UI-R-C1-h-408-Q-U1 s1 UI-R-C1 Rattus norvegicus cDNA clone UI-Rattus norvegicus	Rattus sp.	36,975	5-Jul-99
GB_RO:375965	625	S75965	THP=Tamm-Horsfall protein [promoter] [rat], Genomic, 625 nt.	Rattus sp.	34,400	27-Jul-95
GB_EST5:H69551	459	H69551	yu01q03.r1 Soares, pineal_gland, N3HPG Homo sapiens cDNA clone IMAGE:232564.5, mRNA sequence.	Homo sapiens	32,969	11-DEC-1995
GB_PL1:NEULCCB	2656	M18334	N crassa (strain TS) lacase gene, complete cds.	Neurospora crassa	44,330	03-MAY-1994
GB_OV:MITRACOMPL	16714	Y16884	Rhea americana complete mitochondrial genome.	Mitochondrion Rhea americana	35,094	19-Jul-99
GB_OV:AF090339	16704	AF090339	Rhea americana mitochondrial, complete genome.	Mitochondrion Rhea americana	35,094	27-MAY-1999
GB_PL2:AF111709	52684	AF111709	Oryza sativa subsp. indica Retrosat 1 retrotransposon and Ty3-Gypsy type Retrosat 2 retrotransposon, complete sequences; and unknown genes.	Oryza sativa subsp. indica	37,410	26-Apr-99
GB_IN1:CELZC250	34372	AF003383	Caenorhabditis elegans cosmid ZC250.	Caenorhabditis elegans	35,506	14-MAY-1997
GB_EST1:Z14808	331	Z14808	CEL5E4 Chris Martin sorted cDNA library Caenorhabditis elegans cDNA clone cr5E4.5, mRNA sequence.	Caenorhabditis elegans	36,890	19-Jun-97
GB_BA1:IMTC165	34331	Z95584	Mycobacterium tuberculosis H37Rv complete genome; segment 50/162.	Mycobacterium tuberculosis	59,298	17-Jun-98
GB_BA1:MSGY348	40556	AD000020	Mycobacterium tuberculosis sequence from clone y348.	Mycobacterium tuberculosis	59,227	10-DEC-1998
GB_BA1:SC5C7	41906	AL031515	Streptomyces coelicolor cosmid 5C7.	Streptomyces coelicolor	39,261	7-Sep-98
GB_BA1:TTAJ5043	837	AJ225043	Thermus thermophilus partial narK gene	Thermus thermophilus	55,245	18-Jun-98
GB_PL2:AC010675	84723	AC010675	Arabidopsis thaliana chromosome 1 (BAC T17F3 genomic sequence, complete sequence).	Arabidopsis thaliana	37,058	11-Nov-99
GB_GSS9:AC170662	518	AC170662	HS_3165_B2_F03_T7 CIT Approved Human Genomic Sperm Library D Homo sapiens genomic clone Plae=3165 Col=6 Row=L, genomic survey sequence.	Homo sapiens	38,610	17-OCT-1998
GB_BA1:D90757	17621	D90757	Escherichia coli genomic DNA (27.3 x 27.7 min).	Escherichia coli	55,445	7-Feb-99
GB_BA1:D90757	15942	D90757	E.coli genomic DNA, Kohara clone #276(33.0-33.3 min.).	Escherichia coli	36,815	29-MAY-1997
GB_BA1:D90758	13860	D90758	Escherichia coli genomic DNA (27.6 x 27.9 min).	Escherichia coli	54,942	7-Feb-99
GB_BA1:SCJ12	35302	AL109989	Streptomyces coelicolor cosmid J12.	Streptomyces coelicolor	62,423	24-Aug-99
GB_BA1:BSNARYWII	12450	Z49884	B. subtilis narK(G,H,I,J,K), ywi(C,D,E) and argS genes.	Bacillus subtilis	57,447	24-Jun-98

Table 4 (continued)

GB_BA1:BSUB0020	212150	Z59123	Bacillus subtilis complete genome (section 20 of 21): from 3798401 to Bacillus subtilis 4010550.	37,129	26-Nov-97
GB_GS511:AQ260413	453	AQ250413	CTIBL-E1-2510B12.TF CTIBL-E1 Homo sapiens genomic clone	41,531	24-OCT-1998
GB_EST20:AA840562	326	AA840562	2510B12, genomic survey sequence.	42,901	27-Feb-98
GB_PAT:A39944	3836	A39944	wv77h07.r1 Striatogene mouse heart (#937316) Mus musculus cDNA Mus musculus clone IMAGE:1261021 5' similar to gb:J04181 Mouse A-X actin mRNA, complete cds (MOUSE); mRNA sequence.	38,764	05-MAR-1997
GB_BA1:FVPBENTA	2519	M88557	Sequence 1 from Patent WO9421807.	40,855	26-Apr-93
GB_PAT:H19994	2516	H19994	Flavobacterium sp. pentachlorophenol 4-monoxygenase gene, complete mRNA.	40,855	07-OCT-1996
GB_BA2:AF059680	2410	AF059680	Flavobacterium sp. pentachlorophenol 4-monoxygenase gene, complete cds.	42,993	27-Apr-99
GB_GS512:AQ332469	459	AQ332469	Sequence 2 from patent US 5512478.	38,208	06-MAR-1999
GB_EST27:AA988532	453	AA988532	Sphingomonas sp. UG30 pentachlorophenol 4-monoxygenase (pcpB) gene, complete cds; and pentachlorophenol 4-monoxygenase reductase (pcpD) gene, partial cds.	39,336	08-MAR-1999
GB_HTG1:HS4342D11	178183	AL121748	HS_5003_A1_H08_3P0E RPO111 Human Male BAC Library Homo sapiens genomic clone Plate=579 Col=15 Row=O, genomic survey sequence.	40,550	23-Nov-99
GB_BA2:AE000745	15085	AE000745	UI-R-CO-ic-d-11-0-UI.s1 UI-R-CO Rattus norvegicus cDNA clone UI-R-CO-ic-d-11-0-UI 3', mRNA sequence.	37,694	25-MAR-1998
GB_BA2:AE000745	15085	AE000745	Homo sapiens chromosome 10 clone RP11-342D11, ** SEQUENCING IN PROGRESS ***, in unordered pieces.	35,567	25-MAR-1998
GB_BA1:AB011413	12070	AB011413	Aquifex aeolicus section 77 of 109 of the complete genome.	57,500	7-Aug-98
GB_BA1:AB011413	12070	AB011413	Aquifex aeolicus section 77 of 109 of the complete genome.	35,655	7-Aug-98
GB_PRA:AC005005	133893	AC005005	Streptomyces griseus genes for Orf2, Orf3, Orf4, Orf5, Orf6, Orf8, partial and complete cds.	38,399	02-MAR-1999
GB_HTG3:AC008257	109187	AC008257	Streptomyces griseus genes for Orf2, Orf3, Orf4, Orf5, Orf6, Orf8, partial and complete cds.	33,741	08-OCT-1999
GB_HTG3:AC008257	109187	AC008257	Streptomyces griseus genes for Orf2, Orf3, Orf4, Orf5, Orf6, Orf8, partial and complete cds.	33,741	08-OCT-1999
GB_BA1:MTV003	13246	AL008883	Homo sapiens PAC clone DU412A9 from 22, complete sequence.	39,369	17-Jun-98
GB_BA1:MSG81529CS	36395	L78824	Drosophila melanogaster chromosome 2 clone BACR08A11 (D916) RPCL-98 08.A.11 map 42A-42A strain y; cn bw sp, ** SEQUENCING IN PROGRESS ***, 93 unordered pieces.	60,624	15-Jun-96
GB_BA1:AB024601	14807	AB024601	Drosophila melanogaster chromosome 2 clone BACR08A11 (D916) RPCL-98 08.A.11 map 42A-42A strain y; cn bw sp, ** SEQUENCING IN PROGRESS ***, 93 unordered pieces.	41,603	12-MAR-1999
			Drosophila melanogaster chromosome 2 clone BACR08A11 (D916) RPCL-98 08.A.11 map 42A-42A strain y; cn bw sp, ** SEQUENCING IN PROGRESS ***, 93 unordered pieces.		
			Mycobacterium tuberculosis H37Rv complete genome, segment 125/162.		
			Mycobacterium leprae cosmid B1529 DNA sequence.		
			Pseudomonas aeruginosa dapD gene for tetrahydrodipicolinate N-succinyltransferase, complete cds, strain PAO1.		

Table 4 (continued)

na01654	1119	GB_GSS4:AQ704352	532	AQ704352	HS_2147_A2_H04_MR CIT Approved Human Genomic Sperm Library, D Homo sapiens genomic clone Plate=2147 Col=6 Row=O, genomic survey sequence.	Homo sapiens	37,838	7-Jul-99
		GB_RO:MMAE000663	250611	AEO00663	Mus musculus TCR beta locus from bases 1 to 250611 (section 1 of 3) of the complete sequence.	Mus musculus	35,799	4-Sep-97
		GB_EST23:AI158428	511	AI158428	u224f12.r1 Soares 2NbMT Mus musculus cDNA clone IMAGE:1446863 5', mRNA sequence.	Mus musculus	41,337	30-Sep-98
na01664	945	GB_OV:AF026198	63155	AF026198	Fugu rubripes neural cell adhesion molecule L1 homolog (L1-CAM) gene, complete cds; putative protein 1 (PUT1) gene, partial cds; mitosis-specific chromosome segregation protein SMC1 homolog (SMC1) gene, complete cds; and calcium channel alpha-1 subunit homolog (CCA1) and putative protein 2 (PUT2) genes, partial cds, complete sequence.	Fugu rubripes	35,187	02-MAY-1998
		GB_PR3:AC004466	122186	AC004466	Homo sapiens 12q13.1 PAC RPCIS-105720 (Roswell Park Cancer Institute Human PAC library) complete sequence.	Homo sapiens	37,382	17-Sep-98
		GB_PR3:AC004466	122186	AC004466	Homo sapiens 12q13.1 PAC RPCIS-105720 (Roswell Park Cancer Institute Human PAC library) complete sequence.	Homo sapiens	37,325	17-Sep-98
na01795	720	GB_BA2:CGU13922	4412	U13922	Corynebacterium glutamicum putative type II 5-cytosine methyltransferase (cgIM) and putative type II restriction endonuclease (cgIR) genes, complete cds.	Corynebacterium glutamicum	99,444	3-Feb-98
		GB_BA1:S96113	1044	S96113	ORF 1 [Neisseria gonorrhoeae, Genomic, 1044 nt].	Neisseria gonorrhoeae	58,320	07-MAY-1993
		GB_PAT:J22080	850	J22080	Sequence 1 from patent US 5525717.	Unknown.	57,722	07-OCT-1996
na01802	954	GB_BA2:AE001519	14062	AE001519	Helicobacter pylori, strain J99 section 80 of 132 of the complete genome.	Helicobacter pylori J99	33,510	20-Jan-99
		GB_GSS5:AQ774071	552	AQ774071	HS_2269_B1_C10_TTC CIT Approved Human Genomic Sperm Library D Homo sapiens genomic clone Plate=2269 Col=19 Row=F, genomic survey sequence.	Homo sapiens	37,957	29-Jul-99
		GB_PR4:AC007469	40807	AC007469	Homo sapiens chromosome 16 clone 306C6, complete sequence.	Homo sapiens	39,140	04-MAY-1999
na01838	842	GB_BA1:SCF15	26440	AL049707	Streptomyces coelicolor cosmid E15.	Streptomyces coelicolor	36,297	22-Apr-99
		GB_HTG3:AC009545	165042	AC009545	Homo sapiens chromosome 11 clone 131_J_04 map 11, *** SEQUENCING IN PROGRESS ***, 8 unordered pieces.	Homo sapiens	37,651	01-OCT-1999
		GB_HTG3:AC009545	165042	AC009545	Homo sapiens chromosome 11 clone 131_J_04 map 11, *** SEQUENCING IN PROGRESS ***, 8 unordered pieces.	Homo sapiens	37,651	01-OCT-1999
		GB_BA1:MTCY24A1	20270	Z95207	Mycobacterium tuberculosis H37Rv complete genome; segment 124/162.	Mycobacterium tuberculosis	38,270	17-Jun-98
na01848	867	GB_EST21:C89252	587	C89252	C89252 Mouse early blastocyst cDNA Mus musculus cDNA clone 01B00061UC08, mRNA sequence.	Mus musculus	37,219	28-MAY-1998
		GB_EST14:AA423340	457	AA423340	ve39d04.r1 Soares mouse mammary gland NbMMMG Mus musculus cDNA clone IMAGE:320519 5', mRNA sequence.	Mus musculus	38,377	16-OCT-1997

Table 4 (continued)

ra01849	1224	GB_BA1:MTCY24A1	20270	Z95207	Mycobacterium tuberculosis H37Rv complete genome; segment 124/162.	Mycobacterium tuberculosis	39,960	17-Jun-98
		GB_BA2:RCPHSYNG	48959	Z11165	R. capsulatus complete photosynthesis gene cluster.	Rhodobacter capsulatus	37,344	2-Sep-99
		GB_BA1:RSP010302	40707	AJ010302	Rhodobacter sphaeroides photosynthetic gene cluster.	Rhodobacter sphaeroides	40,898	27-Aug-99
ra01868	2049	GB_BA1:MTV033	21620	AL021928	Mycobacterium tuberculosis H37Rv complete genome; segment 11/162.	Mycobacterium tuberculosis	38,679	17-Jun-98
		GB_BA1:MLCL622	42498	Z95398	Mycobacterium leprae cosmid L622	Mycobacterium leprae	38,911	24-Jun-97
		GB_BA1:MSGB983CS	36788	L78828	Mycobacterium leprae cosmid B983 DNA sequence.	Mycobacterium leprae	38,933	15-Jun-96
ra01885	924	GB_BA1:MTCY1A10	25949	Z95387	Mycobacterium tuberculosis H37Rv complete genome; segment 117/162.	Mycobacterium tuberculosis	51,094	17-Jun-98
		GB_PR3:HSU220B11	41247	Z69908	Human DNA sequence from cosmid cU220B11, between markers DXS8791 and DXS8038 on chromosome X.	Homo sapiens	39,038	23-Nov-99
		GB_BA1:PDU17435	993	U17435	Paracoccus denitrificans Fnr-like transcriptional activator (nnf) gene, complete cds.	Paracoccus denitrificans	39,390	19-Jul-95
ra01914	526	GB_PR3:AC005796	43843	AC005796	Homo sapiens chromosome 19, cosmid R31408, complete sequence.	Homo sapiens	34,961	06-OCT-1998
		GB_PR3:HS390C10	114231	AL008721	Homo sapiens DNA sequence from BAC 390C10 on chromosome 22q11.21-12.1. Contains an Immunoglobulin LIKE gene and a pseudogene similar to Beta Crystallin. Contains ESTs, STSs, GSSs and taga and tat repeat polymorphisms, complete sequence.	Homo sapiens	39,600	23-Nov-99
		GB_PR3:AC005796	43843	AC005796	Homo sapiens chromosome 19, cosmid R31408, complete sequence.	Homo sapiens	37,725	06-OCT-1998
ra01932	1020	GB_PR3:AC003025	112309	AC003025	Human Chromosome 11p12.2 PAC clone pJ465a11, complete sequence.	Homo sapiens	35,585	23-Jul-98
		GB_GSS3:B78728	312	B78728	CIT-HSP-431E3.TV CIT-HSP Homo sapiens genomic clone 431E3, genomic survey sequence.	Homo sapiens	38,907	25-Jun-98
		GB_PR3:AC003025	112309	AC003025	Human Chromosome 11p12.2 PAC clone pJ465a11, complete sequence.	Homo sapiens	35,859	23-Jul-98
ra01933	726	GB_HTG1:HS74016	169401	AL110119	Homo sapiens chromosome 21 clone RPCIP70401674 map 21q21, *** SEQUENCING IN PROGRESS ***. in unordered pieces.	Homo sapiens	35,302	27-Aug-99
		GB_HTG1:HS74016	169401	AL110119	Homo sapiens chromosome 21 clone RPCIP70401674 map 21q21, *** SEQUENCING IN PROGRESS ***. in unordered pieces.	Homo sapiens	35,302	27-Aug-99
		GB_PR4:AC006032	170282	AC006032	Homo sapiens BAC clone NH0115520 from Y, complete sequence.	Homo sapiens	37,640	27-Feb-99
ra01971	954	GB_HTG3:AC008230	108469	AC008230	Drosophila melanogaster chromosome 2 clone BACR17117 (D934) RPCI-98 17.117 map 53A-53C strain y, on bw sp, *** SEQUENCING IN PROGRESS ***. 108 unordered pieces.	Drosophila melanogaster	35,466	10-Aug-99
		GB_HTG3:AC008230	108469	AC008230	Drosophila melanogaster chromosome 2 clone BACR17117 (D934) RPCI-98 17.117 map 53A-53C strain y, on bw sp, *** SEQUENCING IN PROGRESS ***. 108 unordered pieces.	Drosophila melanogaster	35,466	10-Aug-99

Table 4 (continued)

GB_PR3:AF064860	165382	AF064860	Homo sapiens chromosome 21q22.3 PAC 7024, complete sequence.	Homo sapiens	39,716	2-Jun-98
GB_EST7:D48846	459	D48846	RICS15292A Rice green short Oryza sativa cDNA, mRNA sequence.	Oryza sativa	37,118	2-Aug-95
GB_GSS10:AA195886	595	AQ195886	RPC11-56Q13.TJ RPC1-11 Homo sapiens genomic clone RPC1-11-66Q13, genomic survey sequence.	Homo sapiens	41,000	20-Apr-99
GB_GSS10:AA195886	595	AQ195886	RPC11-56Q13.TJ RPC1-11 Homo sapiens genomic clone RPC1-11-66Q13, genomic survey sequence.	Homo sapiens	34,790	20-Apr-99
GB_EST20:AA855266	406	AA855266	vw70b08.r1 Stragagene mouse heart (#937316) Mus musculus cDNA clone IMAGE:1260279 5', mRNA sequence.	Mus musculus	42,638	05-MAR-1998
GB_EST20:AA855266	406	AA855266	vw70b08.r1 Stragagene mouse heart (#937316) Mus musculus cDNA clone IMAGE:1260279 5', mRNA sequence.	Mus musculus	37,183	05-MAR-1998
GB_BA1:SC5C7	41906	ALD31515	Streptomyces coelicolor cosmid 5C7.	Streptomyces coelicolor	41,732	7-Sep-98
GB_BA1:MITC165	34331	Z95584	Mycobacterium tuberculosis H37Rv complete genome; segment 507162.	Mycobacterium tuberculosis	62,395	17-Jun-98
GB_BA1:SCJ12	35302	AL109989	Streptomyces coelicolor cosmid J12.	Streptomyces coelicolor A3(2)	61,603	24-Aug-99
GB_PAT:E15823	2323	E15823	DNA encoding cell surface protein from Corynebacterium ammoniagenes.	Corynebacterium ammoniagenes	53,942	28-Jul-99
GB_OM:SSAMPTDN	3387	Z29522	S.scrofa mRNA for aminopeptidase N.	Sus scrofa	42,672	26-Sep-94
GB_OVD87992	3181	D87992	Gallus gallus mRNA for aminopeptidase Ey, complete cds.	Gallus gallus	41,554	5-Jun-99
GB_BA1:AP000064	247895	AP000064	Aeropyrum pernix genomic DNA, section 177.	Aeropyrum pernix	39,882	22-Jun-99
GB_PL2:ATAC006587	79262	AC006587	Arabidopsis thaliana chromosome II BAC T17D12 genomic sequence, complete sequence.	Arabidopsis thaliana	38,490	23-MAR-1999
GB_PL2:ATAC006587	79262	AC006587	Arabidopsis thaliana chromosome II BAC T17D12 genomic sequence, complete sequence.	Arabidopsis thaliana	34,863	23-MAR-1999
GB_BA2:AF120718	4137	AF120718	Lactobacillus fermentum urease operon, partial sequence.	Lactobacillus fermentum	56,265	31-MAR-1999
GB_PAT:E03531	2886	E03531	DNA sequence coding for acid urease.	Lactobacillus fermentum	56,265	29-Sep-97
GB_BA1:LBAAJURE	2886	D10605	L. fermentum gene for acid urease.	Lactobacillus fermentum	56,265	2-Feb-99
GB_GSS10:AQ242920	451	AQ242920	HS_206T_A1_E08 MR CIT Approved Human Genomic Sperm Library D Homo sapiens genomic clone Plate=2061 Col=15 Row=1, genomic survey sequence.	Homo sapiens	37,916	03-OCT-1998
GB_IN1:SLMMTTFMF	14503	D29637	Physarum polycephalum mitochondrial DNA.	Mitochondrion Physarum polycephalum	40,335	12-MAY-1999
GB_IN2:AF012249	5542	AF012249	Physarum polycephalum strain aux2-S region of mitochondria derived from mtF plasmid, including URFA, URFC, URFD, URFE, URFF, and URFG genes, complete cds, and URFH gene, partial cds.	Physarum polycephalum	40,335	08-MAY-1998
GB_BA2:AF048784	681	AF048784	Actinomyces naeslundii urease accessory protein (ureG) gene, complete cds.	Actinomyces naeslundii	66,814	9-Feb-99

Table 4 (continued)

GB_BA2:AF056321	5482	AF056321	Actinomyces naeslundii urease gamma subunit UreA (ureA), urease beta subunit UreB (ureB), urease alpha subunit UreC (ureC), urease accessory protein UreE (ureE), urease accessory protein UreF (ureF), urease accessory protein UreG (ureG), and urease accessory protein UreD (ureD) genes, complete cds.	63,686	Actinomyces naeslundii	9-Feb-99
GB_BA2:SSU35248	5773	U35248	Streptococcus salivarius ure cluster: nickel transporter homolog (ureI) gene, partial cds, and urease beta subunit (ureA), gamma subunit (ureB), alpha subunit (ureC), and accessory proteins (ureE), (ureF), (ureG), and (ureD) genes, complete cds.	61,931	Streptococcus salivarius	26-Jan-96
GB_GSS3:BA9054	543	BA9054	RPC11-4113.TV RPC1-11: Homo sapiens genomic clone RPC1-11-4113, genomic survey sequence.	39,161	Homo sapiens	8-Apr-99
GB_PL1:PMCMSGI	3363	L27092	Pneumocystis carinii B-cell receptor (msgI) gene, 3' end.	39,819	Pneumocystis carinii	26-Sep-94
GB_PL2:AF038556	12792	AF038556	Pneumocystis carinii f. sp. hominis variant regions of major surface glycoproteins (msg1, msg3, msg4) genes, partial cds.	33,832	Pneumocystis carinii f. sp. hominis	10-Sep-96
GB_GSS8:AQ051031	914	AQ051031	nbx00004G10r: CUGI Rice BAC Library Oryza sativa genomic clone nbx00004G10r, genomic survey sequence.	32,289	Oryza sativa	24-MAR-1999
GB_GSS8:AQ051031	914	AQ051031	nbx00004N20r: CUGI Rice BAC Library Oryza sativa genomic clone nbx00004N20r, genomic survey sequence.	34,573	Oryza sativa	24-MAR-1999
GB_BA1:CGU35023	3195	U35023	Corynebacterium glutamicum thiosulfate sulfurtransferase (thiR) gene, partial cds, acyl CoA carboxylase (accBC) gene, complete cds.	100,000	Corynebacterium glutamicum	16-Jan-97
GB_BA1:MTCY71	42729	Z92771	Mycobacterium tuberculosis H37Rv complete genome, segment 141/162.	60,380	Mycobacterium tuberculosis	10-Feb-99
GB_BA1:U00012	33312	U00012	Mycobacterium leprae cosmid B1308.	37,660	Mycobacterium leprae	30-Jan-96
GB_HTG2:HS225E12	126464	AL031772	Homo sapiens chromosome 6 clone RP1-225E12 map q24, *** SEQUENCING IN PROGRESS *** in unordered pieces.	35,973	Homo sapiens	03-DEC-1999
GB_HTG2:HS225E12	126464	AL031772	Homo sapiens chromosome 6 clone RP1-225E12 map q24, *** SEQUENCING IN PROGRESS *** in unordered pieces.	35,973	Homo sapiens	03-DEC-1999
GB_HTG2:HS225E12	126464	AL031772	Homo sapiens chromosome 6 clone RP1-225E12 map q24, *** SEQUENCING IN PROGRESS *** in unordered pieces.	36,992	Homo sapiens	03-DEC-1999
GB_BA1:AB020624	1805	AB020624	Corynebacterium glutamicum murl gene for D-glutamate racemase, complete cds.	99,227	Corynebacterium glutamicum	24-Jul-99
GB_EST4:H61527	294	H61527	yo33b09.s1 Soares adult brain N2b4HB55Y Homo sapiens cDNA clone IMAGE:179705 3', mRNA sequence.	40,411	Homo sapiens	18-Sep-95
GB_GSS1:CNS003CM	1101	AL064136	Drosophila melanogaster genome survey sequence T7 end of BAC # BACR08C19 of RPC1-98 library from Drosophila melanogaster (fruit fly), genomic survey sequence.	37,674	Drosophila melanogaster	3-Jun-99

Table 4 (continued)

ra02477	744	GB_HTG4:AC010054	130191	AC010054	Drosophila melanogaster chromosome 3L74E2 clone RPC198-15E10, *** SEQUENCING IN PROGRESS ***, 70 unordered pieces.	Drosophila melanogaster	37,466	16-OCT-1999
		GB_HTG4:AC010054	130191	AC010054	Drosophila melanogaster chromosome 3L74E2 clone RPC198-15E10, *** SEQUENCING IN PROGRESS ***, 70 unordered pieces.	Drosophila melanogaster	37,466	16-OCT-1999
		GB_HTG4:AC009375	137069	AC009375	Drosophila melanogaster chromosome 3L75A1 clone RPC198-44L16, *** SEQUENCING IN PROGRESS ***, 59 unordered pieces.	Drosophila melanogaster	39,118	16-OCT-1999
ra02513	832	GB_BA1:INTER260	373	X92572	M.terae gene for 32 kDa protein (partial).	Mycobacterium terrae	42,895	15-Jan-98
		GB_PL1:AB019229	84284	AB019229	Arabidopsis thaliana genomic DNA, chromosome 3, P1 clone: MDC16, complete sequence.	Arabidopsis thaliana	36,084	20-Nov-99
		GB_PL1:AB019229	84284	AB019229	Arabidopsis thaliana genomic DNA, chromosome 3, P1 clone: MDC16, complete sequence.	Arabidopsis thaliana	35,244	20-Nov-99
ra02631	834	GB_BA1:CGLATTB	271	X89850	C.glutamicum DNA for attB region.	Corynebacterium glutamicum	40,590	8-Aug-96
		GB_EST11:AA239557	423	AA239557	mv2504.11 GuayWoodford Beiter mouse kidney day 0 Mus musculus cDNA clone IMAGE:656095 5' similar to gpX52634 Murine tm oncogene for tm protein (MOUSE); mRNA sequence.	Mus musculus	38,760	12-MAR-1997
		GB_BA1:RSPYPPCL	6500	AJ002398	Rhodobacter sphaeroides pyp and pcd genes, and orfA, orfB, orfC, orfD, orfE, orfF.	Rhodobacter sphaeroides	37,091	17-DEC-1998
ra02548	314	GB_BA2:AF127374	63734	AF127374	Streptomyces lavendulae LinA homolog, cytochrome P450 hydroxylase ORF4, cytochrome P450 hydroxylase ORF3, MitT (mitT), Mts (mts), MitR (mitR), MitQ (mitQ), MitP (mitP), MitO (mitO), MitN (mitN), MitM (mitM), MitL (mitL), MitK (mitK), MitJ (mitJ), MitI (mitI), MitH (mitH), MitG (mitG), MitF (mitF), MitE (mitE), MitD (mitD), MitC (mitC), MitB (mitB), MitA (mitA), MmcA (mmcA), MmcB (mmcB), MmcC (mmcC), MmcD (mmcD), MmcE (mmcE), MmcF (mmcF), MmcG (mmcG), MmcH (mmch), MmcI (mmci), MmcJ (mmcj), MmcK (mmck), MmcL (mmcl), MmcM (mmcm), MmcN (mmcn), MmcO (mmco), MmcP (mmp), MmcQ (mmcq), MmcR (mmcr), MmcS (mms), MmcT (mmct), MmcU (mmcu), MmcV (mmcv), Mct (mct), MmcW (mmcw), MmcX (mmcx), and MmcY (mmcy) genes, complete cds, and unknown genes.	Streptomyces lavendulae	66,242	27-MAY-1999

Table 4 (continued)

GB_BA2:AF127374	63734	AF127374	Streptomyces lavendulae LinA homolog, cytochrome P450 hydroxylase ORF4, cytochrome P450 hydroxylase ORF3, MIT (mitT), Mts (mitS), Mtr (mitR), MtrQ (mitQ), MIP (mitP), MIO (mitO), MtrN (mitN), MtrM (mitM), MtrL (mitL), MtrK (mitK), MtrJ (mitJ), MtrI (mitI), MtrH (mitH), MtrG (mitG), MtrF (mitF), MtrE (mitE), MtrD (mitD), MtrC (mitC), MtrB (mitB), MtrA (mitA), MtrC2 (mitC2), MtrCB (mitCB), MtrCD (mitCD), MtrCE (mitCE), MtrCF (mitCF), MtrCG (mitCG), MtrCH (mitCH), MtrCL (mitCL), MtrCJ (mitCJ), MtrCK (mitCK), MtrCL (mitCL), MtrCM (mitCM), MtrCN (mitCN), MtrCO (mitCO), MtrD (mitD), MtrCP (mitCP), MtrCQ (mitCQ), MtrCR (mitCR), MtrCS (mitCS), MtrCT (mitCT), MtrCU (mitCU), MtrCV (mitCV), MtrC (mitC), MtrCW (mitCW), MtrCX (mitCX), and MtrCY (mitCY) genes, complete cds, and unknown genes.	Streptomyces lavendulae	38,411	27-MAY-1999
GB_GSS4:AQ741886	742	AQ741886	HS_5569_B2_B02_SP6 RPC1-11 Human Male BAC Library Homo sapiens genomic clone Plate=1145 Col=4 Row=D, genomic survey sequence.	Homo sapiens	38,907	16-Jul-99
GB_EST18:AA567307	741	AA567307	HL01004.5prime HL Drosophila melanogaster head BlueScript sequence.	Drosophila melanogaster	38,736	28-Nov-98
GB_EST27:A1402394	630	A1402394	Drosophila melanogaster cDNA clone HL01004.5prime, mRNA sequence.	Drosophila melanogaster	41,308	8-Feb-99
GB_GSS10:AC237646	715	AQ237646	RPC111-61B.TJB RPC1-11 Homo sapiens genomic clone RPC1-11-619, genomic survey sequence.	Homo sapiens	44,340	21-Apr-99
GB_EST32:A1726448	562	A1726448	BNLGH5854 Six-day Cotton fiber Gossypium hirsutum cDNA 5' similar to (U53418) UDP-glucose dehydrogenase [Glycine max], mRNA sequence.	Gossypium hirsutum	37,003	11-Jun-99
GB_EST32:A1726198	608	A1726198	BNLGH5243 Six-day Cotton fiber Gossypium hirsutum cDNA 5' similar to (U53418) UDP-glucose dehydrogenase [Glycine max], mRNA sequence.	Gossypium hirsutum	40,925	11-Jun-99
GB_PR4:AC002992	154848	AC002992	Homo sapiens chromosome Y, clone 203M13, complete sequence.	Homo sapiens	38,039	19-OCT-1999
GB_EST4:H29653	415	H29653	ym58101.1 Soares infant brain 1N1B Homo sapiens cDNA clone IMAGE52678 5' similar to SP-OXDD_BOVIN P31228 D-ASPARTATE OXIDASE, mRNA sequence.	Homo sapiens	39,036	17-Jul-95
GB_PR3:HSD.261K5	131974	AL050350	Human DNA sequence from clone 261K5 on chromosome 6q21-22.1. Contains the 3' part of the gene for a novel organic cation transporter (BAC ORF RG331P03), the DDO gene for D-aspartate oxidase (EC 1.4.3.1), ESTs, STSs, GSSs and two putative CpG islands, complete sequence.	Homo sapiens	35,557	23-Nov-99

Table 4 (continued)

rx02589	888	GB_EST2:R20147	494	R20147	yg18n02.r1 Soares infant brain 1MB Homo sapiens cDNA clone IMAGE:32866 5' similar to SP:OXDD_BOVIN F31228 D-ASPARTATE OXIDASE; mRNA sequence.	Homo sapiens	36,437	17-Apr-95
		GB_HTG1:CEY6E2	186306	Z56799	Caenorhabditis elegans chromosome V clone Y6E2 *** SEQUENCING IN PROGRESS *** in unordered pieces.	Caenorhabditis elegans	37,979	02-OCT-1997
		GB_HTG1:CEY6E2	186306	Z56799	Caenorhabditis elegans chromosome V clone Y6E2 *** SEQUENCING IN PROGRESS *** in unordered pieces.	Caenorhabditis elegans	37,979	02-OCT-1997
		GB_HTG3:AC011690	72277	AC011690	Homo sapiens clone 17_E_13, LOW-PASS SEQUENCE SAMPLING.	Homo sapiens	35,814	10-OCT-1998
rx02592	894	GB_BA1:MSG8983CS	36788	L78828	Mycobacterium leprae cosmid B983 DNA sequence.	Mycobacterium leprae	53,235	15-Jun-96
		GB_GSS9:AC107023	487	AC107023	HS_2270_B2_F05_MR CIT Approved Human Genomic Sperm Library D Homo sapiens genomic clone Plate-2270 Col=10 Row=L, genomic survey sequence.	Homo sapiens	39,666	16-OCT-1998
		GB_GSS12:AC349387	781	AC349387	RPC11-118H16.T3 RPC1-11 Homo sapiens genomic clone RPC1-11-118H16, genomic survey sequence.	Homo sapiens	34,204	07-MAY-1999
rx02603	1119	GB_BA1:MTV026	23740	AL022076	Mycobacterium tuberculosis H37Rv complete genome, segment 157/162.	Mycobacterium tuberculosis	37,975	24-Jun-99
		GB_IN2:AC005714	177740	AC005714	Drosophila melanogaster, chromosome 2R, region 58D4-58E2, BAC clone BACR48M13, complete sequence.	Drosophila melanogaster	41,226	01-MAY-1999
		GB_EST18:AA776050	218	AA776050	ac76e10.s1 Stratiagene lung (#937210) Homo sapiens cDNA clone IMAGE:868564, 3' similar to gb:Y00371.mn1 HEAT SHOCK COGNATE 71 KD PROTEIN (HUMAN); mRNA sequence.	Homo sapiens	40,826	5-Feb-98
rx02630	1446	GB_BA1:MLC1373	37304	AL036500	Mycobacterium leprae cosmid L373.	Mycobacterium leprae	49,015	27-Aug-99
		GB_BA1:MTV044	16150	AL021999	Mycobacterium tuberculosis H37Rv complete genome, segment 45/162.	Mycobacterium tuberculosis	49,192	17-Jun-98
rx02643	1167	GB_BA1:MLU16180	38675	U15180	Mycobacterium leprae cosmid B1798.	Mycobacterium leprae	45,621	09-MAR-1995
		GB_EST37:AI950576	308	AI950576	wx52e08.x1 NCI CGAP_Lu28 Homo sapiens cDNA clone IMAGE:2547302, 3' mRNA sequence.	Homo sapiens	40,909	6-Sep-99
		GB_EST37:AI950576	308	AI950576	wx52e08.x1 NCI CGAP_Lu28 Homo sapiens cDNA clone IMAGE:2547302, 3' mRNA sequence.	Homo sapiens	40,288	6-Sep-99
rx02644	774	GB_EST34:AV149547	302	AV149547	AV149547 Mus musculus C57BL/6J 10-11 day embryo Mus musculus cDNA clone 2810489D03, mRNA sequence.	Mus musculus	38,627	5-Jul-99
		GB_EST35:AV156221	271	AV156221	AV156221 Mus musculus head C57BL/6J 12-day embryo Mus musculus cDNA clone 3000001C24, mRNA sequence.	Mus musculus	33,990	7-Jul-99
		GB_EST32:AV054919	274	AV054919	AV054919 Mus musculus pancreas C57BL/6J adult Mus musculus cDNA clone 1810033C08, mRNA sequence.	Mus musculus	36,585	23-Jun-99
rx02745	902	GB_BA1:MTV007	32806	AL021184	Mycobacterium tuberculosis H37Rv complete genome, segment 64/162.	Mycobacterium tuberculosis	39,298	17-Jun-98

Table 4 (continued)

GB_BA2:AF027770	30683	AF027770	Mycobacterium smegmatis FxbA (fxbA) gene, partial cds; FxbB (fxbB), FxbC (fxbC), and FxbD (fxbD) genes, complete cds; and unknown genes.	Mycobacterium smegmatis 55,125	03-DEC-1998
GB_BA2:SAU43537	3938	U43537	Streptomyces argillaceus mitramycin resistance determinant, ATP-binding protein (mtfA) and membrane protein (mtfB) genes, complete cds.	Streptomyces argillaceus 46,868	5-Sep-96
GB_BA1:CAJ10319	5388	AJ010319	Corynebacterium glutamicum amfP, glnB, glnD genes and partial ftsY and srp genes.	Corynebacterium glutamicum 100,000	14-MAY-1989
GB_BA1:MTCY338	29372	Z74697	Mycobacterium tuberculosis H37Rv complete genome; segment 127/162.	Mycobacterium tuberculosis 39,785	17-Jun-98
GB_HTG3:AC008733	216140	AC008733	Homo sapiens chromosome 19 clones CITB-E1_2525/15, *** SEQUENCING IN PROGRESS ***; 72 unordered pieces.	Homo sapiens 35,688	3-Aug-99
GB_BA1:BFU64514	3637	U64514	Bacillus firmus dppABC operon; dipeptide transporter protein dppA gene, partial cds, and dipeptide transporter proteins dppB and dppC genes, complete cds.	Bacillus firmus 36,859	1-Feb-97
GB_INT1:CET04C10	20958	Z69885	Caenorhabditis elegans cosmid T04C10, complete sequence.	Caenorhabditis elegans 35,934	2-Sep-99
GB_EST35:AI823090	720	AI823090	L30-944T3 ice plant Lambda Uni-Zap XR expression library, 30 hours NaCl treatment; Mesembryanthemum crystallinum cDNA clone L30-944 5' similar to 60S ribosomal protein L36 (AC004684) [Arabidopsis thaliana], mRNA sequence.	Mesembryanthemum crystallinum 35,770	21-Jul-99
GB_BA1:CJY13333	3315	Y13333	Campylobacter jejuni cfbB gene.	Campylobacter jejuni 53,400	12-Apr-99
GB_BA2:AF065404	181654	AF065404	Bacillus anthracis virulence plasmid PX01, complete sequence.	Bacillus anthracis 45,168	20-OCT-1999
GB_PL2:AC006601	110684	AC006601	Arabidopsis thaliana chromosome V map near 60.5 cM, complete sequence.	Arabidopsis thaliana 36,680	22-Feb-99

Exemplification

Example 1: Preparation of total genomic DNA of *Corynebacterium glutamicum* ATCC 13032

- 5 A culture of *Corynebacterium glutamicum* (ATCC 13032) was grown overnight at 30°C with vigorous shaking in BHI medium (Difco). The cells were harvested by centrifugation, the supernatant was discarded and the cells were resuspended in 5 ml buffer-I (5% of the original volume of the culture — all indicated volumes have been calculated for 100 ml of culture volume). Composition of buffer-I: 140.34 g/l sucrose,
- 10 2.46 g/l $\text{MgSO}_4 \times 7\text{H}_2\text{O}$, 10 ml/l KH_2PO_4 solution (100 g/l, adjusted to pH 6.7 with KOH), 50 ml/l M12 concentrate (10 g/l $(\text{NH}_4)_2\text{SO}_4$, 1 g/l NaCl, 2 g/l $\text{MgSO}_4 \times 7\text{H}_2\text{O}$, 0.2 g/l CaCl_2 , 0.5 g/l yeast extract (Difco), 10 ml/l trace-elements-mix (200 mg/l $\text{FeSO}_4 \times \text{H}_2\text{O}$, 10 mg/l $\text{ZnSO}_4 \times 7\text{H}_2\text{O}$, 3 mg/l $\text{MnCl}_2 \times 4\text{H}_2\text{O}$, 30 mg/l H_3BO_3 , 20 mg/l $\text{CoCl}_2 \times 6\text{H}_2\text{O}$, 1 mg/l $\text{NiCl}_2 \times 6\text{H}_2\text{O}$, 3 mg/l $\text{Na}_2\text{MoO}_4 \times 2\text{H}_2\text{O}$, 500 mg/l complexing agent
- 15 (EDTA or citric acid), 100 ml/l vitamins-mix (0.2 mg/l biotin, 0.2 mg/l folic acid, 20 mg/l p-amino benzoic acid, 20 mg/l riboflavin, 40 mg/l ca-pantothenate, 140 mg/l nicotinic acid, 40 mg/l pyridoxole hydrochloride, 200 mg/l myo-inositol). Lysozyme was added to the suspension to a final concentration of 2.5 mg/ml. After an approximately 4 h incubation at 37°C, the cell wall was degraded and the resulting
- 20 protoplasts are harvested by centrifugation. The pellet was washed once with 5 ml buffer-I and once with 5 ml TE-buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8). The pellet was resuspended in 4 ml TE-buffer and 0.5 ml SDS solution (10%) and 0.5 ml NaCl solution (5 M) are added. After adding of proteinase K to a final concentration of 200 µg/ml, the suspension is incubated for ca.18 h at 37°C. The DNA was purified by
- 25 extraction with phenol, phenol-chloroform-isoamylalcohol and chloroform-isoamylalcohol using standard procedures. Then, the DNA was precipitated by adding 1/50 volume of 3 M sodium acetate and 2 volumes of ethanol, followed by a 30 min incubation at -20°C and a 30 min centrifugation at 12,000 rpm in a high speed centrifuge using a SS34 rotor (Sorvall). The DNA was dissolved in 1 ml TE-buffer containing 20
- 30 µg/ml RNaseA and dialysed at 4°C against 1000 ml TE-buffer for at least 3 hours. During this time, the buffer was exchanged 3 times. To aliquots of 0.4 ml of the dialysed DNA solution, 0.4 ml of 2 M LiCl and 0.8 ml of ethanol are added. After a 30

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min incubation at -20°C, the DNA was collected by centrifugation (13,000 rpm, Biofuge Fresco, Heraeus, Hanau, Germany). The DNA pellet was dissolved in TE-buffer. DNA prepared by this procedure could be used for all purposes, including southern blotting or construction of genomic libraries.

Example 2: Construction of genomic libraries in *Escherichia coli* of *Corynebacterium glutamicum* ATCC13032.

Using DNA prepared as described in Example 1, cosmid and plasmid libraries were constructed according to known and well established methods (see e.g., Sambrook, J. *et al.* (1989) "Molecular Cloning : A Laboratory Manual", Cold Spring Harbor Laboratory Press, or Ausubel, F.M. *et al.* (1994) "Current Protocols in Molecular Biology", John Wiley & Sons.)

Any plasmid or cosmid could be used. Of particular use were the plasmids pBR322 (Sutcliffe, J.G. (1979) *Proc. Natl. Acad. Sci. USA*, 75:3737-3741); pACYC177 (Change & Cohen (1978) *J. Bacteriol* 134:1141-1156), plasmids of the pBS series (pBSSK+, pBSSK- and others; Stratagene, LaJolla, USA), or cosmids as SuperCos1 (Stratagene, LaJolla, USA) or Lorist6 (Gibson, T.J., Rosenthal A. and Waterson, R.H. (1987) *Gene* 53:283-286. Gene libraries specifically for use in *C. glutamicum* may be constructed using plasmid pSL109 (Lee, H.-S. and A. J. Sinskey (1994) *J. Microbiol. Biotechnol.* 4: 256-263).

Example 3: DNA Sequencing and Computational Functional Analysis

Genomic libraries as described in Example 2 were used for DNA sequencing according to standard methods, in particular by the chain termination method using ABI377 sequencing machines (see e.g., Fleischman, R.D. *et al.* (1995) "Whole-genome Random Sequencing and Assembly of *Haemophilus Influenzae* Rd., *Science*, 269:496-512). Sequencing primers with the following nucleotide sequences were used: 5'-GGAAACAGTATGACCATG-3' or 5'-GTAAAACGACGGCCAGT-3'.

Example 4: *In vivo* Mutagenesis

In vivo mutagenesis of *Corynebacterium glutamicum* can be performed by passage of plasmid (or other vector) DNA through *E. coli* or other microorganisms (e.g. *Bacillus* spp. or yeasts such as *Saccharomyces cerevisiae*) which are impaired in their capabilities to maintain

the integrity of their genetic information. Typical mutator strains have mutations in the genes for the DNA repair system (e.g., mutHLS, mutD, mutT, etc.; for reference, see Rupp, W.D. (1996) DNA repair mechanisms, in: *Escherichia coli* and *Salmonella*, p. 2277-2294, ASM: Washington.) Such strains are well known to those of ordinary skill in the art. The use of such strains is illustrated, for example, in Greener, A. and Callahan, M. (1994) *Strategies* 7: 32-34.

Example 5: DNA Transfer Between *Escherichia coli* and *Corynebacterium glutamicum*

Several *Corynebacterium* and *Brevibacterium* species contain endogenous plasmids (as e.g., pHM1519 or pBL1) which replicate autonomously (for review see, e.g., Martin, J.F. *et al.* (1987) *Biotechnology*, 5:137-146). Shuttle vectors for *Escherichia coli* and *Corynebacterium glutamicum* can be readily constructed by using standard vectors for *E. coli* (Sambrook, J. *et al.* (1989), "Molecular Cloning: A Laboratory Manual", Cold Spring Harbor Laboratory Press or Ausubel, F.M. *et al.* (1994) "Current Protocols in Molecular Biology", John Wiley & Sons) to which a origin of replication for and a suitable marker from *Corynebacterium glutamicum* is added. Such origins of replication are preferably taken from endogenous plasmids isolated from *Corynebacterium* and *Brevibacterium* species. Of particular use as transformation markers for these species are genes for kanamycin resistance (such as those derived from the Tn5 or Tn903 transposons) or chloramphenicol (Winnacker, E.L. (1987) "From Genes to Clones — Introduction to Gene Technology, VCH, Weinheim). There are numerous examples in the literature of the construction of a wide variety of shuttle vectors which replicate in both *E. coli* and *C. glutamicum*, and which can be used for several purposes, including gene over-expression (for reference, see e.g., Yoshihama, M. *et al.* (1985) *J. Bacteriol.* 162:591-597, Martin J.F. *et al.* (1987) *Biotechnology*, 5:137-146 and Eikmanns, B.J. *et al.* (1991) *Gene*, 102:93-98).

Using standard methods, it is possible to clone a gene of interest into one of the shuttle vectors described above and to introduce such a hybrid vectors into strains of *Corynebacterium glutamicum*. Transformation of *C. glutamicum* can be achieved by protoplast transformation (Kastsumata, R. *et al.* (1984) *J. Bacteriol.* 159:306-311), electroporation (Liebl, E. *et al.* (1989) *FEMS Microbiol. Letters*, 53:399-303) and in cases where special vectors are used, also by conjugation (as described e.g. in Schäfer, A *et al.*

(1990) *J. Bacteriol.* 172:1663-1666). It is also possible to transfer the shuttle vectors for *C. glutamicum* to *E. coli* by preparing plasmid DNA from *C. glutamicum* (using standard methods well-known in the art) and transforming it into *E. coli*. This transformation step can be performed using standard methods, but it is advantageous to use an *Mcr*-deficient

5 *E. coli* strain, such as NM522 (Gough & Murray (1983) *J. Mol. Biol.* 166:1-19).

Genes may be overexpressed in *C. glutamicum* strains using plasmids which comprise pCG1 (U.S. Patent No. 4,617,267) or fragments thereof, and optionally the gene for kanamycin resistance from TN903 (Grindley, N.D. and Joyce, C.M. (1980) *Proc. Natl. Acad. Sci. USA* 77(12): 7176-7180). In addition, genes may be

10 overexpressed in *C. glutamicum* strains using plasmid pSL109 (Lee, H.-S. and A. J. Sinskey (1994) *J. Microbiol. Biotechnol.* 4: 256-263).

Aside from the use of replicative plasmids, gene overexpression can also be achieved by integration into the genome. Genomic integration in *C. glutamicum* or other *Corynebacterium* or *Brevibacterium* species may be accomplished by well-known

15 methods, such as homologous recombination with genomic region(s), restriction endonuclease mediated integration (REMI) (see, e.g., DE Patent 19823834), or through the use of transposons. It is also possible to modulate the activity of a gene of interest by modifying the regulatory regions (e.g., a promoter, a repressor, and/or an enhancer) by sequence modification, insertion, or deletion using site-directed methods (such as

20 homologous recombination) or methods based on random events (such as transposon mutagenesis or REMI). Nucleic acid sequences which function as transcriptional terminators may also be inserted 3' to the coding region of one or more genes of the invention; such terminators are well-known in the art and are described, for example, in Winnacker, E.L. (1987) *From Genes to Clones – Introduction to Gene Technology*. VCH:

25 Weinheim.

Example 6: Assessment of the Expression of the Mutant Protein

Observations of the activity of a mutated protein in a transformed host cell rely on the fact that the mutant protein is expressed in a similar fashion and in a similar quantity to

30 that of the wild-type protein. A useful method to ascertain the level of transcription of the mutant gene (an indicator of the amount of mRNA available for translation to the gene product) is to perform a Northern blot (for reference see, for example, Ausubel *et al.*

(1988) Current Protocols in Molecular Biology, Wiley: New York), in which a primer designed to bind to the gene of interest is labeled with a detectable tag (usually radioactive or chemiluminescent), such that when the total RNA of a culture of the organism is extracted, run on gel, transferred to a stable matrix and incubated with this probe, the binding and quantity of binding of the probe indicates the presence and also the quantity of mRNA for this gene. This information is evidence of the degree of transcription of the mutant gene. Total cellular RNA can be prepared from *Corynebacterium glutamicum* by several methods, all well-known in the art, such as that described in Bormann, E.R. *et al.* (1992) *Mol. Microbiol.* 6: 317-326.

- 10 To assess the presence or relative quantity of protein translated from this mRNA, standard techniques, such as a Western blot, may be employed (see, for example, Ausubel *et al.* (1988) Current Protocols in Molecular Biology, Wiley: New York). In this process, total cellular proteins are extracted, separated by gel electrophoresis, transferred to a matrix such as nitrocellulose, and incubated with a probe, such as an antibody, which
- 15 specifically binds to the desired protein. This probe is generally tagged with a chemiluminescent or colorimetric label which may be readily detected. The presence and quantity of label observed indicates the presence and quantity of the desired mutant protein present in the cell.

20 **Example 7: Growth of Genetically Modified *Corynebacterium glutamicum* — Media and Culture Conditions**

- Genetically modified *Corynebacteria* are cultured in synthetic or natural growth media. A number of different growth media for *Corynebacteria* are both well-known and readily available (Lieb *et al.* (1989) *Appl. Microbiol. Biotechnol.*, 32:205-210; von der
- 25 Osten *et al.* (1998) *Biotechnology Letters*, 11:11-16; Patent DE 4,120,867; Liebl (1992) "The Genus *Corynebacterium*, in: The Prokaryotes, Volume II, Balows, A. *et al.*, eds. Springer-Verlag). These media consist of one or more carbon sources, nitrogen sources, inorganic salts, vitamins and trace elements. Preferred carbon sources are sugars, such as
- 30 mono-, di-, or polysaccharides. For example, glucose, fructose, mannose, galactose, ribose, sorbose, ribulose, lactose, maltose, sucrose, raffinose, starch or cellulose serve as very good carbon sources. It is also possible to supply sugar to the media via complex compounds such as molasses or other by-products from sugar refinement. It can also be

advantageous to supply mixtures of different carbon sources. Other possible carbon sources are alcohols and organic acids, such as methanol, ethanol, acetic acid or lactic acid. Nitrogen sources are usually organic or inorganic nitrogen compounds, or materials which contain these compounds. Exemplary nitrogen sources include ammonia gas or
5 ammonia salts, such as NH_4Cl or $(\text{NH}_4)_2\text{SO}_4$, NH_4OH , nitrates, urea, amino acids or complex nitrogen sources like corn steep liquor, soy bean flour, soy bean protein, yeast extract, meat extract and others.

Inorganic salt compounds which may be included in the media include the chloride-, phosphorous- or sulfate- salts of calcium, magnesium, sodium, cobalt,
10 molybdenum, potassium, manganese, zinc, copper and iron. Chelating compounds can be added to the medium to keep the metal ions in solution. Particularly useful chelating compounds include dihydroxyphenols, like catechol or protocatechuate, or organic acids, such as citric acid. It is typical for the media to also contain other growth factors, such as vitamins or growth promoters, examples of which include biotin, riboflavin, thiamin, folic
15 acid, nicotinic acid, pantothenate and pyridoxin. Growth factors and salts frequently originate from complex media components such as yeast extract, molasses, corn steep liquor and others. The exact composition of the media compounds depends strongly on the immediate experiment and is individually decided for each specific case. Information about media optimization is available in the textbook "Applied Microbiol. Physiology, A
20 Practical Approach (*eds.* P.M. Rhodes, P.F. Stanbury, IRL Press (1997) pp. 53-73, ISBN 0 19 963577 3). It is also possible to select growth media from commercial suppliers, like standard 1 (Merck) or BHI (grain heart infusion, DIFCO) or others.

All medium components are sterilized, either by heat (20 minutes at 1.5 bar and 121°C) or by sterile filtration. The components can either be sterilized together or, if
25 necessary, separately. All media components can be present at the beginning of growth, or they can optionally be added continuously or batchwise.

Culture conditions are defined separately for each experiment. The temperature should be in a range between 15°C and 45°C. The temperature can be kept constant or can be altered during the experiment. The pH of the medium should be in the range of 5 to
30 8.5, preferably around 7.0, and can be maintained by the addition of buffers to the media. An exemplary buffer for this purpose is a potassium phosphate buffer. Synthetic buffers such as MOPS, HEPES, ACES and others can alternatively or simultaneously be used. It

is also possible to maintain a constant culture pH through the addition of NaOH or NH₄OH during growth. If complex medium components such as yeast extract are utilized, the necessity for additional buffers may be reduced, due to the fact that many complex compounds have high buffer capacities. If a fermentor is utilized for culturing the micro-organisms, the pH can also be controlled using gaseous ammonia.

The incubation time is usually in a range from several hours to several days. This time is selected in order to permit the maximal amount of product to accumulate in the broth. The disclosed growth experiments can be carried out in a variety of vessels, such as microtiter plates, glass tubes, glass flasks or glass or metal fermentors of different sizes.

For screening a large number of clones, the microorganisms should be cultured in microtiter plates, glass tubes or shake flasks, either with or without baffles. Preferably 100 ml shake flasks are used, filled with 10% (by volume) of the required growth medium. The flasks should be shaken on a rotary shaker (amplitude 25 mm) using a speed-range of 100 – 300 rpm. Evaporation losses can be diminished by the maintenance of a humid atmosphere; alternatively, a mathematical correction for evaporation losses should be performed.

If genetically modified clones are tested, an unmodified control clone or a control clone containing the basic plasmid without any insert should also be tested. The medium is inoculated to an OD₆₀₀ of 0.5 – 1.5 using cells grown on agar plates, such as CM plates (10 g/l glucose, 2,5 g/l NaCl, 2 g/l urea, 10 g/l polypeptone, 5 g/l yeast extract, 5 g/l meat extract, 22 g/l NaCl, 2 g/l urea, 10 g/l polypeptone, 5 g/l yeast extract, 5 g/l meat extract, 22 g/l agar, pH 6.8 with 2M NaOH) that had been incubated at 30°C. Inoculation of the media is accomplished by either introduction of a saline suspension of *C. glutamicum* cells from CM plates or addition of a liquid preculture of this bacterium.

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Example 8 – *In vitro* Analysis of the Function of Mutant Proteins

The determination of activities and kinetic parameters of enzymes is well established in the art. Experiments to determine the activity of any given altered enzyme must be tailored to the specific activity of the wild-type enzyme, which is well within the ability of one of ordinary skill in the art. Overviews about enzymes in general, as well as specific details concerning structure, kinetics, principles, methods,

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- applications and examples for the determination of many enzyme activities may be found, for example, in the following references: Dixon, M., and Webb, E.C., (1979) *Enzymes*. Longmans: London; Fersht, (1985) *Enzyme Structure and Mechanism*. Freeman: New York; Walsh, (1979) *Enzymatic Reaction Mechanisms*. Freeman: San Francisco; Price, N.C., Stevens, L. (1982) *Fundamentals of Enzymology*. Oxford Univ. Press: Oxford; Boyer, P.D., ed. (1983) *The Enzymes*, 3rd ed. Academic Press: New York; Bisswanger, H., (1994) *Enzymkinetik*, 2nd ed. VCH: Weinheim (ISBN 3527300325); Bergmeyer, H.U., Bergmeyer, J., Graßl, M., eds. (1983-1986) *Methods of Enzymatic Analysis*, 3rd ed., vol. I-XII, Verlag Chemie: Weinheim; and Ullmann's Encyclopedia of Industrial Chemistry (1987) vol. A9, "Enzymes". VCH: Weinheim, p. 352-363.

- The activity of proteins which bind to DNA can be measured by several well-established methods, such as DNA band-shift assays (also called gel retardation assays). The effect of such proteins on the expression of other molecules can be measured using reporter gene assays (such as that described in Kolmar, H. *et al.* (1995) *EMBO J.* 14: 3895-3904 and references cited therein). Reporter gene test systems are well known and established for applications in both pro- and eukaryotic cells, using enzymes such as beta-galactosidase, green fluorescent protein, and several others.

- The determination of activity of membrane-transport proteins can be performed according to techniques such as those described in Gennis, R.B. (1989) "Pores, Channels and Transporters", in *Biomembranes, Molecular Structure and Function*, Springer: Heidelberg, p. 85-137; 199-234; and 270-322.

Example 9: Analysis of Impact of Mutant Protein on the Production of the Desired Product

- The effect of the genetic modification in *C. glutamicum* on production of a desired compound (such as an amino acid) can be assessed by growing the modified microorganism under suitable conditions (such as those described above) and analyzing the medium and/or the cellular component for increased production of the desired product (*i.e.*, an amino acid). Such analysis techniques are well known to one of ordinary skill in the art, and include spectroscopy, thin layer chromatography, staining methods of various kinds, enzymatic and microbiological methods, and analytical

- chromatography such as high performance liquid chromatography (see, for example, Ullman, Encyclopedia of Industrial Chemistry, vol. A2, p. 89-90 and p. 443-613, VCH: Weinheim (1985); Fallon, A. *et al.*, (1987) "Applications of HPLC in Biochemistry" in: Laboratory Techniques in Biochemistry and Molecular Biology, vol. 17; Rehm *et al.*
- 5 (1993) Biotechnology, vol. 3, Chapter III: "Product recovery and purification", page 469-714, VCH: Weinheim; Belter, P.A. *et al.* (1988) Bioseparations: downstream processing for biotechnology, John Wiley and Sons; Kennedy, J.F. and Cabral, J.M.S. (1992) Recovery processes for biological materials, John Wiley and Sons; Shaeiwitz, J.A. and Henry, J.D. (1988) Biochemical separations, in: Ulmann's Encyclopedia of
- 10 Industrial Chemistry, vol. B3, Chapter 11, page 1-27, VCH: Weinheim; and Dechow, F.J. (1989) Separation and purification techniques in biotechnology, Noyes Publications.)

- In addition to the measurement of the final product of fermentation, it is also possible to analyze other components of the metabolic pathways utilized for the
- 15 production of the desired compound, such as intermediates and side-products, to determine the overall efficiency of production of the compound. Analysis methods include measurements of nutrient levels in the medium (e.g., sugars, hydrocarbons, nitrogen sources, phosphate, and other ions), measurements of biomass composition and growth, analysis of the production of common metabolites of biosynthetic pathways, and
- 20 measurement of gasses produced during fermentation. Standard methods for these measurements are outlined in Applied Microbial Physiology, A Practical Approach, P.M. Rhodes and P.F. Stanbury, eds., IRL Press, p. 103-129; 131-163; and 165-192 (ISBN: 0199635773) and references cited therein.

25 **Example 10: Purification of the Desired Product from *C. glutamicum* Culture**

- Recovery of the desired product from the *C. glutamicum* cells or supernatant of the above-described culture can be performed by various methods well known in the art. If the desired product is not secreted from the cells, the cells can be harvested from the culture by low-speed centrifugation, the cells can be lysed by standard techniques, such
- 30 as mechanical force or sonication. The cellular debris is removed by centrifugation, and the supernatant fraction containing the soluble proteins is retained for further purification of the desired compound. If the product is secreted from the *C. glutamicum*

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cells, then the cells are removed from the culture by low-speed centrifugation, and the supernate fraction is retained for further purification.

The supernatant fraction from either purification method is subjected to chromatography with a suitable resin, in which the desired molecule is either retained on a chromatography resin while many of the impurities in the sample are not, or where the impurities are retained by the resin while the sample is not. Such chromatography steps may be repeated as necessary, using the same or different chromatography resins. One of ordinary skill in the art would be well-versed in the selection of appropriate chromatography resins and in their most efficacious application for a particular molecule to be purified. The purified product may be concentrated by filtration or ultrafiltration, and stored at a temperature at which the stability of the product is maximized.

There are a wide array of purification methods known to the art and the preceding method of purification is not meant to be limiting. Such purification techniques are described, for example, in Bailey, J.E. & Ollis, D.F. *Biochemical Engineering Fundamentals*, McGraw-Hill: New York (1986).

The identity and purity of the isolated compounds may be assessed by techniques standard in the art. These include high-performance liquid chromatography (HPLC), spectroscopic methods, staining methods, thin layer chromatography, NIRS, enzymatic assay, or microbiologically. Such analysis methods are reviewed in: Patek *et al.* (1994) *Appl. Environ. Microbiol.* 60: 133-140; Malakhova *et al.* (1996) *Biotekhnologiya* 11: 27-32; and Schmidt *et al.* (1998) *Bioprocess Engineer.* 19: 67-70. *Ullmann's Encyclopedia of Industrial Chemistry*, (1996) vol. A27, VCH: Weinheim, p. 89-90, p. 521-540, p. 540-547, p. 559-566, 575-581 and p. 581-587; Michal, G. (1999) *Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology*, John Wiley and Sons; Fallon, A. *et al.* (1987) *Applications of HPLC in Biochemistry in: Laboratory Techniques in Biochemistry and Molecular Biology*, vol. 17.

Example 11: Analysis of the Gene Sequences of the Invention

The comparison of sequences and determination of percent homology between two sequences are art-known techniques, and can be accomplished using a mathematical algorithm, such as the algorithm of Karlin and Altschul (1990) *Proc. Natl. Acad. Sci. USA* 87:2264-68, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA*

90:5873-77. Such an algorithm is incorporated into the NBLAST and XBLAST programs (version 2.0) of Altschul, *et al.* (1990) *J. Mol. Biol.* 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences homologous to HA nucleic acid molecules of the invention. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength = 3 to obtain amino acid sequences homologous to HA protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul *et al.*, (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, one of ordinary skill in the art will know how to optimize the parameters of the program (*e.g.*, XBLAST and NBLAST) for the specific sequence being analyzed.

Another example of a mathematical algorithm utilized for the comparison of sequences is the algorithm of Meyers and Miller ((1988) *Comput. Appl. Biosci.* 4: 11-17). Such an algorithm is incorporated into the ALIGN program (version 2.0) which is part of the GCG sequence alignment software package. When utilizing the ALIGN program for comparing amino acid sequences, a PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used. Additional algorithms for sequence analysis are known in the art, and include ADVANCE and ADAM. described in Torelli and Robotti (1994) *Comput. Appl. Biosci.* 10:3-5; and FASTA, described in Pearson and Lipman (1988) *P.N.A.S.* 85:2444-8.

The percent homology between two amino acid sequences can also be accomplished using the GAP program in the GCG software package (available at <http://www.gcg.com>), using either a Blosum 62 matrix or a PAM250 matrix, and a gap weight of 12, 10, 8, 6, or 4 and a length weight of 2, 3, or 4. The percent homology between two nucleic acid sequences can be accomplished using the GAP program in the GCG software package, using standard parameters, such as a gap weight of 50 and a length weight of 3.

A comparative analysis of the gene sequences of the invention with those present in Genbank has been performed using techniques known in the art (see, *e.g.*, Bexevanis and Ouellette, eds. (1998) *Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins.* John Wiley and Sons: New York). The gene sequences of the invention

were compared to genes present in Genbank in a three-step process. In a first step, a BLASTN analysis (e.g., a local alignment analysis) was performed for each of the sequences of the invention against the nucleotide sequences present in Genbank, and the top 500 hits were retained for further analysis. A subsequent FASTA search (e.g., a
5 combined local and global alignment analysis, in which limited regions of the sequences are aligned) was performed on these 500 hits. Each gene sequence of the invention was subsequently globally aligned to each of the top three FASTA hits, using the GAP program in the GCG software package (using standard parameters). In order to obtain correct results, the length of the sequences extracted from Genbank were adjusted to the
10 length of the query sequences by methods well-known in the art. The results of this analysis are set forth in Table 4. The resulting data is identical to that which would have been obtained had a GAP (global) analysis alone been performed on each of the genes of the invention in comparison with each of the references in Genbank, but required significantly reduced computational time as compared to such a database-wide GAP
15 (global) analysis. Sequences of the invention for which no alignments above the cutoff values were obtained are indicated on Table 4 by the absence of alignment information. It will further be understood by one of ordinary skill in the art that the GAP alignment homology percentages set forth in Table 4 under the heading "% homology (GAP)" are listed in the European numerical format, wherein a ',' represents a decimal point. For
20 example, a value of "40,345" in this column represents "40.345%".

Example 12: Construction and Operation of DNA Microarrays

The sequences of the invention may additionally be used in the construction and application of DNA microarrays (the design, methodology, and uses of DNA arrays are
25 well known in the art, and are described, for example, in Schena, M. *et al.* (1995) *Science* 270: 467-470; Wodicka, L. *et al.* (1997) *Nature Biotechnology* 15: 1359-1367; DeSaizieu, A. *et al.* (1998) *Nature Biotechnology* 16: 45-48; and DeRisi, J.L. *et al.* (1997) *Science* 278: 680-686).

DNA microarrays are solid or flexible supports consisting of nitrocellulose,
30 nylon, glass, silicone, or other materials. Nucleic acid molecules may be attached to the surface in an ordered manner. After appropriate labeling, other nucleic acids or nucleic acid mixtures can be hybridized to the immobilized nucleic acid molecules, and the label

may be used to monitor and measure the individual signal intensities of the hybridized molecules at defined regions. This methodology allows the simultaneous quantification of the relative or absolute amount of all or selected nucleic acids in the applied nucleic acid sample or mixture. DNA microarrays, therefore, permit an analysis of the

5 expression of multiple (as many as 6800 or more) nucleic acids in parallel (see, *e.g.*, Schena, M. (1996) *BioEssays* 18(5): 427-431).

The sequences of the invention may be used to design oligonucleotide primers which are able to amplify defined regions of one or more *C. glutamicum* genes by a nucleic acid amplification reaction such as the polymerase chain reaction. The choice

10 and design of the 5' or 3' oligonucleotide primers or of appropriate linkers allows the covalent attachment of the resulting PCR products to the surface of a support medium described above (and also described, for example, Schena, M. *et al.* (1995) *Science* 270: 467-470).

Nucleic acid microarrays may also be constructed by *in situ* oligonucleotide

15 synthesis as described by Wodicka, L. *et al.* (1997) *Nature Biotechnology* 15: 1359-1367. By photolithographic methods, precisely defined regions of the matrix are exposed to light. Protective groups which are photolabile are thereby activated and undergo nucleotide addition, whereas regions that are masked from light do not undergo any modification. Subsequent cycles of protection and light activation permit the

20 synthesis of different oligonucleotides at defined positions. Small, defined regions of the genes of the invention may be synthesized on microarrays by solid phase oligonucleotide synthesis.

The nucleic acid molecules of the invention present in a sample or mixture of nucleotides may be hybridized to the microarrays. These nucleic acid molecules can be

25 labeled according to standard methods. In brief, nucleic acid molecules (*e.g.*, mRNA molecules or DNA molecules) are labeled by the incorporation of isotopically or fluorescently labeled nucleotides, *e.g.*, during reverse transcription or DNA synthesis. Hybridization of labeled nucleic acids to microarrays is described (*e.g.*, in Schena, M. *et al.* (1995) *supra*; Wodicka, L. *et al.* (1997), *supra*; and DeSaizieu A. *et al.* (1998),

30 *supra*). The detection and quantification of the hybridized molecule are tailored to the specific incorporated label. Radioactive labels can be detected, for example, as

described in Schena, M. *et al.* (1995) *supra* and fluorescent labels may be detected, for example, by the method of Shalon *et al.* (1996) *Genome Research* 6: 639-645).

The application of the sequences of the invention to DNA microarray technology, as described above, permits comparative analyses of different strains of *C. glutamicum* or other Corynebacteria. For example, studies of inter-strain variations based on individual transcript profiles and the identification of genes that are important for specific and/or desired strain properties such as pathogenicity, productivity and stress tolerance are facilitated by nucleic acid array methodologies. Also, comparisons of the profile of expression of genes of the invention during the course of a fermentation reaction are possible using nucleic acid array technology.

Example 13: Analysis of the Dynamics of Cellular Protein Populations (Proteomics)

The genes, compositions, and methods of the invention may be applied to study the interactions and dynamics of populations of proteins, termed 'proteomics'. Protein populations of interest include, but are not limited to, the total protein population of *C. glutamicum* (e.g., in comparison with the protein populations of other organisms), those proteins which are active under specific environmental or metabolic conditions (e.g., during fermentation, at high or low temperature, or at high or low pH), or those proteins which are active during specific phases of growth and development.

Protein populations can be analyzed by various well-known techniques, such as gel electrophoresis. Cellular proteins may be obtained, for example, by lysis or extraction, and may be separated from one another using a variety of electrophoretic techniques. Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) separates proteins largely on the basis of their molecular weight. Isoelectric focusing polyacrylamide gel electrophoresis (IEF-PAGE) separates proteins by their isoelectric point (which reflects not only the amino acid sequence but also posttranslational modifications of the protein). Another, more preferred method of protein analysis is the consecutive combination of both IEF-PAGE and SDS-PAGE, known as 2-D-gel electrophoresis (described, for example, in Hermann *et al.* (1998) *Electrophoresis* 19: 3217-3221; Fountoulakis *et al.* (1998) *Electrophoresis* 19: 1193-1202; Langen *et al.* (1997) *Electrophoresis* 18: 1184-1192; Antelmann *et al.* (1997) *Electrophoresis* 18:

1451-1463). Other separation techniques may also be utilized for protein separation, such as capillary gel electrophoresis; such techniques are well known in the art.

Proteins separated by these methodologies can be visualized by standard techniques, such as by staining or labeling. Suitable stains are known in the art, and include Coomassie Brilliant Blue, silver stain, or fluorescent dyes such as Sypro Ruby (Molecular Probes). The inclusion of radioactively labeled amino acids or other protein precursors (e.g., ^{35}S -methionine, ^{35}S -cysteine, ^{14}C -labelled amino acids, ^{15}N -amino acids, $^{15}\text{NO}_3$ or $^{15}\text{NH}_4^+$ or ^{13}C -labelled amino acids) in the medium of *C. glutamicum* permits the labeling of proteins from these cells prior to their separation. Similarly, fluorescent labels may be employed. These labeled proteins can be extracted, isolated and separated according to the previously described techniques.

Proteins visualized by these techniques can be further analyzed by measuring the amount of dye or label used. The amount of a given protein can be determined quantitatively using, for example, optical methods and can be compared to the amount of other proteins in the same gel or in other gels. Comparisons of proteins on gels can be made, for example, by optical comparison, by spectroscopy, by image scanning and analysis of gels, or through the use of photographic films and screens. Such techniques are well-known in the art.

To determine the identity of any given protein, direct sequencing or other standard techniques may be employed. For example, N- and/or C-terminal amino acid sequencing (such as Edman degradation) may be used, as may mass spectrometry (in particular MALDI or ESI techniques (see, e.g., Langen *et al.* (1997) *Electrophoresis* 18: 1184-1192)). The protein sequences provided herein can be used for the identification of *C. glutamicum* proteins by these techniques.

The information obtained by these methods can be used to compare patterns of protein presence, activity, or modification between different samples from various biological conditions (e.g., different organisms, time points of fermentation, media conditions, or different biotopes, among others). Data obtained from such experiments alone, or in combination with other techniques, can be used for various applications, such as to compare the behavior of various organisms in a given (e.g., metabolic) situation, to increase the productivity of strains which produce fine chemicals or to increase the efficiency of the production of fine chemicals.

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Equivalents

Those of ordinary skill in the art will recognize, or will be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the

5 following claims.

What is claimed:

1. An isolated nucleic acid molecule from *Corynebacterium glutamicum* encoding an HA protein, or a portion thereof, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.
2. The isolated nucleic acid molecule of claim 1, wherein said nucleic acid molecule encodes an HA protein involved in the production of a fine chemical.
3. An isolated *Corynebacterium glutamicum* nucleic acid molecule selected from the group consisting of those sequences set forth as odd-numbered SEQ ID NOs of the Sequence Listing, or a portion thereof, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.
4. An isolated nucleic acid molecule which encodes a polypeptide sequence selected from the group consisting of those sequences set forth as even-numbered SEQ ID NOs of the Sequence Listing, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.
5. An isolated nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide selected from the group of amino acid sequences consisting of those sequences set forth as even-numbered SEQ ID NOs of the Sequence Listing, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.
6. An isolated nucleic acid molecule comprising a nucleotide sequence which is at least 50% homologous to a nucleotide sequence selected from the group consisting of those sequences set forth as odd-numbered SEQ ID NOs of the Sequence Listing, or a portion thereof, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.

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7. An isolated nucleic acid molecule comprising a fragment of at least 15 nucleotides of a nucleic acid comprising a nucleotide sequence selected from the group consisting of those sequences set forth as odd-numbered SEQ ID NOs of the Sequence Listing, provided that the nucleic acid molecule does not consist of any of the F-designated genes set forth in Table 1.
8. An isolated nucleic acid molecule which hybridizes to the nucleic acid molecule of any one of claims 1-7 under stringent conditions.
9. An isolated nucleic acid molecule comprising the nucleic acid molecule of any one of claims 1-8 or a portion thereof and a nucleotide sequence encoding a heterologous polypeptide.
10. A vector comprising the nucleic acid molecule of any one of claims 1-9.
11. The vector of claim 10, which is an expression vector.
12. A host cell transfected with the expression vector of claim 11.
13. The host cell of claim 12, wherein said cell is a microorganism.
14. The host cell of claim 13, wherein said cell belongs to the genus *Corynebacterium* or *Brevibacterium*.
15. The host cell of claim 12, wherein the expression of said nucleic acid molecule results in the modulation in production of a fine chemical from said cell.
16. The host cell of claim 15, wherein said fine chemical is selected from the group consisting of: organic acids, proteinogenic and nonproteinogenic amino acids, purine and pyrimidine bases, nucleosides, nucleotides, lipids, saturated and unsaturated fatty acids, diols, carbohydrates, aromatic compounds, vitamins, cofactors, polyketides, and enzymes.

17. A method of producing a polypeptide comprising culturing the host cell of claim 12 in an appropriate culture medium to, thereby, produce the polypeptide.
- 5 18. An isolated HA polypeptide from *Corynebacterium glutamicum*, or a portion thereof.
19. The polypeptide of claim 18, wherein said polypeptide is involved in the production of a fine chemical production.
- 10 20. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of those sequences set forth as even-numbered SEQ ID NOs of the Sequence Listing, provided that the amino acid sequence is not encoded by any of the F-designated genes set forth in Table 1.
- 15 21. An isolated polypeptide comprising a naturally occurring allelic variant of a polypeptide comprising an amino acid sequence selected from the group consisting of those sequences set forth in as even-numbered SEQ ID NOs of the Sequence Listing, or a portion thereof, provided that the amino acid sequence is not encoded
- 20 by any of the F-designated genes set forth in Table 1.
22. The isolated polypeptide of any of claims 18-21, further comprising heterologous amino acid sequences.
- 25 23. An isolated polypeptide which is encoded by a nucleic acid molecule comprising a nucleotide sequence which is at least 50% homologous to a nucleic acid selected from the group consisting of those sequences set forth as odd-numbered SEQ ID NOs of the Sequence Listing, provided that the nucleic acid molecule does not consist of any of the F-designated nucleic acid molecules set forth in Table 1.
- 30 24. An isolated polypeptide comprising an amino acid sequence which is at least 50% homologous to an amino acid sequence selected from the group consisting of those

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sequences set forth as even-numbered SEQ ID NOs of the Sequence Listing, provided that the amino acid sequence is not encoded by any of the F-designated genes set forth in Table 1.

- 5 25. A method for producing a fine chemical, comprising culturing a cell containing a vector of claim 12 such that the fine chemical is produced.
26. The method of claim 25, wherein said method further comprises the step of recovering the fine chemical from said culture.
- 10 27. The method of claim 25, wherein said method further comprises the step of transfecting said cell with the vector of claim 11 to result in a cell containing said vector.
- 15 28. The method of claim 25, wherein said cell belongs to the genus *Corynebacterium* or *Brevibacterium*.
29. The method of claim 25, wherein said cell is selected from the group consisting of: *Corynebacterium glutamicum*, *Corynebacterium herculis*, *Corynebacterium lilium*,
20 *Corynebacterium acetoacidophilum*, *Corynebacterium acetoglutamicum*,
Corynebacterium acetophilum, *Corynebacterium ammoniagenes*, *Corynebacterium fujiokense*, *Corynebacterium nitrilophilus*, *Brevibacterium ammoniagenes*,
Brevibacterium butanicum, *Brevibacterium divaricatum*, *Brevibacterium flavum*,
Brevibacterium healii, *Brevibacterium ketoglutamicum*, *Brevibacterium*
25 *ketosoreductum*, *Brevibacterium lactofermentum*, *Brevibacterium linens*,
Brevibacterium paraffinolyticum, and those strains set forth in Table 3.
30. The method of claim 25, wherein expression of the nucleic acid molecule from said vector results in modulation of production of said fine chemical.
- 30 31. The method of claim 25, wherein said fine chemical is selected from the group consisting of: organic acids, proteinogenic and nonproteinogenic amino acids, purine

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and pyrimidine bases, nucleosides, nucleotides, lipids, saturated and unsaturated fatty acids, diols, carbohydrates, aromatic compounds, vitamins, cofactors, polyketides, and enzymes.

- 5 32. The method of claim 25, wherein said fine chemical is an amino acid.
33. The method of claim 32, wherein said amino acid is drawn from the group consisting of: lysine, glutamate, glutamine, alanine, aspartate, glycine, serine, threonine, methionine, cysteine, valine, leucine, isoleucine, arginine, proline, histidine, 10 tyrosine, phenylalanine, and tryptophan.
34. A method for producing a fine chemical, comprising culturing a cell whose genomic DNA has been altered by the inclusion of a nucleic acid molecule of any one of 15 claims 1-9.
35. A method for diagnosing the presence or activity of *Corynebacterium diphtheriae* in a subject, comprising detecting the presence of one or more of SEQ ID NOs 1 through 440 of the Sequence Listing in the subject, provided that the sequences are not or are not 20 encoded by any of the F-designated sequences set forth in Table 1, thereby diagnosing the presence or activity of *Corynebacterium diphtheriae* in the subject.
36. A host cell comprising a nucleic acid molecule selected from the group consisting of the nucleic acid molecules set forth as odd-numbered SEQ ID NOs of the Sequence Listing, wherein the nucleic acid molecule is disrupted.
- 25 37. A host cell comprising a nucleic acid molecule selected from the group consisting of the nucleic acid molecules set forth as odd-numbered SEQ ID NOs in the Sequence Listing, wherein the nucleic acid molecule comprises one or more nucleic acid modifications from the sequence set forth as odd-numbered SEQ ID NOs of the 30 Sequence Listing s.

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38. A host cell comprising a nucleic acid molecule selected from the group consisting of the nucleic acid molecules set forth as odd-numbered SEQ ID NOs of the Sequence Listing, wherein the regulatory region of the nucleic acid molecule is modified relative to the wild-type regulatory region of the molecule.

SEQUENCE LISTING

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 INVOLVED IN HOMEOSTASIS AND ADAPTATION

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acc ggt gct gtg ctc tcc gaa gcc cgc acc att gac gat gtg atc gaa      192
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gag atc gcc acc tcc acc ctt acc gaa cgt ggc gca acc cgc gcc gat      240
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ctg gca tct gca ctt gat gtc aca gac acc cac gtt gtt cca atc tct 240
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tac acc gga cca acc gtg ctg gaa atc ctg gaa aac gta gaa gtt tcc 336
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cac ggc cgt gca cac gac ctg ggc ttc cgc ttc cca atc cag tac gtc 384
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gaa gaa aac agc acg gat tat gtc ttc tac gca gtt cgc ctc ccc tac Glu Glu Asn Ser Thr Asp Tyr Val Phe Tyr Ala Val Arg Leu Pro Tyr 70 75 80 85			355
gcg atc cag gca gat gag gac gat gcg caa gtt gca ttg gaa ttc atc Ala Ile Gln Ala Asp Glu Asp Asp Ala Gln Val Ala Leu Glu Phe Ile 90 95 100			403
gca cct gac aag agc gtg acc gtc aac gtt aaa gac gca acg gac gcc Ala Pro Asp Lys Ser Val Thr Val Asn Val Lys Asp Ala Thr Asp Ala 105 110 115			451
acc gaa gca act gtt gca gct gct ttg gaa ctt cct gag ctg acc gac Thr Glu Ala Thr Val Ala Ala Leu Glu Leu Pro Glu Leu Thr Asp 120 125 130			499
ttc aat cgg ggc aat att aaa gct cgc caa cgc atg gtt gcc cag tac Phe Asn Arg Gly Asn Ile Lys Ala Arg Gln Arg Met Val Ala Gln Tyr 135 140 145			547
gca atc gca ggc cag ttg ggc ttg ctg gtt att ggc act gat cac gcg Ala Ile Ala Gly Gln Leu Gly Leu Leu Val Ile Gly Thr Asp His Ala 150 155 160 165			595
gct gaa aac gtc acg ggg ttc ttc acc aaa ttc ggt gat ggc gca gct Ala Glu Asn Val Thr Gly Phe Phe Thr Lys Phe Gly Asp Gly Ala Ala 170 175 180			643
gac ctg ctt cct ttg gca ggt ttg agc aag cgt caa gga gct gcc att Asp Leu Leu Pro Leu Ala Gly Leu Ser Lys Arg Gln Gly Ala Ala Ile 185 190 195			691
ctg gag cac ctg ggt gca cct tca agc acg tgg acc aag gtt cct acc Leu Glu His Leu Gly Ala Pro Ser Ser Thr Trp Thr Lys Val Pro Thr 200 205 210			739
gct gat ttg gaa gag gat cgc cca gcg ttg cca gat gag gaa gca ctt Ala Asp Leu Glu Glu Asp Arg Pro Ala Leu Pro Asp Glu Glu Ala Leu 215 220 225			787
ggt gtg tcg tat gcg gac atc gat aat tac ctg gaa aac aag ccc gat Gly Val Ser Tyr Ala Asp Ile Asp Asn Tyr Leu Glu Asn Lys Pro Asp 230 235 240 245			835
gtc agt gaa aaa gcc cag cag cgc att gag cac ctg tgg aag gtg ggc Val Ser Glu Lys Ala Gln Gln Arg Ile Glu His Leu Trp Lys Val Gly 250 255 260			883
cag cac aag cgc cac ctc cct gct acc ccg cag gaa aat tgg tgg cgt Gln His Lys Arg His Leu Pro Ala Thr Pro Gln Glu Asn Trp Trp Arg 265 270 275			931
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 35 40 45
 Gly Gly Gln Asp Ser Thr Leu Ala Gly Arg Leu Thr Gln Leu Ala Val
 50 55 60
 Glu Arg Ile Arg Ala Glu Asn Ser Thr Asp Tyr Val Phe Tyr Ala
 65 70 75 80
 Val Arg Leu Pro Tyr Ala Ile Gln Ala Asp Glu Asp Asp Ala Gln Val
 85 90 95
 Ala Leu Glu Phe Ile Ala Pro Asp Lys Ser Val Thr Val Asn Val Lys
 100 105 110
 Asp Ala Thr Asp Ala Thr Glu Ala Thr Val Ala Ala Leu Glu Leu
 115 120 125
 Pro Glu Leu Thr Asp Phe Asn Arg Gly Asn Ile Lys Ala Arg Gln Arg
 130 135 140
 Met Val Ala Gln Tyr Ala Ile Ala Gly Gln Leu Gly Leu Leu Val Ile
 145 150 155 160
 Gly Thr Asp His Ala Ala Glu Asn Val Thr Gly Phe Phe Thr Lys Phe
 165 170 175
 Gly Asp Gly Ala Ala Asp Leu Leu Pro Leu Ala Gly Leu Ser Lys Arg
 180 185 190
 Gln Gly Ala Ala Ile Leu Glu His Leu Gly Ala Pro Ser Ser Thr Trp
 195 200 205
 Thr Lys Val Pro Thr Ala Asp Leu Glu Glu Asp Arg Pro Ala Leu Pro
 210 215 220
 Asp Glu Glu Ala Leu Gly Val Ser Tyr Ala Asp Ile Asp Asn Tyr Leu
 225 230 235 240
 Glu Asn Lys Pro Asp Val Ser Glu Lys Ala Gln Gln Arg Ile Glu His
 245 250 255
 Leu Trp Lys Val Gly Gln His Lys Arg His Leu Pro Ala Thr Pro Gln
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Val Gly Arg Glu Ala Lys Thr Ile Glu Ile Ile Asn Thr Gly Asp Arg
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Pro Val Gln Ile Gly Ser His Phe His Phe Ala Glu Val Asn Pro Ser
      35           40           45
Ile Ser Phe Asp Arg Ser Glu Gly Tyr Gly Phe Arg Leu Asp Ile Pro
      50           55           60
Ser Gly Thr Ala Val Arg Leu Glu Pro Gly Asp Ala Arg Thr Val Asn
      65           70           75           80
Leu Val Ala Ile Gly Gly Asp Arg Ile Val Ala Gly Phe Arg Asp Leu
          85           90           95
Val Asp Gly Pro Leu Glu Asp Leu Lys Val Asn Val Trp Glu Gly Arg
      100           105           110
Glu Asp Gly Trp Arg Arg Ser Ser Ala Ala Gly Asp Ala Pro Gln Glu
      115           120           125
Leu Pro Gln Val Glu Ala Ala Glu Arg Gly Arg Lys Leu Asp Asp Ala
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Thr Asp Val Asp Thr Asn Val Gly Thr Glu Glu Gly Phe Glu Glu Gly
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Arg Asn
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<223> FRXA02264

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atcccatccg ataacccttg atgttttttag gagttttgtc atg atc cca ggc gag 115
                                     Met Ile Pro Gly Glu
                                     1           5
tac atc ctg tcc agc gaa tca ctc acc gga aat gtt ggg cgc gag gcc 163
Tyr Ile Leu Ser Ser Glu Ser Leu Thr Gly Asn Val Gly Arg Glu Ala
      10           15           20
aaa acc atc gaa atc atc aac acc ggt gat agg cct gtg cag att ggt 211
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Lys Thr Ile Glu Ile Ile Asn Thr Gly Asp Arg Pro Val Gln Ile Gly
 25 30 35

tcg cat ttc
 Ser His Phe
 40

220

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 <213> Corynebacterium glutamicum

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 Pro Val Gln Ile Gly Ser His Phe
 35 40

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 Met Ser Phe Glu Ile
 1 5
 tcc cgc aag cag tac acc gac ctt tat ggt cca acc gtt ggc gat tca 163
 Ser Arg Lys Gln Tyr Thr Asp Leu Tyr Gly Pro Thr Val Gly Asp Ser
 10 15 20
 gta cgt ctt gct gat act gag ctt ttt ctc tgt gtg gaa aaa gat tac 211
 Val Arg Leu Ala Asp Thr Glu Leu Phe Leu Cys Val Glu Lys Asp Tyr
 25 30 35
 gca gca atc ggc gaa gaa gta gca ttc ggc ggt ggc aag gtc att cgt 259
 Ala Ala Ile Gly Glu Glu Val Ala Phe Gly Gly Gly Lys Val Ile Arg
 40 45 50
 gat ggc atg ggc caa aat ggc acc ttg gtt cgc gat gta gat att ccc 307
 Asp Gly Met Gly Gln Asn Gly Thr Leu Val Arg Asp Val Asp Ile Pro
 55 60 65
 gat acc gtc atc acc aac gtc atc gtc ctt gac tat acg ggt gtg tac 355
 Asp Thr Val Ile Thr Asn Val Ile Val Leu Asp Tyr Thr Gly Val Tyr
 70 75 80 85

aaa gct gac gtt gcg ctt cga gat ggc aaa atc ttc cga atc gga aag	403
Lys Ala Asp Val Ala Leu Arg Asp Gly Lys Ile Phe Arg Ile Gly Lys	
90 95 100	
gcc gga aac ccg aat gtc atg gaa aac gtc gac atc gtc atc ggc gtt	451
Ala Gly Asn Pro Asn Val Met Glu Asn Val Asp Ile Val Ile Gly Val	
105 110 115	
gcc acc gac atc att gct ggt gaa ggc aaa atc ctt acc gca ggt ggc	499
Ala Thr Asp Ile Ile Ala Gly Glu Gly Lys Ile Leu Thr Ala Gly Gly	
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Ile Asp Thr His Val His Phe Leu Gly Thr Asp Gln Val Asn Thr Ala	
135 140 145	
tta gca tca ggt atc acc acg atg atc ggt gga ggc acc ggc cca agc	595
Leu Ala Ser Gly Ile Thr Thr Met Ile Gly Gly Thr Gly Pro Ser	
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cag gcg tcg atg gct aca act gtc acg cca ggt cag tgg aat acc tac	643
Gln Ala Ser Met Ala Thr Thr Val Thr Pro Gly Gln Trp Asn Thr Tyr	
170 175 180	
aac atg ctt agt gct ttt gaa ggc atg ccc atg aac ttt ggc att ttg	691
Asn Met Leu Ser Ala Phe Glu Gly Met Pro Met Asn Phe Gly Ile Leu	
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Gly Lys Gly His Gly Ser Ser Lys Ser Pro Leu Ala Glu Gln Val Arg	
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gcg ggt gca atc ggt ctg aaa att cac gag gac tgg ggt gcc aca cca	787
Ala Gly Ala Ile Gly Leu Lys Ile His Glu Asp Trp Gly Ala Thr Pro	
215 220 225	
tcg tcg atc aac act gcc cta gaa gta gcc gat gac atg gac atc cag	835
Ser Ser Ile Asn Thr Ala Leu Glu Val Ala Asp Asp Met Asp Ile Gln	
230 235 240 245	
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Val Ala Leu His Ser Asp Thr Leu Asn Glu Ala Gly Phe Val Glu Asp	
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acc att gaa gcc att gcg ggc cga gtc atc cat acc ttc cac acc gaa	931
Thr Ile Glu Ala Ile Ala Gly Arg Val Ile His Thr Phe His Thr Glu	
265 270 275	
ggt gct ggt ggt gga cac gct cct gac cta atc cga gtg gct gct ctg	979
Gly Ala Gly Gly His Ala Pro Asp Leu Ile Arg Val Ala Ala Leu	
280 285 290	
cca aac gtg ttg cct gca tcc acc aac cca acg ctc cca tac acc cga	1027
Pro Asn Val Leu Pro Ala Ser Thr Asn Pro Thr Leu Pro Tyr Thr Arg	
295 300 305	
aac act gtt gaa gag cac ctg gac atg gtg atg gtt gcc cac cac ctc	1075
Asn Thr Val Glu Glu His Leu Asp Met Val Met Val Ala His His Leu	
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gcc	gaa	acg	att	gca	gcc	gaa	gat	gtg	ctt	cac	gat	atg	ggt	atc	ttc	1171	
Ala	Glu	Thr	Ile	Ala	Ala	Glu	Asp	Val	Leu	His	Asp	Met	Gly	Ile	Phe		
			345					350					355				
tct	atc	acc	tct	tcg	gat	tcc	cag	gcg	atg	ggc	cga	gta	gga	gag	acc	1219	
Ser	Ile	Thr	Ser	Ser	Asp	Ser	Gln	Ala	Met	Gly	Arg	Val	Gly	Glu	Thr		
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atc	acg	cgc	acg	tgg	cag	gtc	gcc	gac	cat	atg	aaa	cgc	acc	cgt	gga	1267	
Ile	Thr	Arg	Thr	Trp	Gln	Val	Ala	Asp	His	Met	Lys	Arg	Thr	Arg	Gly		
			375			380					385						
tca	cta	acg	gga	gat	gct	cca	tac	aac	gac	aac	aac	cgc	ttg	cgt	cga	1315	
Ser	Leu	Thr	Gly	Asp	Ala	Pro	Tyr	Asn	Asp	Asn	Asn	Arg	Leu	Arg	Arg		
390					395					400					405		
ttc	atc	gca	aaa	tac	acc	atc	aac	cct	gcg	att	gcg	cac	ggt	gtg	gat	1363	
Phe	Ile	Ala	Lys	Tyr	Thr	Ile	Asn	Pro	Ala	Ile	Ala	His	Gly	Val	Asp		
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tat	gtt	gtt	cgt	tca	gtg	gag	gaa	ggc	aag	ttc	gct	gac	ctc	gtg	ctg	1411	
Tyr	Val	Val	Arg	Ser	Val	Glu	Glu	Gly	Lys	Phe	Ala	Asp	Leu	Val	Leu		
			425					430					435				
tgg	gat	cca	aag	ttc	ttt	ggt	gtg	aaa	cct	gat	ctg	gtg	atc	aag	ggt	1459	
Trp	Asp	Pro	Lys	Phe	Phe	Gly	Val	Lys	Pro	Asp	Leu	Val	Ile	Lys	Gly		
			440			445						450					
ggg	ttg	atg	gtc	aat	tcc	ctc	atg	ggt	gat	tcc	aac	ggt	tcc	att	cca	1507	
Gly	Leu	Met	Val	Asn	Ser	Leu	Met	Gly	Asp	Ser	Asn	Gly	Ser	Ile	Pro		
			455			460					465						
act	ccg	cag	ccc	cgc	acc	ctg	cgc	aat	act	tgg	ggt	gcg	ttt	ggc	cag	1555	
Thr	Pro	Gln	Pro	Arg	Thr	Leu	Arg	Asn	Thr	Trp	Gly	Ala	Phe	Gly	Gln		
					475					480					485		
gca	gtt	tcc	aga	agc	tcc	att	aca	ttc	cta	tcc	cag	gac	gct	atc	gat	1603	
Ala	Val	Ser	Arg	Ser	Ser	Ile	Thr	Phe	Leu	Ser	Gln	Asp	Ala	Ile	Asp		
				490					495					500			
gca	aat	gtt	cct	gat	ctg	ctg	aat	ctg	agg	aag	cag	atc	cgg	ggc	gtt	1651	
Ala	Asn	Val	Pro	Asp	Leu	Leu	Asn	Leu	Arg	Lys	Gln	Ile	Arg	Gly	Val		
			505					510					515				
cga	ggt	gta	agg	aat	ctg	acc	aaa	cga	gac	atg	aaa	ctc	aat	gca	gaa	1699	
Arg	Gly	Val	Arg	Asn	Leu	Thr	Lys	Arg	Asp	Met	Lys	Asn	Ala	Glu			
			520				525					530					
atg	cct	gat	att	cgt	gtc	gat	cca	gag	acc	tac	cag	gtg	ttt	gtc	aac	1747	
Met	Pro	Asp	Ile	Arg	Val	Asp	Pro	Glu	Thr	Tyr	Gln	Val	Phe	Val	Asn		
						540					545						
ggt	gag	ttg	atc	acc	agc	aag	cca	gca	gag	aca	gtg	cca	atg	gca	cgt	1795	
Gly	Glu	Leu	Ile	Thr	Ser	Lys	Pro	Ala	Glu	Thr	Val	Pro	Met	Ala	Arg		
					555					560					565		
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 <213> *Corynebacterium glutamicum*

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Val Glu Lys Asp Tyr Ala Ala Ile Gly Glu Glu Val Ala Phe Gly Gly
      35           40           45

Gly Lys Val Ile Arg Asp Gly Met Gly Gln Asn Gly Thr Leu Val Arg
      50           55           60

Asp Val Asp Ile Pro Asp Thr Val Ile Thr Asn Val Ile Val Leu Asp
      65           70           75           80

Tyr Thr Gly Val Tyr Lys Ala Asp Val Ala Leu Arg Asp Gly Lys Ile
          85           90           95

Phe Arg Ile Gly Lys Ala Gly Asn Pro Asn Val Met Glu Asn Val Asp
          100          105          110

Ile Val Ile Gly Val Ala Thr Asp Ile Ile Ala Gly Glu Gly Lys Ile
          115          120          125

Leu Thr Ala Gly Gly Ile Asp Thr His Val His Phe Leu Gly Thr Asp
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Gln Val Asn Thr Ala Leu Ala Ser Gly Ile Thr Thr Met Ile Gly Gly
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Gly Thr Gly Pro Ser Gln Ala Ser Met Ala Thr Thr Val Thr Pro Gly
          165          170          175

Gln Trp Asn Thr Tyr Asn Met Leu Ser Ala Phe Glu Gly Met Pro Met
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Asn Phe Gly Ile Leu Gly Lys Gly His Gly Ser Ser Lys Ser Pro Leu
          195          200          205

Ala Glu Gln Val Arg Ala Gly Ala Ile Gly Leu Lys Ile His Glu Asp
      210          215          220

Trp Gly Ala Thr Pro Ser Ser Ile Asn Thr Ala Leu Glu Val Ala Asp
      225          230          235          240

Asp Met Asp Ile Gln Val Ala Leu His Ser Asp Thr Leu Asn Glu Ala
          245          250          255

Gly Phe Val Glu Asp Thr Ile Glu Ala Ile Ala Gly Arg Val Ile His
          260          265          270

Thr Phe His Thr Glu Gly Ala Gly Gly Gly His Ala Pro Asp Leu Ile

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Leu Pro Tyr Thr Arg Asn Thr Val Glu Glu His Leu Asp Met Val Met		
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Val Ala His His Leu Asn Pro Asp Ile Pro Glu Asp Val Ala Phe Ala		
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Asp Ser Arg Ile Arg Ala Glu Thr Ile Ala Ala Glu Asp Val Leu His		
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Asp Met Gly Ile Phe Ser Ile Thr Ser Ser Asp Ser Gln Ala Met Gly		
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Arg Val Gly Glu Thr Ile Thr Arg Thr Trp Gln Val Ala Asp His Met		
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Lys Arg Thr Arg Gly Ser Leu Thr Gly Asp Ala Pro Tyr Asn Asp Asn		
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Asn Arg Leu Arg Arg Phe Ile Ala Lys Tyr Thr Ile Asn Pro Ala Ile		
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Ala His Gly Val Asp Tyr Val Val Arg Ser Val Glu Glu Gly Lys Phe		
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Ala Asp Leu Val Leu Trp Asp Pro Lys Phe Phe Gly Val Lys Pro Asp		
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Leu Val Ile Lys Gly Gly Leu Met Val Asn Ser Leu Met Gly Asp Ser		
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Asn Gly Ser Ile Pro Thr Pro Gln Pro Arg Thr Leu Arg Asn Thr Trp		
465	470	475
Gly Ala Phe Gly Gln Ala Val Ser Arg Ser Ser Ile Thr Phe Leu Ser		
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Gln Asp Ala Ile Asp Ala Asn Val Pro Asp Leu Leu Asn Leu Arg Lys		
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Gln Ile Arg Gly Val Arg Gly Val Arg Asn Leu Thr Lys Arg Asp Met		
	515	520
Lys Leu Asn Ala Glu Met Pro Asp Ile Arg Val Asp Pro Glu Thr Tyr		
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Gln Val Phe Val Asn Gly Glu Leu Ile Thr Ser Lys Pro Ala Glu Thr		
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Val Pro Met Ala Arg Arg Tyr Phe Leu Phe		
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<222> (1)..(1602)

<223> FRXA02274

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cgt gat ggc atg ggc caa aat ggc acc ttg gtt cgc gat gta gat att	96
Arg Asp Gly Met Gly Gln Asn Gly Thr Leu Val Arg Asp Val Asp Ile	
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ccc gat acc gtc atc acc aac gtc atc gtc ctt gac tat acg ggt gtg	144
Pro Asp Thr Val Ile Thr Asn Val Ile Val Leu Asp Tyr Thr Gly Val	
35 40 45	
tac aaa gct gac gtt gcg ctt cga gat ggc aaa atc ttc cga atc gga	192
Tyr Lys Ala Asp Val Ala Leu Arg Asp Gly Lys Ile Phe Arg Ile Gly	
50 55 60	
aag gcc gga aac cgc aat gtc atg gaa aac gtc gac atc gtc atc ggc	240
Lys Ala Gly Asn Pro Asn Val Met Glu Asn Val Asp Ile Val Ile Gly	
65 70 75 80	
gtt gcc acc gac atc att gct ggt gaa ggc aaa atc ctt acc gca ggt	288
Val Ala Thr Asp Ile Ile Ala Gly Glu Gly Lys Ile Leu Thr Ala Gly	
85 90 95	
ggc atc gac acg cac gtg cac ttc ttg ggc aca gac cag gtc aac act	336
Gly Ile Asp Thr His Val His Phe Leu Gly Thr Asp Gln Val Asn Thr	
100 105 110	
gca tta gca tca ggt atc acc acg atg atc ggt gga ggc acc ggc cca	384
Ala Leu Ala Ser Gly Ile Thr Thr Met Ile Gly Gly Gly Thr Gly Pro	
115 120 125	
agc cag gcg tcg atg gct aca act gtc acg cca ggt cag tgg aat acc	432
Ser Gln Ala Ser Met Ala Thr Thr Val Thr Pro Gly Gln Trp Asn Thr	
130 135 140	
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Tyr Asn Met Leu Ser Ala Phe Glu Gly Met Pro Met Asn Phe Gly Ile	
145 150 155 160	
ttg ggt aaa ggc cat ggt tct tcc aaa tct cgc ctg gct gag cag gtt	528
Leu Gly Lys Gly His Gly Ser Ser Lys Ser Pro Leu Ala Glu Gln Val	
165 170 175	
cgt gcg ggt gca atc ggt ctg aaa att cac gag gac tgg ggt gcc aca	576
Arg Ala Gly Ala Ile Gly Leu Lys Ile His Glu Asp Trp Gly Ala Thr	
180 185 190	
cca tcg tcg atc aac act gcc cta gaa gta gcc gat gac atg gac atc	624
Pro Ser Ser Ile Asn Thr Ala Leu Glu Val Ala Asp Asp Met Asp Ile	
195 200 205	
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Gln Val Ala Leu His Ser Asp Thr Leu Asn Glu Ala Gly Phe Val Glu	

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ctg cca aac gtg ttg cct gca tcc acc aac cca acg ctc cca tac acc Leu Pro Asn Val Leu Pro Ala Ser Thr Asn Pro Thr Leu Pro Tyr Thr 260 265 270			816
cga aac act gtt gaa gag cac ctg gac atg gtg atg gtt gcc cac cac Arg Asn Thr Val Glu Glu His Leu Asp Met Val Met Val Ala His His 275 280 285			864
ctc aac cca gat att cca gaa gac gtg gct ttt gcg gat tcc cga att Leu Asn Pro Asp Ile Pro Glu Asp Val Ala Phe Ala Asp Ser Arg Ile 290 295 300			912
cgt gcc gaa acg att gca gcc gaa gat gtg ctt cac gat atg ggt atc Arg Ala Glu Thr Ile Ala Ala Glu Asp Val Leu His Asp Met Gly Ile 305 310 315			960
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gga tca cta acg gga gat gct cca tac aac gac aac aac cgc ttg cgt Gly Ser Leu Thr Gly Asp Ala Pro Tyr Asn Asp Asn Asn Arg Leu Arg 355 360 365			1104
cga ttc atc gca aaa tac acc atc aac cct gcg att gcg cac ggt gtg Arg Phe Ile Ala Lys Tyr Thr Ile Asn Pro Ala Ile Ala His Gly Val 370 375 380			1152
gat tat gtt gtt cgt tca gtg gag gaa ggc aag ttc gct gac ctc gtg Asp Tyr Val Val Arg Ser Val Glu Glu Gly Lys Phe Ala Asp Leu Val 385 390 395 400			1200
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 Asp Ala Asn Val Pro Asp Leu Leu Asn Leu Arg Lys Gln Ile Arg Gly 480
 465 470 475

gtt cga ggt gta agg aat ctg acc aaa cga gac atg aaa ctc aat gca 1488
 Val Arg Gly Val Arg Asn Leu Thr Lys Arg Asp Met Lys Leu Asn Ala 495
 485 490

gaa atg cct gat att cgt gtc gat cca gag acc tac cag gtg ttt gtc 1536
 Glu Met Pro Asp Ile Arg Val Asp Pro Glu Thr Tyr Gln Val Phe Val 510
 500 505 510

aac ggt gag ttg atc acc agc aag cca gca gag aca gtg cca atg gca 1584
 Asn Gly Glu Leu Ile Thr Ser Lys Pro Ala Glu Thr Val Pro Met Ala 525
 515 520

cgt cgc tac ttc ttg ttc taatccgccca acaaggaagg aag 1625
 Arg Arg Tyr Phe Leu Phe 530

<210> 16

<211> 534

<212> PRT

<213> Corynebacterium glutamicum

<400> 16

Tyr Ala Ala Ile Gly Glu Glu Val Ala Phe Gly Gly Gly Lys Val Ile
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Arg Asp Gly Met Gly Gln Asn Gly Thr Leu Val Arg Asp Val Asp Ile
 20 25 30

Pro Asp Thr Val Ile Thr Asn Val Ile Val Leu Asp Tyr Thr Gly Val
 35 40 45

Tyr Lys Ala Asp Val Ala Leu Arg Asp Gly Lys Ile Phe Arg Ile Gly
 50 55 60

Lys Ala Gly Asn Pro Asn Val Met Glu Asn Val Asp Ile Val Ile Gly
 65 70 75 80

Val Ala Thr Asp Ile Ile Ala Gly Glu Gly Lys Ile Leu Thr Ala Gly
 85 90 95

Gly Ile Asp Thr His Val His Phe Leu Gly Thr Asp Gln Val Asn Thr
 100 105 110

Ala Leu Ala Ser Gly Ile Thr Thr Met Ile Gly Gly Gly Thr Gly Pro
 115 120 125

Ser Gln Ala Ser Met Ala Thr Thr Val Thr Pro Gly Gln Trp Asn Thr
 130 135 140

Tyr Asn Met Leu Ser Ala Phe Glu Gly Met Pro Met Asn Phe Gly Ile
 145 150 155 160

Leu Gly Lys Gly His Gly Ser Ser Lys Ser Pro Leu Ala Glu Gln Val
 165 170 175

Arg Ala Gly Ala Ile Gly Leu Lys Ile His Glu Asp Trp Gly Ala Thr
 180 185 190
 Pro Ser Ser Ile Asn Thr Ala Leu Glu Val Ala Asp Asp Met Asp Ile
 195 200 205
 Gln Val Ala Leu His Ser Asp Thr Leu Asn Glu Ala Gly Phe Val Glu
 210 215 220
 Asp Thr Ile Glu Ala Ile Ala Gly Arg Val Ile His Thr Phe His Thr
 225 230 235 240
 Glu Gly Ala Gly Gly Gly His Ala Pro Asp Leu Ile Arg Val Ala Ala
 245 250 255
 Leu Pro Asn Val Leu Pro Ala Ser Thr Asn Pro Thr Leu Pro Tyr Thr
 260 265 270
 Arg Asn Thr Val Glu Glu His Leu Asp Met Val Met Val Ala His His
 275 280 285
 Leu Asn Pro Asp Ile Pro Glu Asp Val Ala Phe Ala Asp Ser Arg Ile
 290 295 300
 Arg Ala Glu Thr Ile Ala Ala Glu Asp Val Leu His Asp Met Gly Ile
 305 310 315 320
 Phe Ser Ile Thr Ser Ser Asp Ser Gln Ala Met Gly Arg Val Gly Glu
 325 330 335
 Thr Ile Thr Arg Thr Trp Gln Val Ala Asp His Met Lys Arg Thr Arg
 340 345 350
 Gly Ser Leu Thr Gly Asp Ala Pro Tyr Asn Asp Asn Asn Arg Leu Arg
 355 360 365
 Arg Phe Ile Ala Lys Tyr Thr Ile Asn Pro Ala Ile Ala His Gly Val
 370 375 380
 Asp Tyr Val Val Arg Ser Val Glu Glu Gly Lys Phe Ala Asp Leu Val
 385 390 395 400
 Leu Trp Asp Pro Lys Phe Phe Gly Val Lys Pro Asp Leu Val Ile Lys
 405 410 415
 Gly Gly Leu Met Val Asn Ser Leu Met Gly Asp Ser Asn Gly Ser Ile
 420 425 430
 Pro Thr Pro Gln Pro Arg Thr Leu Arg Asn Thr Trp Gly Ala Phe Gly
 435 440 445
 Gln Ala Val Ser Arg Ser Ser Ile Thr Phe Leu Ser Gln Asp Ala Ile
 450 455 460
 Asp Ala Asn Val Pro Asp Leu Leu Asn Leu Arg Lys Gln Ile Arg Gly
 465 470 475 480
 Val Arg Gly Val Arg Asn Leu Thr Lys Arg Asp Met Lys Leu Asn Ala
 485 490 495
 Glu Met Pro Asp Ile Arg Val Asp Pro Glu Thr Tyr Gln Val Phe Val

Ala Asp Leu Ala Arg Arg Arg Lys Asp Arg Gly Leu Lys Leu Asn His
 20 25 30
 Pro Glu Ala Val Ala Leu Ile Thr Tyr Glu Leu Ile Glu Gly Ala Arg
 35 40 45
 Asp Gly Arg Thr Val Ala Asp Leu Met Ser Trp Gly Ser Thr Ile Leu
 50 55 60
 Thr Arg Asp Asp Val Leu Glu Gly Ile Pro Glu Met Ile Pro Asp Ile
 65 70 75 80
 Gln Val Glu Ala Thr Phe Asp Asp Gly Thr Lys Leu Val Thr Val His
 85 90 95
 Asn Pro Ile Arg
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<210> 19
 <211> 972
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
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 <222> (101)..(949)
 <223> RXA02278

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 Met Thr Gln Thr Gln
 1 5
 cca gtg gga acc ctg cga ctg acc atc gat gat caa gga ccc caa ggt 163
 Pro Val Gly Thr Leu Arg Leu Thr Ile Asp Asp Gln Gly Pro Gln Gly
 10 15 20
 caa agc cgt gcg gtg gag caa ttt cac cag ggt gcg ctt cga gtc atc 211
 Gln Ser Arg Ala Val Glu Gln Phe His Gln Gly Ala Leu Arg Val Ile
 25 30 35
 cgg cca cac tac ttg gat gat tcc gga cag gtt tgc tac acc atc att 259
 Arg Pro His Tyr Leu Asp Asp Ser Gly Gln Val Cys Tyr Thr Ile Ile
 40 45 50
 gcc att ggt ggc gga tac ctg ggc ggc gat gtg tat gag cag caa ttc 307
 Ala Ile Gly Gly Gly Tyr Leu Gly Gly Asp Val Tyr Glu Gln Gln Phe
 55 60 65
 acg atc aaa gac aac gca aaa gct ttg atc acc acg caa tgc gcc acc 355
 Thr Ile Lys Asp Asn Ala Lys Ala Leu Ile Thr Thr Gln Ser Ala Thr
 70 75 80 85
 aag att tat cgc aca ccg caa gga cca gcc acg cag cac acc gaa atc 403
 Lys Ile Tyr Arg Thr Pro Gln Gly Pro Ala Thr Gln His Thr Glu Ile
 90 95 100

aac gtc ggt gaa aat gct gtg ctg gaa tac ttg gcg gat caa acc atc 451
 Asn Val Gly Glu Asn Ala Val Leu Glu Tyr Leu Ala Asp Gln Thr Ile
 105 110 115
 gcg tac cgg gag gcc acc tat cat caa ttc acc aag gtg gcg ctg cac 499
 Ala Tyr Arg Glu Ala Thr Tyr His Gln Phe Thr Lys Val Ala Leu His
 120 125 130
 ccg agc gca acg ttt gtg atg agc gaa caa atc acc cca ggc tgg cac 547
 Pro Ser Ala Thr Phe Val Met Ser Glu Gln Ile Thr Pro Gly Trp His
 135 140 145
 ccc gac ggc aaa cac ttt gct tac gat gaa atg cgt cta cac acc gaa 595
 Pro Asp Gly Lys His Phe Ala Tyr Asp Glu Met Arg Leu His Thr Glu
 150 155 160 165
 atc acg gac tcc acc aca ggg cga ctc gtg ctc ttg gat aat tta ctg 643
 Ile Thr Asp Ser Thr Thr Gly Arg Leu Val Leu Leu Asp Asn Leu Leu
 170 175 180
 ctc cgg ccg gac tcc cga gag gga agt ttt ggg tgg acg gaa cag tac 691
 Leu Arg Pro Ser Arg Glu Gly Ser Phe Gly Trp Thr Glu Gln Tyr
 185 190 195
 aca cat tca ggg cag atg att gtg atg ggg gaa ggc gtc gat aag cag 739
 Thr His Ser Gly Gln Met Ile Val Met Gly Glu Gly Val Asp Lys Gln
 200 205 210
 ctt gtt gct gag ctg aat gag caa ctt gcc gcg cac cct gat gtg tac 787
 Leu Val Ala Glu Leu Asn Glu Gln Leu Ala Ala His Pro Asp Val Tyr
 215 220 225
 ggc gcc gtc aat ttc tta agc gcg ccg ggc acg tta ctg cgc gga ttt 835
 Gly Ala Val Asn Phe Leu Ser Ala Pro Gly Thr Leu Leu Arg Gly Phe
 230 235 240 245
 att gcg cgc acg ctg agc aac cgc act gag gag ttg att aac ctg cac 883
 Ile Ala Arg Thr Leu Ser Asn Arg Thr Glu Glu Leu Ile Asn Leu His
 250 255 260
 gaa cac att gcg tcg ctg ttg cgc ggg ccg tgg cgc ggg cag gaa ccg 931
 Glu His Ile Ala Ser Leu Leu Arg Gly Arg Trp Arg Gly Gln Glu Pro
 265 270 275
 gtg aat ttg ccg aag tac tagacggcgt cgagaaatcg aag 972
 Val Asn Leu Arg Lys Tyr
 280

<210> 20

<211> 283

<212> PRT

<213> Corynebacterium glutamicum

<400> 20

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Gln Gly Pro Gln Gly Gln Ser Arg Ala Val Glu Gln Phe His Gln Gly
 20 25 30

Ala Leu Arg Val Ile Arg Pro His Tyr Leu Asp Asp Ser Gly Gln Val
 35 40 45
 Cys Tyr Thr Ile Ile Ala Ile Gly Gly Gly Tyr Leu Gly Gly Asp Val
 50 55 60
 Tyr Glu Gln Gln Phe Thr Ile Lys Asp Asn Ala Lys Ala Leu Ile Thr
 65 70 75 80
 Thr Gln Ser Ala Thr Lys Ile Tyr Arg Thr Pro Gln Gly Pro Ala Thr
 85 90 95
 Gln His Thr Glu Ile Asn Val Gly Glu Asn Ala Val Leu Glu Tyr Leu
 100 105 110
 Ala Asp Gln Thr Ile Ala Tyr Arg Glu Ala Thr Tyr His Gln Phe Thr
 115 120 125
 Lys Val Ala Leu His Pro Ser Ala Thr Phe Val Met Ser Glu Gln Ile
 130 135 140
 Thr Pro Gly Trp His Pro Asp Gly Lys His Phe Ala Tyr Asp Glu Met
 145 150 155 160
 Arg Leu His Thr Glu Ile Thr Asp Ser Thr Thr Gly Arg Leu Val Leu
 165 170 175
 Leu Asp Asn Leu Leu Leu Arg Pro Asp Ser Arg Glu Gly Ser Phe Gly
 180 185 190
 Trp Thr Glu Gln Tyr Thr His Ser Gly Gln Met Ile Val Met Gly Glu
 195 200 205
 Gly Val Asp Lys Gln Leu Val Ala Glu Leu Asn Glu Gln Leu Ala Ala
 210 215 220
 His Pro Asp Val Tyr Gly Ala Val Asn Phe Leu Ser Ala Pro Gly Thr
 225 230 235 240
 Leu Leu Arg Gly Phe Ile Ala Arg Thr Leu Ser Asn Arg Thr Glu Glu
 245 250 255
 Leu Ile Asn Leu His Glu His Ile Ala Ser Leu Leu Arg Gly Arg Trp
 260 265 270
 Arg Gly Gln Glu Pro Val Asn Leu Arg Lys Tyr
 275 280

<210> 21
 <211> 594
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
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 <222> (101)..(571)
 <223> RXA02275

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ctacttcttg ttctaataccg ccaacaagga aggaagatcc atg att atc act gcg 115
Met Ile Ile Thr Ala
1 5

atc gac acc aac atc tac gat gaa ccg gag ttt gtt gaa gga cgc gat 163
Ile Asp Thr Asn Ile Tyr Asp Glu Pro Glu Phe Val Glu Gly Arg Asp
10 15 20

gtc atc ggt gtg cgc ttt gaa gat tta gtt ttg gat aag cgc att caa 211
Val Ile Gly Val Arg Phe Glu Asp Leu Val Leu Asp Lys Arg Ile Gln
25 30 35

cgg gtt gca ctc ccc gga gga gaa gaa ctg ggg ttg cgg tta aac cac 259
Arg Val Ala Leu Pro Gly Gly Glu Leu Leu Gly Leu Arg Leu Asn His
40 45 50

ggg cat ccg att ctg cgt gaa ggt gat gtg ttg aaa gct gat gat aag 307
Gly His Pro Ile Leu Arg Glu Gly Asp Val Leu Lys Ala Asp Asp Lys
55 60 65

acg gta ttt gtg gtg gag att atc ccc acg gat gtt tta gtt atc acg 355
Thr Val Phe Val Val Glu Ile Ile Pro Thr Asp Val Leu Val Ile Thr
70 75 80 85

cca agc gat att cac cag atg gga ttt gtg gcg cac tcc ctg gga aac 403
Pro Ser Asp Ile His Gln Met Gly Phe Val Ala His Ser Leu Gly Asn
90 95 100

agg cac ctg cca gca cag ttt tcc aag cca ggt gaa ttg aca gag aag 451
Arg His Leu Pro Ala Gln Phe Ser Lys Pro Gly Glu Leu Thr Glu Lys
105 110 115

gca gcc atg atc gtg caa tac gat cac acg gtg gtc agc ttc ttg gat 499
Ala Ala Met Ile Val Gln Tyr Asp His Thr Val Val Ser Phe Leu Asp
120 125 130

gag cac ggc atc gag tat cag cgc acc gaa ctt gtt ccg cca att cct 547
Glu His Gly Ile Glu Tyr Gln Arg Thr Glu Leu Val Pro Pro Ile Pro
135 140 145

ttc agg cat agc ggg cac aca cat tgatggatct tgacgctgat ttt 594
Phe Arg His Ser Gly His Thr His
150 155

<210> 22

<211> 157

<212> PRT

<213> Corynebacterium glutamicum

<400> 22

Met Ile Ile Thr Ala Ile Asp Thr Asn Ile Tyr Asp Glu Pro Glu Phe
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Val Glu Gly Arg Asp Val Ile Gly Val Arg Phe Glu Asp Leu Val Leu
20 25 30

Asp Lys Arg Ile Gln Arg Val Ala Leu Pro Gly Gly Glu Glu Leu Gly
35 40 45

Leu Arg Leu Asn His Gly His Pro Ile Leu Arg Glu Gly Asp Val Leu
 50 55 60
 Lys Ala Asp Asp Lys Thr Val Phe Val Val Glu Ile Ile Pro Thr Asp
 65 70 75 80
 Val Leu Val Ile Thr Pro Ser Asp Ile His Gln Met Gly Phe Val Ala
 85 90 95
 His Ser Leu Gly Asn Arg His Leu Pro Ala Gln Phe Ser Lys Pro Gly
 100 105 110
 Glu Leu Thr Glu Lys Ala Ala Met Ile Val Gln Tyr Asp His Thr Val
 115 120 125
 Val Ser Phe Leu Asp Glu His Gly Ile Glu Tyr Gln Arg Thr Glu Leu
 130 135 140
 Val Pro Pro Ile Pro Phe Arg His Ser Gly His Thr His
 145 150 155

<210> 23
 <211> 801
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(778)
 <223> RXA02276

<400> 23
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 Met Asp Leu Asp Ala
 1 5
 gat ttt ctg ctg ttg cat tta tct gat tca gca ctt cca acg gga gcg 163
 Asp Phe Leu Leu Leu His Leu Ser Asp Ser Ala Leu Pro Thr Gly Ala
 10 15 20
 ttt gcg cac tca ttt gga ttt gaa act tat atg gat gca gag cga atc 211
 Phe Ala His Ser Phe Gly Phe Glu Thr Tyr Met Asp Ala Glu Arg Ile
 25 30 35
 acc aat gca gag gag ttc caa gac tgg ctg aaa gtc ctg ctt aag gtg 259
 Thr Asn Ala Glu Glu Phe Gln Asp Trp Leu Lys Val Leu Leu Lys Val
 40 45 50
 caa ttg acc agc tct gat gct ttg gca atg agg atg ttt tac gcc acc 307
 Gln Leu Thr Ser Ser Asp Ala Leu Ala Met Arg Met Phe Tyr Ala Thr
 55 60 65
 ccg acg gtg tct gag ctg aaa cgg ctg gat gag cgc ctt ttt gct gga 355
 Pro Thr Val Ser Glu Leu Lys Arg Leu Asp Glu Arg Leu Phe Ala Gly
 70 75 80 85
 act ccg gcg aga gaa att cgg gaa gct aat gct cga atg ggt acg cgc 403
 Thr Pro Ala Arg Glu Ile Arg Glu Ala Asn Ala Arg Met Gly Thr Arg

	90	95	100	
atg gca gag atc gtg gct gaa acc tac tcc gtg ccc ctg att gtt gag				451
Met Ala Glu Ile Val Ala Glu Thr Tyr Ser Val Pro Leu Ile Val Glu	105	110	115	
tat ctc gaa ttg att caa cat cga gag cta tca ggg cac ccg gct ttg				499
Tyr Leu Glu Leu Ile Gln His Arg Glu Leu Ser Gly His Pro Ala Leu	120	125	130	
gct ttg gct ctt gcc acc cac agc gcg ggg att gat gtg gat cga gca				547
Ala Leu Ala Leu Ala Thr His Ser Ala Gly Ile Asp Val Asp Arg Ala	135	140	145	
atc cac gct cac ctc acg gca acg gtg agt tcg ctg atc caa aat gcg				595
Ile His Ala His Leu Thr Ala Thr Val Ser Ser Leu Ile Gln Asn Ala	150	155	160	165
gtt cgt ggc atc cca ctg ggg caa atg gca ggt cag ccg gtg atg ttc				643
Val Arg Gly Ile Pro Leu Gly Gln Met Ala Gly Gln Arg Val Met Phe	170	175	180	
gcc atg cgt gag cat atc ggt gcg gcc gtg aaa cgt agc gcg aac ttg				691
Ala Met Arg Glu His Ile Gly Ala Val Lys Arg Ser Ala Asn Leu	185	190	195	
gat gag att gat ttc tgt tcg ggt gat cca ggc ttg gat att tca caa				739
Asp Glu Ile Asp Phe Cys Ser Gly Asp Pro Gly Leu Asp Ile Ser Gln	200	205	210	
atg gtt cat gaa acc caa cgc gca cga cta ttt atg agt taagaaggag				788
Met Val His Glu Thr Gln Arg Ala Arg Leu Phe Met Ser	215	220	225	
aaaagaaaca tgg				801
 <210> 24				
<211> 226				
<212> PRT				
<213> Corynebacterium glutamicum				
 <400> 24				
Met Asp Leu Asp Ala Asp Phe Leu Leu Leu His Leu Ser Asp Ser Ala	1	5	10	15
Leu Pro Thr Gly Ala Phe Ala His Ser Phe Gly Phe Glu Thr Tyr Met	20	25	30	
Asp Ala Glu Arg Ile Thr Asn Ala Glu Glu Phe Gln Asp Trp Leu Lys	35	40	45	
Val Leu Leu Lys Val Gln Leu Thr Ser Ser Asp Ala Leu Ala Met Arg	50	55	60	
Met Phe Tyr Ala Thr Pro Thr Val Ser Glu Leu Lys Arg Leu Asp Glu	65	70	75	80
Arg Leu Phe Ala Gly Thr Pro Ala Arg Glu Ile Arg Glu Ala Asn Ala	85	90	95	

Gly Arg Glu Ile Ala Ala Lys Tyr Ala Glu Ala Ile Tyr Ser Val Ala
 200 205 210

787
 ttg gat ttg gag caa gcg caa gat tat cgc tct gat att cat gct cgt
 Trp Asp Leu Glu Gln Ala Gln Asp Tyr Arg Ser Asp Ile His Ala Arg
 215 220 225

835
 gcc act gcc cag ggt cgc gag ccc atg ccg gtg ctt cct ggt ttg gtg
 Ala Thr Ala Gln Gly Arg Glu Pro Met Pro Val Leu Pro Gly Leu Val
 230 235 240 245

883
 act ttt gtt gcc acg acc gtg gaa gaa gcg cgt gca aaa cag cag gct
 Thr Phe Val Gly Thr Thr Val Glu Glu Ala Arg Ala Lys Gln Gln Ala
 250 255 260

931
 ctt aat gcg ttg ctg ccg gtc aaa gac tca cta aat cag ttg agt ttc
 Leu Asn Ala Leu Leu Pro Val Lys Asp Ser Leu Asn Gln Leu Ser Phe
 265 270 275

979
 ttt gtg ggt caa gat tgc tcg acg tgg gat ttg gat gca cct ccc cca
 Phe Val Gly Gln Asp Cys Ser Thr Trp Asp Leu Asp Ala Pro Pro Pro
 280 285 290

1027
 cca ctg cca ccg cta gaa gag ttt tcc ggt cct aaa gcc agg tac gaa
 Pro Leu Pro Pro Leu Glu Glu Phe Ser Gly Pro Lys Gly Arg Tyr Glu
 295 300 305

1039
 acg gtc ctg cgg
 Thr Val Leu Arg
 310

<210> 440
 <211> 313
 <212> PRT
 <213> *Corynebacterium glutamicum*

<400> 440
 Val Glu Gly Ser Val Glu Lys Leu Gly Leu Ile Ser Trp Trp Glu Glu
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Leu Ala Arg Thr Ala Glu Arg Gly Lys Leu Asp Ala Val Phe Leu Ala
 20 25 30

Asp Gly Gln Ala Ile Asn Pro Val Gly Leu Glu Asn Gly Pro Gly Trp
 35 40 45

Phe Leu Glu Pro Val Thr Ala Leu Thr Ala Met Ala Arg Ala Thr Asn
 50 55 60

Asn Ile Gly Leu Ile Ser Thr Ile Ser Ser Thr Phe Trp Gln Pro Phe
 65 70 75 80

His Ala Ala Arg Met Ile Ala Ser Leu Asp His Ile Ser Gly Gly Arg
 85 90 95

Ala Gly Ile Asn Val Val Thr Ser Met Thr Asp Ala Glu Ala Arg Asn
 100 105 110

His Gly Met Asp Ala Leu Pro Gly His Asp Val Arg Tyr Ala Arg Ala
 115 120 125

Ala Glu Phe Ile Glu Thr Ile Thr Ala Leu Trp Asp Ser Trp Pro Ala
130 135 140

Glu Ser Leu Val Met Asp Arg Ala Gly Lys Phe Ala Asp Ser Ser Leu
145 150 155 160

Ile Lys Ser Ile Asp His Asp Gly Glu Phe Phe Gln Val Ala Gly Pro
165 170 175

Leu Asn Ile Pro Ser Pro Pro Gln Gly Arg Pro Val Leu Phe Gln Ala
180 185 190

Gly Ser Ser Pro Gln Gly Arg Glu Ile Ala Ala Lys Tyr Ala Glu Ala
195 200 205

Ile Tyr Ser Val Ala Trp Asp Leu Glu Gln Ala Gln Asp Tyr Arg Ser
210 215 220

Asp Ile His Ala Arg Ala Thr Ala Gln Gly Arg Glu Pro Met Pro Val
225 230 235 240

Leu Pro Gly Leu Val Thr Phe Val Gly Thr Thr Val Glu Glu Ala Arg
245 250 255

Ala Lys Gln Gln Ala Leu Asn Ala Leu Leu Pro Val Lys Asp Ser Leu
260 265 270

Asn Gln Leu Ser Phe Phe Val Gly Gln Asp Cys Ser Thr Trp Asp Leu
275 280 285

Asp Ala Pro Pro Pro Pro Leu Pro Pro Leu Glu Glu Phe Ser Gly Pro
290 295 300

Lys Gly Arg Tyr Glu Thr Val Leu Arg
305 310

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2

Ile Phe Val Glu Ser Gly Gly Asp Asn Leu Ser Ala Thr Phe Ser Pro
 100 105 110
 Glu Leu Val Asp Phe Ser Ile Tyr Ile Ile Asp Val Ala Gln Gly Glu
 115 120 125
 Lys Ile Pro Arg Lys Ala Gly Gln Gly Met Ile Lys Ser Asp Leu Phe
 130 135 140
 Ile Ile Asn Lys Thr Asp Leu Ala Pro Tyr Val Gly Ala Asn Leu Asp
 145 150 155 160
 Val Met Val Glu Asp Ala Lys Ala Phe Arg Lys Asn Lys Pro Phe Cys
 165 170 175
 Leu Thr Asn Leu Arg Thr Asp Asp Gly Leu Asp Lys Val Leu Glu Trp
 180 185 190
 Ile Arg His Glu Val Met Met Gln Asp Leu Gln Glu Ala
 195 200 205

<210> 27
 <211> 1119
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(1096)
 <223> RXA02603

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 cgaccgcaga ggccgtccat aagatcgaaa gggctacgaa gtg agc gaa cac gcc 115
 Val Ser Glu His Ala
 1 5
 gct gaa cat cac cgc gat acc caa aat ttc tta acc tcc gaa ccg cac 163
 Ala Glu His His Arg Asp Thr Gln Asn Phe Leu Thr Ser Glu Pro His
 10 15 20
 acc acg gca atc gaa gac aac aag aag cgc caa ccg ccg aaa aac ctt 211
 Thr Thr Ala Ile Glu Asp Asn Lys Lys Arg Gln Pro Pro Lys Asn Leu
 25 30 35
 gct gac ggc atg atc aag gcg ctg cgc ccc aag cag tgg gtc aag aac 259
 Ala Asp Gly Met Ile Lys Ala Leu Arg Pro Lys Gln Trp Val Lys Asn
 40 45 50
 gtt ctt gtg cta gca gca cca ctt gct gct ggt gca gat gcg atc ttc 307
 Val Leu Val Leu Ala Ala Pro Leu Ala Ala Gly Ala Asp Ala Ile Phe
 55 60 65
 aac cag cgc acg atc atc gac gtt gct atc gca ttc gta gtg ttc tgc 355
 Asn Gln Arg Thr Ile Ile Asp Val Ala Ile Ala Phe Val Val Phe Cys
 70 75 80 85
 ttc ggt gca tca gcc att tac ttg gtt aat gat gcc cgt gac gtg gaa 403
 Phe Gly Ala Ser Ala Ile Tyr Leu Val Asn Asp Ala Arg Asp Val Glu

90										95										100									
gct	gac	cgc	gag	cac	cca	acc	aag	cgt	ttc	cgc	ccc	atc	gct	gca	gga					451									
Ala	Asp	Arg	Glu	His	Pro	Thr	Lys	Arg	Phe	Arg	Pro	Ile	Ala	Ala	Gly														
			105					110					115																
gtc	ctg	cca	gta	gga	atg	gca	tac	ggc	atg	gcc	gtg	gcg	ctc	att	gca					499									
Val	Leu	Pro	Val	Gly	Met	Ala	Tyr	Gly	Met	Ala	Val	Ala	Leu	Ile	Ala														
			120				125						130																
cta	tcc	atc	gga	ctg	tct	ttc	ctc	gcc	acc	gac	ggc	gtg	gca	ctt	gcc					547									
Leu	Ser	Ile	Gly	Leu	Ser	Phe	Leu	Ala	Thr	Asp	Gly	Val	Ala	Leu	Ala														
			135				140						145																
tgc	gtg	att	ggc	gtg	tac	att	gcg	ctg	cag	ctg	gga	tac	tgc	ttc	ggc					595									
Cys	Val	Ile	Gly	Val	Tyr	Ile	Ala	Leu	Gln	Leu	Gly	Tyr	Cys	Phe	Gly														
					155				160					165															
tgg	aag	cac	atg	cca	gtg	atc	gat	att	gcg	ctt	gtc	tcc	tcc	gga	ttc					643									
Trp	Lys	His	Met	Pro	Val	Ile	Asp	Ile	Ala	Leu	Val	Ser	Ser	Gly	Phe														
					170				175					180															
atg	ctc	cgc	gca	atg	gca	ggc	ggc	gtc	gca	gca	ggc	atc	gag	cta	tcc					691									
Met	Leu	Arg	Ala	Met	Ala	Gly	Gly	Val	Ala	Ala	Gly	Ile	Glu	Leu	Ser														
			185					190					195																
cag	tgg	ttc	ctg	cta	gtc	gct	gcg	ttt	ggc	tcc	ctg	ttc	atg	gca	tct					739									
Gln	Trp	Phe	Leu	Leu	Val	Ala	Ala	Phe	Gly	Ser	Leu	Phe	Met	Ala	Ser														
			200				205						210																
gga	aag	cgc	tac	gca	gaa	atc	ctt	ctg	cac	gag	cgc	acc	ggc	gct	aag					787									
Gly	Lys	Arg	Tyr	Ala	Glu	Ile	Leu	Leu	His	Glu	Arg	Thr	Gly	Ala	Lys														
			215			220						225																	
atc	cgc	aag	tcc	ctg	gaa	agc	tac	acc	ccc	acc	tac	ctg	cgc	ttc	gtt					835									
Ile	Arg	Lys	Ser	Leu	Glu	Ser	Tyr	Thr	Pro	Thr	Tyr	Leu	Arg	Phe	Val														
			230		235				240					245															
tgg	acc	atg	gca	gca	aca	gca	gtg	gtc	atg	tcc	tac	gca	ctg	tgg	ggc					883									
Trp	Thr	Met	Ala	Ala	Thr	Ala	Val	Val	Met	Ser	Tyr	Ala	Leu	Trp	Gly														
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Phe	Asp	Leu	Ser	Gln	His	Ser	Thr	Asp	Ala	Gly	Pro	Trp	Tyr	Gln	Ile														
			265					270						275															
tcc	atg	gtt	cca	ttc	acc	atc	gcc	atc	ctg	cgc	tac	gca	gcc	ggc	gta					979									
Ser	Met	Val	Pro	Phe	Thr	Ile	Ala	Ile	Leu	Arg	Tyr	Ala	Ala	Gly	Val														
			280				285						290																
gac	acc	ggc	gac	ggc	ggc	gcc	cct	gac	gaa	gtg	gca	ctc	agc	gac	aaa					1027									
Asp	Thr	Gly	Asp	Gly	Gly	Ala	Pro	Asp	Glu	Val	Ala	Leu	Ser	Asp	Lys														
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Val	Leu	Gln	Val	Leu	Ala	Ala	Leu	Ala	Trp	Val	Phe	Cys	Ile	Val	Met	Ala													
			310		315					320				325															
gtg	tac	atc	atg	ccg	atg	ttt	tgaatatttta	ccaatgaaca	tgc											1119									
Val	Tyr	Ile	Met	Pro	Met	Phe																							
				330																									

<210> 28
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 <212> PRT
 <213> Corynebacterium glutamicum

<400> 28

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Pro Pro Lys Asn Leu Ala Asp Gly Met Ile Lys Ala Leu Arg Pro Lys
      35          40          45

Gln Trp Val Lys Asn Val Leu Val Leu Ala Ala Pro Leu Ala Ala Gly
  50          55          60

Ala Asp Ala Ile Phe Asn Gln Arg Thr Ile Ile Asp Val Ala Ile Ala
  65          70          75      80

Phe Val Val Phe Cys Phe Gly Ala Ser Ala Ile Tyr Leu Val Asn Asp
      85          90          95

Ala Arg Asp Val Glu Ala Asp Arg Glu His Pro Thr Lys Arg Phe Arg
      100          105          110

Pro Ile Ala Ala Gly Val Leu Pro Val Gly Met Ala Tyr Gly Met Ala
      115          120          125

Val Ala Leu Ile Ala Leu Ser Ile Gly Leu Ser Phe Leu Ala Thr Asp
      130          135          140

Gly Val Ala Leu Ala Cys Val Ile Gly Val Tyr Ile Ala Leu Gln Leu
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Gly Tyr Cys Phe Gly Trp Lys His Met Pro Val Ile Asp Ile Ala Leu
      165          170          175

Val Ser Ser Gly Phe Met Leu Arg Ala Met Ala Gly Gly Val Ala Ala
      180          185          190

Gly Ile Glu Leu Ser Gln Trp Phe Leu Leu Val Ala Ala Phe Gly Ser
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Leu Phe Met Ala Ser Gly Lys Arg Tyr Ala Glu Ile Leu Leu His Glu
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Arg Thr Gly Ala Lys Ile Arg Lys Ser Leu Glu Ser Tyr Thr Pro Thr
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Tyr Leu Arg Phe Val Trp Thr Met Ala Ala Thr Ala Val Val Met Ser
      245          250          255

Tyr Ala Leu Trp Gly Phe Asp Leu Ser Gln His Ser Thr Asp Ala Gly
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Pro Trp Tyr Gln Ile Ser Met Val Pro Phe Thr Ile Ala Ile Leu Arg
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Tyr Ala Ala Gly Val Asp Thr Gly Asp Gly Gly Ala Pro Asp Glu Val
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 <212> DNA
 <213> Corynebacterium glutamicum

<220>
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 <222> (101)..(1981)
 <223> RXA01385

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 Met Gln Phe His Tyr
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 gaa gga tac gca acc ggt gac cca atg gag atg cgc gcg gaa ggt agc 163
 Glu Gly Tyr Ala Thr Gly Asp Pro Met Glu Met Arg Ala Glu Gly Ser
 10 15 20
 gga atc aac cgc ccg gac gat ctc ccc gag gtc atg gat gtt ctc atc 211
 Gly Ile Asn Arg Pro Asp Asp Leu Pro Glu Val Met Asp Val Leu Ile
 25 30 35
 gtt ggt gca ggt ccg gct ggc acc atc gca gcg gct cag ctt tcc cga 259
 Val Gly Ala Gly Pro Ala Gly Thr Ile Ala Ala Ala Gln Leu Ser Arg
 40 45 50
 ttc ccc aat gtg acc acc cgc ctc gta gag aga agc gac cgt cgc ctc 307
 Phe Pro Asn Val Thr Thr Arg Leu Val Glu Arg Ser Asp Arg Arg Leu
 55 60 65
 gaa cta gcc aat gca gat ggc gtg cac tcc cga acc att gaa act ttc 355
 Glu Leu Ala Asn Ala Asp Gly Val His Ser Arg Thr Ile Glu Thr Phe
 70 75 80 85
 cag gca ttt ggt ttc gcc cac gag atc ctc gcc gaa gct cat gaa atc 403
 Gln Ala Phe Gly Phe Ala His Glu Ile Leu Ala Glu Ala His Glu Ile
 90 95 100
 acc gac atg gcg ttc tgg aag ccg gac ccg caa aac cct cgt gag atc 451
 Thr Asp Met Ala Phe Trp Lys Pro Asp Pro Gln Asn Pro Arg Glu Ile
 105 110 115
 att cgc gac aac agc acc cgc gag ctg cca cag cac atc agt gaa ttt 499
 Ile Arg Asp Asn Ser Thr Arg Glu Leu Pro Gln His Ile Ser Glu Phe
 120 125 130
 ccg atg gcg ttg ctc acc cag acc cgc atc atc gac cac ttc aac cgg 547

Pro Met Ala Leu Leu Thr Gln Thr Arg Ile Ile Asp His Phe Asn Arg	
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Phe Met Lys Asn Ser Pro Thr Arg Met Lys Pro Asp Tyr Gly Tyr Glu	
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Phe Val Asp Phe Glu Val Glu Glu Asp Ala Glu Tyr Pro Val Ile Val	
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acc ctc cgc cgc acc agt ggc gag caa act ggc gaa ttg gtc acc gtc	691
Thr Leu Arg Arg Thr Ser Gly Glu Gln Thr Gly Glu Leu Val Thr Val	
185 190 195	
cga acc aag tac ctg gtc ggt gcc gat ggt gca cga agc caa gtg cgc	739
Arg Thr Lys Tyr Leu Val Gly Ala Asp Gly Ala Arg Ser Gln Val Arg	
200 205 210	
aaa tca ctg gga tac cga ctc caa ggt aag cag gct aac cac gct tgg	787
Lys Ser Leu Gly Tyr Arg Leu Gln Gly Lys Gln Ala Asn His Ala Trp	
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Gly Val Met Asp Ile His Ala Asn Thr Glu Phe Pro Asp Val Arg Lys	
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Lys Cys Thr Ile Lys Ser Asp Ser Gly Arg Thr Ile Leu Leu Ile Pro	
250 255 260	
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Arg Glu Gly Gly Phe Leu Phe Arg Leu Tyr Val Asp Leu Gly Glu Val	
265 270 275	
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Pro Asp Asp Gly Ser Lys Ala Val Arg Asp Thr Pro Leu Gln Asp Val	
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Ile Asp Thr Ala Asn Gln Ile Met Ala Pro Phe Thr Leu Asp Val Lys	
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Asn Val Val Trp Asn Ser Ile Tyr Glu Val Gly His Arg Val Ala Asp	
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His Phe Asp Asp Arg Val Ser Glu Lys Thr Ser Ser Glu His Pro Arg	
330 335 340	
att ttc att gct ggc gac gcc tgc cac acc cac agc gct aag gct ggc	1171
Ile Phe Ile Ala Gly Asp Ala Cys His Thr His Ser Ala Lys Ala Gly	
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Gln Gly Met Asn Val Ser Met Gln Asp Gly Phe Asn Leu Gly Trp Lys	
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Leu Gly His Val Ala Ser Gly Asn Ser Pro Arg Glu Leu Leu Gln Thr	

375	380	385	
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gcc caa gac ctt gag gat ttc tac cgc gcg aac tct gag ttc aat gcc Ala Gln Asp Leu Glu Asp Phe Tyr Arg Ala Asn Ser Glu Phe Asn Ala 425 430 435			1411
ggc tac atg acc cac tat cct cct tct tcc atc aca atg gat ggc agc Gly Tyr Met Thr His Tyr Pro Pro Ser Ser Ile Thr Met Asp Gly Ser 440 445 450			1459
aac caa gat ctg gca aag ggc tac cca att ggc cga cgc ttc aag tca Asn Gln Asp Leu Ala Lys Gly Tyr Pro Ile Gly Arg Arg Phe Lys Ser 455 460 465			1507
gcg atg gtt ggt cga gtc tgc gac ttc acc gaa aca cac ctc ggt cac Ala Met Val Gly Arg Val Cys Asp Phe Thr Glu Thr His Leu Gly His 470 475 480 485			1555
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gat gca ctt aac ggc gag ggt tct gag cta gac cgc tgg gca gaa tgg Asp Ala Leu Asn Gly Glu Gly Ser Glu Leu Asp Arg Trp Ala Glu Trp 505 510 515			1651
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ggt atc acc acg gac tcc gac atc ttt gat agt cgc gag atc tcc cgc Gly Ile Thr Thr Asp Ser Asp Ile Phe Asp Ser Arg Glu Ile Ser Arg 570 575 580			1843
gat ggt gtc gtg gtg gta gtc cga cca gac caa tac gtt tcc gga atc Asp Gly Val Val Val Val Arg Pro Asp Gln Tyr Val Ser Gly Ile 585 590 595			1891
ttc cca ctc act gat acc caa ggg ctt ggc gaa ttc ctc acc gga tac Phe Pro Leu Thr Asp Thr Gln Gly Leu Gly Glu Phe Leu Thr Gly Tyr 600 605 610			1939
ttc ccc aaa atg aaa ggc gca cat cag cta atc aac gcg aac Phe Pro Lys Met Lys Gly Ala His Gln Leu Ile Asn Ala Asn 615 620 625			1981

taaggcacag ctgttaaac agt

2004

<210> 30

<211> 627

<212> PRT

<213> *Corynebacterium glutamicum*

<400> 30

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Arg	Ala	Glu	Gly	Ser	Gly	Ile	Asn	Arg	Pro	Asp	Asp	Leu	Pro	Glu	Val
		20						25					30		

Met	Asp	Val	Leu	Ile	Val	Gly	Ala	Gly	Pro	Ala	Gly	Thr	Ile	Ala	Ala
		35					40					45			

Ala	Gln	Leu	Ser	Arg	Phe	Pro	Asn	Val	Thr	Thr	Arg	Leu	Val	Glu	Arg
	50						55				60				

Ser	Asp	Arg	Arg	Leu	Glu	Leu	Ala	Asn	Ala	Asp	Gly	Val	His	Ser	Arg
	65				70					75					80

Thr	Ile	Glu	Thr	Phe	Gln	Ala	Phe	Gly	Phe	Ala	His	Glu	Ile	Leu	Ala
				85					90					95	

Glu	Ala	His	Glu	Ile	Thr	Asp	Met	Ala	Phe	Trp	Lys	Pro	Asp	Pro	Gln
			100					105					110		

Asn	Pro	Arg	Glu	Ile	Ile	Arg	Asp	Asn	Ser	Thr	Arg	Glu	Leu	Pro	Gln
		115					120					125			

His	Ile	Ser	Glu	Phe	Pro	Met	Ala	Leu	Leu	Thr	Gln	Thr	Arg	Ile	Ile
	130					135					140				

Asp	His	Phe	Asn	Arg	Phe	Met	Lys	Asn	Ser	Pro	Thr	Arg	Met	Lys	Pro
	145				150					155					160

Asp	Tyr	Gly	Tyr	Glu	Phe	Val	Asp	Phe	Glu	Val	Glu	Glu	Asp	Ala	Glu
				165					170					175	

Tyr	Pro	Val	Ile	Val	Thr	Leu	Arg	Arg	Thr	Ser	Gly	Glu	Gln	Thr	Gly
			180					185					190		

Glu	Leu	Val	Thr	Val	Arg	Thr	Lys	Tyr	Leu	Val	Gly	Ala	Asp	Gly	Ala
		195					200					205			

Arg	Ser	Gln	Val	Arg	Lys	Ser	Leu	Gly	Tyr	Arg	Leu	Gln	Gly	Lys	Gln
	210					215					220				

Ala	Asn	His	Ala	Trp	Gly	Val	Met	Asp	Ile	His	Ala	Asn	Thr	Glu	Phe
	225				230					235					240

Pro	Asp	Val	Arg	Lys	Lys	Cys	Thr	Ile	Lys	Ser	Asp	Ser	Gly	Arg	Thr
				245					250					255	

Ile	Leu	Leu	Ile	Pro	Arg	Glu	Gly	Gly	Phe	Leu	Phe	Arg	Leu	Tyr	Val
			260					265					270		

Asp Leu Gly Glu Val Pro Asp Asp Gly Ser Lys Ala Val Arg Asp Thr
 275 280 285
 Pro Leu Gln Asp Val Ile Asp Thr Ala Asn Gln Ile Met Ala Pro Phe
 290 295 300
 Thr Leu Asp Val Lys Asn Val Val Trp Asn Ser Ile Tyr Glu Val Gly
 305 310 315 320
 His Arg Val Ala Asp His Phe Asp Asp Arg Val Ser Glu Lys Thr Ser
 325 330 335
 Ser Glu His Pro Arg Ile Phe Ile Ala Gly Asp Ala Cys His Thr His
 340 345 350
 Ser Ala Lys Ala Gly Gln Gly Met Asn Val Ser Met Gln Asp Gly Phe
 355 360 365
 Asn Leu Gly Trp Lys Leu Gly His Val Ala Ser Gly Asn Ser Pro Arg
 370 375 380
 Glu Leu Leu Gln Thr Tyr Ala Glu Glu Arg Glu Asp Ile Ala Tyr Lys
 385 390 395 400
 Leu Ile Glu Tyr Asp Lys Asn Trp Ser Thr Leu Met Ala Lys Pro Ser
 405 410 415
 Ser Glu Met Gly Ser Ala Gln Asp Leu Glu Asp Phe Tyr Arg Ala Asn
 420 425 430
 Ser Glu Phe Asn Ala Gly Tyr Met Thr His Tyr Pro Pro Ser Ser Ile
 435 440 445
 Thr Met Asp Gly Ser Asn Gln Asp Leu Ala Lys Gly Tyr Pro Ile Gly
 450 455 460
 Arg Arg Phe Lys Ser Ala Met Val Gly Arg Val Cys Asp Phe Thr Glu
 465 470 475 480
 Thr His Leu Gly His Gln Ala Thr Ala Asp Gly Arg Met Arg Ala Tyr
 485 490 495
 Val Phe Ala Gly Ser Asp Ala Leu Asn Gly Glu Gly Ser Glu Leu Asp
 500 505 510
 Arg Trp Ala Glu Trp Ala Glu Ala Asn Leu Asp Pro Thr Leu Val Asp
 515 520 525
 Ala Lys Val Ile Tyr Gln Ser Pro Tyr Thr Glu Leu Asp Thr Arg Gln
 530 535 540
 Val Pro Ser Val Phe Lys Pro Ala Val Gly Ile Phe Glu Leu Thr Asn
 545 550 555 560
 Val Glu Asn Ser Phe Gly Ile Thr Thr Asp Ser Asp Ile Phe Asp Ser
 565 570 575
 Arg Glu Ile Ser Arg Asp Gly Val Val Val Val Arg Pro Asp Gln
 580 585 590
 Tyr Val Ser Gly Ile Phe Pro Leu Thr Asp Thr Gln Gly Leu Gly Glu

38

aac cgt ttg acc gat gtc tcc cac gct ctc gag gtc gca acc cgc aag 595
 Asn Arg Leu Thr Asp Val Ser His Ala Leu Glu Val Ala Thr Arg Lys
 150 155 160 165
 gct gag tcc aag ttc ggc gtc gcg ctc ggc atc gtc gat ggc tac ggc 643
 Ala Glu Ser Lys Phe Gly Val Ala Leu Gly Ile Val Asp Gly Tyr Gly
 170 175 180
 gga cac ggc att ggc cgc cac atg cac gag gag cca tac ttg gct aat 691
 Gly His Gly Ile Gly Arg His Met His Glu Glu Pro Tyr Leu Ala Asn
 185 190 195
 gag ggc aag gcc ggc aag ggc cct gtg att cag gag ggc tcc gtg ctc 739
 Glu Gly Lys Ala Gly Lys Gly Pro Val Ile Gln Glu Gly Ser Val Leu
 200 205 210
 gcc att gag cct atg ctc acc ctc ggc acc gaa gat tcc gca gtg ctg 787
 Ala Ile Glu Pro Met Leu Thr Leu Gly Thr Glu Asp Ser Ala Val Leu
 215 220 225
 gaa gat gat tgg act gtc gtg act ctc gac ggt tca tgg gca tca cac 835
 Glu Asp Asp Trp Thr Val Val Thr Leu Asp Gly Ser Trp Ala Ser His
 230 235 240 245
 tgg gag cac acc gtt gca gcc acc aag ggc ggc ccg cgc atc ctc acg 883
 Trp Glu His Thr Val Ala Ala Thr Lys Gly Gly Pro Arg Ile Leu Thr
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 ccg cgt tat taaaatgatg cttttcgacg cat 915
 Pro Arg Tyr

<210> 32

<211> 264

<212> PRT

<213> *Corynebacterium glutamicum*

<400> 32

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 20 25 30
 Gln Ala Val Arg Ala Glu Ala Lys Ala Gly Met Ser Thr Trp Asp Leu
 35 40 45
 Asp Gln Ile Ala Glu Gln Val Ile Arg Asp Ala Gly Ala Val Pro Thr
 50 55 60
 Phe Leu Gly Tyr Gln Gly Phe Pro Ala Ser Val Cys Ala Ser Val Asn
 65 70 75 80
 Glu Val Ile Val His Gly Ile Pro Ser Lys Glu Thr Ile Leu Glu Glu
 85 90 95
 Gly Asp Leu Val Ser Ile Asp Cys Gly Ala Thr Phe Asp Gly Trp Val
 100 105 110

Gly Asp Ser Ala Trp Ser Phe Gly Ile Gly Glu Leu Asp Glu Asp Val
 115 120 125
 Gln Gly Leu Asn Leu Ala Thr Glu Trp Val Leu Met Glu Gly Met Lys
 130 135 140
 Ala Met Val Pro Gly Asn Arg Leu Thr Asp Val Ser His Ala Leu Glu
 145 150 155 160
 Val Ala Thr Arg Lys Ala Glu Ser Lys Phe Gly Val Ala Leu Gly Ile
 165 170 175
 Val Asp Gly Tyr Gly Gly His Gly Ile Gly Arg His Met His Glu Glu
 180 185 190
 Pro Tyr Leu Ala Asn Glu Gly Lys Ala Gly Lys Gly Pro Val Ile Gln
 195 200 205
 Glu Gly Ser Val Leu Ala Ile Glu Pro Met Leu Thr Leu Gly Thr Glu
 210 215 220
 Asp Ser Ala Val Leu Glu Asp Asp Trp Thr Val Val Thr Leu Asp Gly
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 Ser Trp Ala Ser His Trp Glu His Thr Val Ala Ala Thr Lys Gly Gly
 245 250 255
 Pro Arg Ile Leu Thr Pro Arg Tyr
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<210> 33
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 <212> DNA
 <213> Corynebacterium glutamicum

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 <222> (1)..(483)
 <223> FRXA00675

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 Gly Ile Gly Glu Leu Asp Glu Asp Val Gln Gly Leu Asn Leu Ala Thr
 20 25 30
 gag tgg gtc ctc atg gaa ggc atg aag gcc atg gtt cca ggc aac cgt 144
 Glu Trp Val Leu Met Glu Gly Met Lys Ala Met Val Pro Gly Asn Arg
 35 40 45
 ttg acc gat gtc tcc cac gct ctc gag gtc gca acc cgc aag gct gag 192
 Leu Thr Asp Val Ser His Ala Leu Glu Val Ala Thr Arg Lys Ala Glu
 50 55 60
 tcc aag ttc ggc gtc gcg ctc ggc atc gtc gat ggc tac ggc gga cac 240
 Ser Lys Phe Gly Val Ala Leu Gly Ile Val Asp Gly Tyr Gly Gly His
 65 70 75 80

ggc att ggc cgc cac atg cac gag gag cca tac ttg gct aat gag ggc 288
 Gly Ile Gly Arg His Met His Glu Glu Pro Tyr Leu Ala Asn Glu Gly
 85 90 95

aag gcc ggc aag ggc cct gtg att cag gag ggc tcc gtg ctc gcc att 336
 Lys Ala Gly Lys Gly Pro Val Ile Gln Glu Gly Ser Val Leu Ala Ile
 100 105 110

gag cct atg ctc acc ctc ggc acc gaa gat tcc gca gtg ctg gaa gat 384
 Glu Pro Met Leu Thr Leu Gly Thr Glu Asp Ser Ala Val Leu Glu Asp
 115 120 125

gat tgg act gtc gtg act ctc gac ggt tca tgg gca tca cac tgg gag 432
 Asp Trp Thr Val Val Thr Leu Asp Gly Ser Trp Ala Ser His Trp Glu
 130 135 140

cac acc gtt gca gcc acc aag ggc ggc ccg cgc atc ctc acg ccg cgt 480
 His Thr Val Ala Ala Thr Lys Gly Gly Pro Arg Ile Leu Thr Pro Arg
 145 150 155 160

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<210> 34

<211> 161

<212> PRT

<213> *Corynebacterium glutamicum*

<400> 34

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 20 25 30

Glu Trp Val Leu Met Glu Gly Met Lys Ala Met Val Pro Gly Asn Arg
 35 40 45

Leu Thr Asp Val Ser His Ala Leu Glu Val Ala Thr Arg Lys Ala Glu
 50 55 60

Ser Lys Phe Gly Val Ala Leu Gly Ile Val Asp Gly Tyr Gly Gly His
 65 70 75 80

Gly Ile Gly Arg His Met His Glu Glu Pro Tyr Leu Ala Asn Glu Gly
 85 90 95

Lys Ala Gly Lys Gly Pro Val Ile Gln Glu Gly Ser Val Leu Ala Ile
 100 105 110

Glu Pro Met Leu Thr Leu Gly Thr Glu Asp Ser Ala Val Leu Glu Asp
 115 120 125

Asp Trp Thr Val Val Thr Leu Asp Gly Ser Trp Ala Ser His Trp Glu
 130 135 140

His Thr Val Ala Ala Thr Lys Gly Gly Pro Arg Ile Leu Thr Pro Arg
 145 150 155 160

Tyr

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<210> 35
<211> 996
<212> DNA
<213> Corynebacterium glutamicum

<220>
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<222> (101)..(973)
<223> RXA01609

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Met Ser Lys Met Arg
1 5

gca cca ctt gta ccc gga att cct acc cca atc agg gaa gta cct gca 163
Ala Pro Leu Val Pro Gly Ile Pro Thr Pro Ile Arg Glu Val Pro Ala
10 15 20

cat att gaa cgt cca gaa tat gtg tgg aag gac gaa gtc caa gaa gca 211
His Ile Glu Arg Pro Glu Tyr Val Trp Lys Asp Glu Val Gln Glu Ala
25 30 35

atc ggt gag cct ttt gtg cag gcc cct gag gtc atc gag aag atg cgt 259
Ile Gly Glu Pro Phe Val Gln Ala Pro Glu Val Ile Glu Lys Met Arg
40 45 50

gag aca tct cgc atc gct gca aac tca ctg aaa atc gcg ggc gaa gcc 307
Glu Thr Ser Arg Ile Ala Ala Asn Ser Leu Lys Ile Ala Gly Glu Ala
55 60 65

gtc aag cca ggc gtg acc act gat gaa ctt gat cgc att gtg cat gag 355
Val Lys Pro Gly Val Thr Thr Asp Glu Leu Asp Arg Ile Val His Glu
70 75 80 85

tac acc tgc gat atg ggc gca tac cct tca gat ctt ggt tac cgg gga 403
Tyr Thr Cys Asp Met Gly Ala Tyr Pro Ser Asp Leu Gly Tyr Arg Gly
90 95 100

ttc acc aag tcc tca tgc att tcc ctc aat gag atc gtg tgc cac ggt 451
Phe Thr Lys Ser Ser Cys Ile Ser Leu Asn Glu Ile Val Cys His Gly
105 110 115

att cct gat tcc acc gtc att gaa gag ggc gat att gtt aac atc gat 499
Ile Pro Asp Ser Thr Val Ile Glu Glu Gly Asp Ile Val Asn Ile Asp
120 125 130

gtc acc gcg ttc aag cac ggc gtc cac ggc gac tgc aat gcc acc ttc 547
Val Thr Ala Phe Lys His Gly Val His Gly Asp Cys Asn Ala Thr Phe
135 140 145

tta gcg ggt gat gtt tct gaa gaa cac cgc ctg ctg gtt gag cgc acc 595
Leu Ala Gly Asp Val Ser Glu Glu His Arg Leu Leu Val Glu Arg Thr
150 155 160 165

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gaa gaa gcc atg atg cgt tcc atc cgt gca gca aag cct gga cgt gaa 643
 Glu Glu Ala Met Met Arg Ser Ile Arg Ala Ala Lys Pro Gly Arg Glu
 170 175 180

 atc aac gtc att ggg cgt gtc att gag tct tac gcc aag cgt ttt ggc 691
 Ile Asn Val Ile Gly Arg Val Ile Glu Ser Tyr Ala Lys Arg Phe Gly
 185 190 195

 tac aac gtg gtc cgc gat ttc acc gga cac ggc atc ggc cca act ttc 739
 Tyr Asn Val Val Arg Asp Phe Thr Gly His Gly Ile Gly Pro Thr Phe
 200 205 210

 cac aac ggc ctt gtg gtg ctg cac tac gac aac act cag tac cgc gat 787
 His Asn Gly Leu Val Val Leu His Tyr Asp Asn Thr Gln Tyr Arg Asp
 215 220 225

 ctg ctc gtg cca ggc atg acc ttg acc atc gag cca atg atc aac ctt 835
 Leu Leu Val Pro Gly Met Thr Leu Thr Ile Glu Pro Met Ile Asn Leu
 230 235 240 245

 ggt tcc ctc gac tac gag atc tgg gaa gat gat tgg act gtc caa aac 883
 Gly Ser Leu Asp Tyr Glu Ile Trp Glu Asp Asp Trp Thr Val Gln Asn
 250 255 260

 gtt gac cgt aag ttc agc gcg cag ttc gag cac acc att gtc atc acc 931
 Val Asp Arg Lys Phe Ser Ala Gln Phe Glu His Thr Ile Val Ile Thr
 265 270 275

 gaa gac ggc aat gag atc ctc acc ctc cca gac gat tcc gtc 973
 Glu Asp Gly Asn Glu Ile Leu Thr Leu Pro Asp Asp Ser Val
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 <213> Corynebacterium glutamicum

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 Glu Val Gln Glu Ala Ile Gly Glu Pro Phe Val Gln Ala Pro Glu Val
 35 40 45

 Ile Glu Lys Met Arg Glu Thr Ser Arg Ile Ala Ala Asn Ser Leu Lys
 50 55 60

 Ile Ala Gly Glu Ala Val Lys Pro Gly Val Thr Thr Asp Glu Leu Asp
 65 70 75 80

 Arg Ile Val His Glu Tyr Thr Cys Asp Met Gly Ala Tyr Pro Ser Asp
 85 90 95

 Leu Gly Tyr Arg Gly Phe Thr Lys Ser Ser Cys Ile Ser Leu Asn Glu

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100          105          110
Ile Val Cys His Gly Ile Pro Asp Ser Thr Val Ile Glu Glu Gly Asp
115          120          125

Ile Val Asn Ile Asp Val Thr Ala Phe Lys His Gly Val His Gly Asp
130          135          140

Cys Asn Ala Thr Phe Leu Ala Gly Asp Val Ser Glu Glu His Arg Leu
145          150          155          160

Leu Val Glu Arg Thr Glu Glu Ala Met Met Arg Ser Ile Arg Ala Ala
165          170          175

Lys Pro Gly Arg Glu Ile Asn Val Ile Gly Arg Val Ile Glu Ser Tyr
180          185          190

Ala Lys Arg Phe Gly Tyr Asn Val Val Arg Asp Phe Thr Gly His Gly
195          200          205

Ile Gly Pro Thr Phe His Asn Gly Leu Val Val Leu His Tyr Asp Asn
210          215          220

Thr Gln Tyr Arg Asp Leu Leu Val Pro Gly Met Thr Leu Thr Ile Glu
225          230          235          240

Pro Met Ile Asn Leu Gly Ser Leu Asp Tyr Glu Ile Trp Glu Asp Asp
245          250          255

Trp Thr Val Gln Asn Val Asp Arg Lys Phe Ser Ala Gln Phe Glu His
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Thr Ile Val Ile Thr Glu Asp Gly Asn Glu Ile Leu Thr Leu Pro Asp
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Asp Ser Val
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<223> RXA01358

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Met Ala Leu Gly Arg
1 5

act ata tcc act gcg caa tta ggt gtg cag gca aaa atc gtt cgt gtg 163
Thr Ile Ser Thr Ala Gln Leu Gly Val Gln Ala Lys Ile Val Arg Val
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gag gct aat gtt ggc cca gga ttg cct ggt acc tac att gtt gga tta 211

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Ala	Asp	Thr	Ala	Ile	Ser	Glu	Ser	Arg	Asp	Arg	Ile	Lys	Thr	Ala	Val	
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Gln	Asn	Ser	Gly	Leu	Met	Trp	Pro	Lys	Thr	Lys	Val	Ile	Ile	Asn	Leu	
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Ser	Pro	Ala	Ser	Met	Arg	Lys	Gln	Gly	Ser	Gln	Cys	Asp	Leu	Ala	Met	
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acc	gtt	gca	gtt	ctc	gtt	gcc	cat	ggc	tct	aac	ccc	aaa	gcg	aag	ttt	403
Thr	Val	Ala	Val	Leu	Val	Ala	His	Gly	Ser	Asn	Pro	Lys	Ala	Lys	Phe	
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cat	gcg	cag	aac	acg	tta	ttt	ctg	ggg	gag	gtg	gcg	ctt	gat	gga	acc	451
His	Ala	Gln	Asn	Thr	Leu	Phe	Leu	Gly	Glu	Val	Ala	Leu	Asp	Gly	Thr	
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Leu	Leu	Pro	Val	Thr	Gly	Val	Leu	Pro	Ala	Leu	Leu	Ala	Ala	Lys	Glu	
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gaa	ggg	att	ggc	aag	att	gtg	atc	ccg	gag	gga	aat	gcc	caa	gaa	gca	547
Glu	Gly	Ile	Gly	Lys	Ile	Val	Ile	Pro	Glu	Gly	Asn	Ala	Gln	Glu	Ala	
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Val	Leu	Arg	Trp	Leu	Asp	Gly	Glu	Glu	Ala	Leu	Pro	Gln	Pro	Gly	Leu	
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Phe	Asn	Asp	Glu	Asn	Ser	Leu	Lys	Leu	Pro	Asp	Met	Arg	Asp	Val	Val	
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Gly	Gln	Pro	Glu	Ala	Arg	Phe	Ala	Ala	Glu	Val	Ala	Ala	Ala	Gly	Gly	
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Ala	Glu	Arg	Ile	Pro	Ser	Leu	Leu	Pro	Glu	Leu	Ser	Pro	Gln	Gln	Met	
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atc	gag	gcg	acg	gca	gtg	cat	tcc	gtt	gtg	ggg	cga	acc	ttt	tca	ggg	883
Ile	Glu	Ala	Thr	Ala	Val	His	Ser	Val	Val	Gly	Arg	Thr	Phe	Ser	Gly	
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ccg	gtg	tcg	agg	gct	ccg	ttt	att	tcc	cca	cac	cac	aat	gtc	agc	aag	931
Pro	Val	Ser	Arg	Ala	Pro	Phe	Ile	Ser	Pro	His	His	Asn	Val	Ser	Lys	

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agc ctg gcg cat cat ggt gtg ttg ttc ctg gat gag gtt agt gag att Ser Leu Ala His His Gly Val Leu Phe Leu Asp Glu Val Ser Glu Ile 295 300 305			1027
cca gcg tca atc ctt gat tct ttg agg act cca ttg gaa tac ggc tgc Pro Ala Ser Ile Leu Asp Ser Leu Arg Thr Pro Leu Glu Tyr Gly Ser 310 315 320 325			1075
atc cgc atc atc aga tcc cgc cat gat gtc acc ttc ccc gca cag ttc Ile Arg Ile Ile Arg Ser Arg His Asp Val Thr Phe Pro Ala Gln Phe 330 335 340			1123
cag ctc atc ctc gcg gcc aat ccg tgt aga tgc ggt gca gaa cag cct Gln Leu Ile Leu Ala Ala Asn Pro Cys Arg Cys Gly Ala Glu Gln Pro 345 350 355			1171
caa gaa tgt gtc tgt tct ggc tca gct cgc gcg acg tac ctt aat aat Gln Glu Cys Val Cys Ser Gly Ser Ala Arg Ala Thr Tyr Leu Asn Asn 360 365 370			1219
ctt tgc ggt ccg ttg agg gat cgc ttg gac atg gtt gtt gcc acc cac Leu Ser Gly Pro Leu Arg Asp Arg Leu Asp Met Val Val Ala Thr His 375 380 385			1267
tct aaa ggt gca gtg ctg cgt agt gat gac gtt gag gca tct gct ccc Ser Lys Gly Ala Val Leu Arg Ser Asp Asp Val Glu Ala Ser Ala Pro 390 395 400 405			1315
att gct gat cgg gtg gca caa gct cgt gag agg gca gct ttc cga tgg Ile Ala Asp Arg Val Ala Gln Ala Arg Glu Arg Ala Ala Phe Arg Trp 410 415 420			1363
cgc cgt tct gga ctg gga aat ctt gtt aat gca cac gta gat cca cac Arg Arg Ser Gly Leu Gly Asn Leu Val Asn Ala His Val Asp Pro His 425 430 435			1411
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ggc gcg ttc ctg gcg gaa gga aca atc tcc caa cgt ggc tgc gat cgg Gly Ala Phe Leu Ala Glu Gly Thr Ile Ser Gln Arg Gly Cys Asp Arg 455 460 465			1507
gcc ata aaa ctg ggt tgg acc ttg tgc gat ttg gat ggg gaa cag cag Ala Ile Lys Leu Gly Trp Thr Leu Cys Asp Leu Asp Gly Glu Gln Gln 470 475 480 485			1555
ccc aat ctt gac cat att gcg cga gcc atg gag ctt cgg ggc act aca Pro Asn Leu Asp His Ile Ala Arg Ala Met Glu Leu Arg Gly Thr Thr 490 495 500			1603
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 <213> *Corynebacterium glutamicum*

<400> 38

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Lys Ile Val Arg Val Glu Ala Asn Val Gly Pro Gly Leu Pro Gly Thr
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Tyr Ile Val Gly Leu Ala Asp Thr Ala Ile Ser Glu Ser Arg Asp Arg
          35           40           45

Ile Lys Thr Ala Val Gln Asn Ser Gly Leu Met Trp Pro Lys Thr Lys
          50           55           60

Val Ile Ile Asn Leu Ser Pro Ala Ser Met Arg Lys Gln Gly Ser Gln
 65           70           75           80

Cys Asp Leu Ala Met Thr Val Ala Val Leu Val Ala His Gly Ser Asn
          85           90           95

Pro Lys Ala Lys Phe His Ala Gln Asn Thr Leu Phe Leu Gly Glu Val
          100          105          110

Ala Leu Asp Gly Thr Leu Leu Pro Val Thr Gly Val Leu Pro Ala Leu
          115          120          125

Leu Ala Ala Lys Glu Glu Gly Ile Gly Lys Ile Val Ile Pro Glu Gly
          130          135          140

Asn Ala Gln Glu Ala Gly Leu Val Glu Asp Pro Ser Val Phe Leu Ala
          145          150          155          160

His Ser Ile Asp Gln Val Leu Arg Trp Leu Asp Gly Glu Glu Ala Leu
          165          170          175

Pro Gln Pro Gly Leu Phe Asn Asp Glu Asn Ser Leu Lys Leu Pro Asp
          180          185          190

Met Arg Asp Val Val Gly Gln Pro Glu Ala Arg Phe Ala Ala Glu Val
          195          200          205

Ala Ala Ala Gly Gly His His Met Leu Met Ile Gly Pro Pro Gly Ser
          210          215          220

Gly Lys Ser Met Ile Ala Glu Arg Ile Pro Ser Leu Leu Pro Glu Leu
          225          230          235          240

Ser Pro Gln Gln Met Ile Glu Ala Thr Ala Val His Ser Val Val Gly
          245          250          255

Arg Thr Phe Ser Gly Pro Val Ser Arg Ala Pro Phe Ile Ser Pro His
          260          265          270

His Asn Val Ser Lys Ala Ala Leu Leu Gly Gly Gly Ser Gly Ser Pro
          275          280          285
  
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Leu Pro Gly Ala Ile Ser Leu Ala His His Gly Val Leu Phe Leu Asp
 290 295 300
 Glu Val Ser Glu Ile Pro Ala Ser Ile Leu Asp Ser Leu Arg Thr Pro
 305 310 315 320
 Leu Glu Tyr Gly Ser Ile Arg Ile Ile Arg Ser Arg His Asp Val Thr
 325 330 335
 Phe Pro Ala Gln Phe Gln Leu Ile Leu Ala Ala Asn Pro Cys Arg Cys
 340 345 350
 Gly Ala Glu Gln Pro Gln Glu Cys Val Cys Ser Gly Ser Ala Arg Ala
 355 360 365
 Thr Tyr Leu Asn Asn Leu Ser Gly Pro Leu Arg Asp Arg Leu Asp Met
 370 375 380
 Val Val Ala Thr His Ser Lys Gly Ala Val Leu Arg Ser Asp Asp Val
 385 390 395 400
 Glu Ala Ser Ala Pro Ile Ala Asp Arg Val Ala Gln Ala Arg Glu Arg
 405 410 415
 Ala Ala Phe Arg Trp Arg Arg Ser Gly Leu Gly Asn Leu Val Asn Ala
 420 425 430
 His Val Asp Pro His Phe Leu Arg Arg Asn Phe Ala Ala Thr Glu Asp
 435 440 445
 Ala Met Val Tyr Leu Gly Ala Phe Leu Ala Glu Gly Thr Ile Ser Gln
 450 455 460
 Arg Gly Cys Asp Arg Ala Ile Lys Leu Gly Trp Thr Leu Cys Asp Leu
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 <223> RXA01458

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 Val Asn Arg Arg Ile
 1 5

aag act ctg acg tgg ggt gct atc cct ttg gtg ctg ctg gca tgc ttg	163
Lys Thr Leu Thr Trp Gly Ala Ile Pro Leu Val Leu Leu Ala Ser Leu	
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gta agc att gac cat att ccg gga aca aac atc aac ttg agc gtg cct	211
Val Ser Ile Asp His Ile Pro Gly Thr Asn Ile Asn Leu Ser Val Pro	
25 30 35	
tat gcc gct gaa ggc cca ggt cct acg atc aat acg ctt ggt cag gtc	259
Tyr Ala Ala Glu Gly Pro Gly Pro Thr Ile Asn Thr Leu Gly Gln Val	
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gac ggc gag gat gtt gtg tcc atc agt agt gct gat ctg gat gag acc	307
Asp Gly Glu Asp Val Val Ser Ile Ser Ser Ala Asp Leu Asp Glu Thr	
55 60 65	
gaa ggt aac ctg aac atg acc act gtg tgc gtt cgt tcc ggc atg aca	355
Glu Gly Asn Leu Asn Met Thr Thr Val Ser Val Arg Ser Gly Met Thr	
70 75 80 85	
ttg tgc cag gta att tcc cga tgg ctg ttt acc gat gac aca atc gtt	403
Leu Ser Gln Val Ile Ser Arg Trp Leu Phe Thr Asp Asp Thr Ile Val	
90 95 100	
ccc atc gag cag gtt ttc cct ccc ggc caa tcc acc gag gaa gtc gaa	451
Pro Ile Glu Gln Val Phe Pro Pro Gly Gln Ser Thr Glu Glu Val Glu	
105 110 115	
gaa tcc aac cgc acc gcg ttc atc tct tgc gag tct tcc gca acg atc	499
Glu Ser Asn Arg Thr Ala Phe Ile Ser Ser Glu Ser Ser Ala Thr Ile	
120 125 130	
gcc gcg atg aat tac ctc aac att ccc gtc gaa gtt gaa gtt gca gaa	547
Ala Ala Met Asn Tyr Leu Asn Ile Pro Val Glu Val Glu Val Ala Glu	
135 140 145	
gtc ctc acc gac agc gcc gca acc gga att ttc gaa ccc ggc gac aaa	595
Val Leu Thr Asp Ser Ala Ala Thr Gly Ile Phe Glu Pro Gly Asp Lys	
150 155 160 165	
ctt ctc agc atc gac ggc acc gca atc tcc act ccc ggc gat gca caa	643
Leu Leu Ser Ile Asp Gly Thr Ala Ile Ser Thr Pro Gly Asp Ala Gln	
170 175 180	
acc atc gtg cga tgc aaa gct ccc ggc gat gag atc acg att tcc tac	691
Thr Ile Val Arg Ser Lys Ala Pro Gly Asp Glu Ile Thr Ile Ser Tyr	
185 190 195	
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Glu Arg Asn Asp Ala Glu Ser Gln Ala Thr Ile Thr Leu Arg Glu His	
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ccg gat gat tct tgc gtg gcg ttg ttg ggt att tca atg ttg tgc gtg	787
Pro Asp Asp Ser Ser Val Ala Leu Leu Gly Ile Ser Met Leu Ser Val	
215 220 225	
cct tgc agc gcg att gag gtt gat tac aac ttg gaa gat atc ggt ggt	835
Pro Ser Ser Ala Ile Glu Val Asp Tyr Asn Leu Glu Asp Ile Gly Gly	
230 235 240 245	
ccg agc gct ggc atg atg ttt tgc ttg gcg gtc gtc gat aag ctt tgc	883

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Pro Ser Ala Gly Met Met Phe Ser Leu Ala Val Val Asp Lys Leu Ser
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Pro Gly Ala Leu Asn Gly Gly Lys Phe Val Ala Gly Thr Gly Thr Ile
      265                      270                      275

gcg gag gac ggg tcg gtg ggc ccg att ggc ggt att gcg cac aag gtg 979
Ala Glu Asp Gly Ser Val Gly Pro Ile Gly Gly Ile Ala His Lys Val
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cgc gct gcg gag gac gcg ggc gcg gaa gtg ttt ttg agc cct gcg gac 1027
Arg Ala Ala Glu Asp Ala Gly Ala Glu Val Phe Leu Ser Pro Ala Asp
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aat tgc gcg gag gcg atg agt gcg aag cct cag gat atg acg atc ttg 1075
Asn Cys Ala Glu Ala Met Ser Ala Lys Pro Gln Asp Met Thr Ile Leu
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Lys Val Asp Ser Leu Ser Gln Ala Ile Asp Gln Met Ala Ala Tyr Asn
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Glu Gly Ser Asp Phe Gln Thr Cys Gly
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ttc 1173

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<213> Corynebacterium glutamicum

<400> 40
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Leu Leu Ala Ser Leu Val Ser Ile Asp His Ile Pro Gly Thr Asn Ile
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Asn Leu Ser Val Pro Tyr Ala Ala Glu Gly Pro Gly Pro Thr Ile Asn
      35                      40                      45

Thr Leu Gly Gln Val Asp Gly Glu Asp Val Val Ser Ile Ser Ser Ala
      50                      55                      60

Asp Leu Asp Glu Thr Glu Gly Asn Leu Asn Met Thr Thr Val Ser Val
      65                      70                      75                      80

Arg Ser Gly Met Thr Leu Ser Gln Val Ile Ser Arg Trp Leu Phe Thr
      85                      90                      95

Asp Asp Thr Ile Val Pro Ile Glu Gln Val Phe Pro Pro Gly Gln Ser
      100                      105                      110

Thr Glu Glu Val Glu Glu Ser Asn Arg Thr Ala Phe Ile Ser Ser Glu
      115                      120                      125

Ser Ser Ala Thr Ile Ala Ala Met Asn Tyr Leu Asn Ile Pro Val Glu

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Val Glu Val Ala Glu Val Leu Thr Asp Ser Ala Ala Thr Gly Ile Phe		
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Glu Pro Gly Asp Lys Leu Leu Ser Ile Asp Gly Thr Ala Ile Ser Thr		
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Pro Gly Asp Ala Gln Thr Ile Val Arg Ser Lys Ala Pro Gly Asp Glu		
180	185	190
Ile Thr Ile Ser Tyr Glu Arg Asn Asp Ala Glu Ser Gln Ala Thr Ile		
195	200	205
Thr Leu Arg Glu His Pro Asp Asp Ser Ser Val Ala Leu Leu Gly Ile		
210	215	220
Ser Met Leu Ser Val Pro Ser Ser Ala Ile Glu Val Asp Tyr Asn Leu		
225	230	235
Glu Asp Ile Gly Gly Pro Ser Ala Gly Met Met Phe Ser Leu Ala Val		
245	250	255
Val Asp Lys Leu Ser Pro Gly Ala Leu Asn Gly Gly Lys Phe Val Ala		
260	265	270
Gly Thr Gly Thr Ile Ala Glu Asp Gly Ser Val Gly Pro Ile Gly Gly		
275	280	285
Ile Ala His Lys Val Arg Ala Ala Glu Asp Ala Gly Ala Glu Val Phe		
290	295	300
Leu Ser Pro Ala Asp Asn Cys Ala Glu Ala Met Ser Ala Lys Pro Gln		
305	310	315
Asp Met Thr Ile Leu Lys Val Asp Ser Leu Ser Gln Ala Ile Asp Gln		
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Met Ala Ala Tyr Asn Glu Gly Ser Asp Phe Gln Thr Cys Gly		
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<211> 1119

<212> DNA

<213> Corynebacterium glutamicum

<220>

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<222> (101)..(1096)

<223> RXA01654

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Asn	Ala	Ala	Glu	Ala	Pro	Ala	Ser	Glu	Trp	Val	Asn	Thr	Thr	Ala	Ile

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Val	Asp	Gln	Ala	Asn	Ala	Gln	Leu	Ser	Gln	Phe	Gly	Val	Ser	Leu	Asp														
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Arg	Ser	Ala	Ala	Glu	Leu	Phe	Asp	Asp	Gln	Ala	Asn	Ser	Gln	Ile	Asp														
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Ala	Ala	Leu	Ser	Pro	Tyr	Ala	Asp	Lys	Val	Pro	Thr	Ser	Gly	Gly	Gln														
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Leu	Pro	Asn	Tyr	Glu	Ile	Arg	Thr	Asp	Leu	Gln	Ser	Gln	Val	Met	Gly														
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Ala	Thr	Leu	Gly	Glu	Val	Leu	His	Arg	Val	Pro	Gly	Ser	Trp	Phe	Asp														
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Ala	Pro	Ala	Val	Pro	Glu	Glu	Ser	Arg	Ile	Val	Glu	Glu	Gln	Gly	Lys														
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Ser	Leu	Tyr	Gly	Pro	Gly	Thr	Pro	Ile	Tyr	Leu	Asn	Gly	Asn	Ser	Met														
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Ile	Thr	Ala	Gly	His	Cys	Gly	Lys	Ser	Gly	Asp	Ala	Val	Arg	Ser	Ala														
				170					175					180															
gac	tcc	ttc	tgg	gtc	ggc	gat	acc	gga	aca	gtg	gtg	tac	aac	gcg	cct					691									
Asp	Ser	Phe	Trp	Val	Gly	Asp	Thr	Gly	Thr	Val	Val	Tyr	Asn	Ala	Pro														
			185					190						195															
aac	gct	gac	tac	tcc	gtg	atc	gag	ttc	ggg	tcc	aat	gca	gag	ttg	agc					739									
Asn	Ala	Asp	Tyr	Ser	Val	Ile	Glu	Phe	Gly	Ser	Asn	Glu	Leu	Ser															
			200					205					210																
aat	acc	tac	aac	ggg	gtc	acc	gcg	aat	gct	gtc	ggc	ggg	ggc	gtg	act					787									
Asn	Thr	Tyr	Asn	Gly	Val	Thr	Ala	Asn	Ala	Val	Gly	Gly	Gly	Val	Thr														
			215				220						225																
aat	ggc	caa	gaa	gta	tgc	aaa	aac	gga	gtt	gct	act	ggc	tac	acc	tgt					835									
Asn	Gly	Gln	Glu	Val	Cys	Lys	Asn	Gly	Val	Ala	Thr	Gly	Tyr	Thr	Cys														
			230				235					240			245														
ggg	ttg	gtg	tgg	act	gct	gat	gag	cgc	atg	acg	atg	tct	cag	gtg	tgt					883									
Gly	Leu	Val	Trp	Thr	Ala	Asp	Glu	Arg	Met	Thr	Met	Ser	Gln	Val	Cys														
				250					255						260														

gcg ggt cgt ggt gat tcg ggt gct ccg ctg att gca gat ggt cgt gtg 931
 Ala Gly Arg Gly Asp Ser Gly Ala Pro Leu Ile Ala Asp Gly Arg Val
 265 270 275

gtt ggt ctt gta tct ggt ggt gta att cct gat tac aac ctg gca tgc 979
 Val Gly Leu Val Ser Gly Gly Val Ile Pro Asp Tyr Asn Leu Ala Cys
 280 285 290

gcc act ccg ttg cag gga cct ttc ttc atg cca acg ctg tca gtg aac 1027
 Ala Thr Pro Leu Gln Gly Pro Phe Phe Met Pro Thr Leu Ser Val Asn
 295 300 305

atg gat act gtc cta act gat ttg gat tcg cag gat ctt ccc ggt cga 1075
 Met Asp Thr Val Leu Thr Asp Leu Asp Ser Gln Asp Leu Pro Gly Arg
 310 315 320 325

ggt ttt cag cca act gct gga tagaatttag aaaatccgcc gtt 1119
 Gly Phe Gln Pro Thr Ala Gly
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<210> 42

<211> 332

<212> PRT

<213> *Corynebacterium glutamicum*

<400> 42

Met Ser Pro Ser Ala Asn Ala Ala Glu Ala Pro Ala Ser Glu Trp Val
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Asn Thr Thr Ala Ile Val Asp Gln Ala Asn Ala Gln Leu Ser Gln Phe
 20 25 30

Gly Val Ser Leu Asp Arg Ser Ala Ala Glu Leu Phe Asp Asp Gln Ala
 35 40 45

Asn Ser Gln Ile Asp Ala Ala Leu Ser Pro Tyr Ala Asp Lys Val Pro
 50 55 60

Thr Ser Gly Gly Gln Val Val Glu Gln Ser Leu Gln Val Val Glu Gln
 65 70 75 80

Glu Val Gln Lys Ala Leu Pro Asn Tyr Glu Ile Arg Thr Asp Leu Gln
 85 90 95

Ser Gln Val Met Gly Ala Thr Leu Gly Glu Val Leu His Arg Val Pro
 100 105 110

Gly Ser Trp Phe Asp Ala Pro Ala Val Pro Glu Glu Ser Arg Ile Val
 115 120 125

Glu Glu Gln Gly Lys Ser Leu Tyr Gly Pro Gly Thr Pro Ile Tyr Leu
 130 135 140

Asn Gly Asn Ser Met Cys Thr Leu Ala Val Thr Gly Thr Asp Ala Asp
 145 150 155 160

Gly Arg Lys Ile Gly Ile Thr Ala Gly His Cys Gly Lys Ser Gly Asp
 165 170 175

Ala Val Arg Ser Ala Asp Ser Phe Trp Val Gly Asp Thr Gly Thr Val
 180 185 190

Val Tyr Asn Ala Pro Asn Ala Asp Tyr Ser Val Ile Glu Phe Gly Ser
 195 200 205

Asn Ala Glu Leu Ser Asn Thr Tyr Asn Gly Val Thr Ala Asn Ala Val
 210 215 220

Gly Gly Gly Val Thr Asn Gly Gln Glu Val Cys Lys Asn Gly Val Ala
 225 230 235 240

Thr Gly Tyr Thr Cys Gly Leu Val Trp Thr Ala Asp Glu Arg Met Thr
 245 250 255

Met Ser Gln Val Cys Ala Gly Arg Gly Asp Ser Gly Ala Pro Leu Ile
 260 265 270

Ala Asp Gly Arg Val Val Gly Leu Val Ser Gly Gly Val Ile Pro Asp
 275 280 285

Tyr Asn Leu Ala Cys Ala Thr Pro Leu Gln Gly Pro Phe Phe Met Pro
 290 295 300

Thr Leu Ser Val Asn Met Asp Thr Val Leu Thr Asp Leu Asp Ser Gln
 305 310 315 320

Asp Leu Pro Gly Arg Gly Phe Gln Pro Thr Ala Gly
 325 330

<210> 43
 <211> 2049
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(2026)
 <223> RXN01868

<400> 43
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ccacaaaccc tgtggcggta aatcccctag agtaggccac atg aag gat ctt tat 115
 Met Lys Asp Leu Tyr
 1 5

cgc ttt gtc aat ggc ctg tgg ctt gac acc cac atc att ccc gac gat 163
 Arg Phe Val Asn Gly Leu Trp Leu Asp Thr His Ile Ile Pro Asp Asp
 10 15 20

cgc gcg gtg gac ggc acg ttc cac aag ctg cgc gat gat gct gaa gaa 211
 Arg Ala Val Asp Gly Thr Phe His Lys Leu Arg Asp Asp Ala Glu Glu
 25 30 35

gac gtc cat gag atc gtc aag gaa gac act gga cgc gca ggc aca ctt 259
 Asp Val His Glu Ile Val Lys Glu Asp Thr Gly Arg Ala Gly Thr Leu
 40 45 50

tat gcc tca ttt atg gat act gac gcc atc aac gct gct ggt gtt gca 307

Tyr	Ala	Ser	Phe	Met	Asp	Thr	Asp	Ala	Ile	Asn	Ala	Ala	Gly	Val	Ala		
55						60					65						
ccg	ctc	gat	gcg	gat	ctg	aac	agg	ctg	tct	gtt	gct	aac	tca	tcg	ttt	355	
Pro	Leu	Asp	Ala	Asp	Leu	Asn	Arg	Leu	Ser	Val	Ala	Asn	Ser	Ser	Phe		
70					75					80					85		
ttc	gca	gct	gct	ctc	ggc	gaa	ctg	gac	cgt	gaa	ggc	gtt	ggc	gcg	cca	403	
Phe	Ala	Ala	Ala	Leu	Gly	Glu	Leu	Asp	Arg	Glu	Gly	Val	Gly	Ala	Pro		
				90					95					100			
gta	ggt	ttc	tgg	gtg	gag	aag	gat	tct	tcc	tcc	aac	gaa	tcc	gtc	gcc	451	
Val	Gly	Phe	Trp	Val	Glu	Lys	Asp	Ser	Ser	Ser	Asn	Glu	Ser	Val	Ala		
			105					110					115				
tat	gtc	atc	cag	tcc	ggc	ctc	ggc	ctg	ccc	gat	gag	gct	tat	tac	cgc	499	
Tyr	Val	Ile	Gln	Ser	Gly	Leu	Gly	Leu	Pro	Asp	Glu	Ala	Tyr	Tyr	Arg		
			120				125					130					
gag	gag	gca	cac	gcc	gaa	act	ctc	gcg	gcc	tac	aaa	gag	cac	gtt	gag	547	
Glu	Glu	Ala	His	Ala	Glu	Thr	Leu	Ala	Ala	Tyr	Lys	Glu	His	Val	Glu		
			135			140					145						
cgc	atg	ctc	ggc	tac	ttg	gat	aac	agc	cgc	ctc	ttc	ggt	ctg	tcg	gct	595	
Arg	Met	Leu	Gly	Tyr	Leu	Asp	Asn	Ser	Arg	Leu	Phe	Gly	Leu	Ser	Ala		
150					155					160					165		
gct	tcc	gct	gcc	gca	cga	att	gtc	gcc	ctg	gaa	acg	gaa	atc	gct	gct	643	
Ala	Ser	Ala	Ala	Ala	Arg	Ile	Val	Ala	Leu	Glu	Thr	Glu	Ile	Ala	Ala		
				170					175					180			
ggc	cac	tgg	gat	gtc	gtg	aag	acc	cgc	gac	gcc	gta	gcc	acc	tac	aac	691	
Gly	His	Trp	Asp	Val	Val	Lys	Thr	Arg	Asp	Ala	Val	Ala	Thr	Tyr	Asn		
			185					190					195				
ccc	acc	gaa	ctc	ggc	gcg	ctg	cca	cca	aag	gtc	cgc	acg	ctg	ctc	agt	739	
Pro	Thr	Glu	Leu	Gly	Ala	Leu	Pro	Pro	Lys	Val	Arg	Thr	Leu	Leu	Ser		
			200				205						210				
tcc	gca	ggc	ctc	ccg	gac	cag	cgc	ctg	gta	tcg	atg	atg	ccg	tca	tac	787	
Ser	Ala	Gly	Leu	Pro	Asp	Gln	Arg	Leu	Val	Ser	Met	Met	Pro	Ser	Tyr		
			215				220					225					
ctc	gac	cac	ctc	aac	ggc	ttg	ctt	gtc	gac	gac	cgc	ctc	ccc	gat	tgg	835	
Leu	Asp	His	Leu	Asn	Gly	Leu	Leu	Val	Asp	Asp	Arg	Leu	Pro	Asp	Trp		
230					235					240					245		
cag	cta	tgg	gca	acc	tgg	cac	atc	ttg	agg	tct	cga	gca	gga	ctg	ttg	883	
Gln	Leu	Trp	Ala	Thr	Trp	His	Ile	Leu	Arg	Ser	Arg	Ala	Gly	Leu	Leu		
				250					255					260			
acc	gag	gaa	att	agc	caa	gca	aac	ttc	gac	ttc	tat	ggc	acc	aaa	ctg	931	
Thr	Glu	Glu	Ile	Ser	Gln	Ala	Asn	Phe	Asp	Phe	Tyr	Gly	Thr	Lys	Leu		
			265					270					275				
tcc	ggc	gcc	acc	gag	caa	aaa	gat	cga	tgg	aag	cgt	gct	gtc	ggc	ctg	979	
Ser	Gly	Ala	Thr	Glu	Gln	Lys	Asp	Arg	Trp	Lys	Arg	Ala	Val	Gly	Leu		
			280				285					290					
gca	gag	cgc	atg	gtg	ggc	gag	gaa	atc	ggg	caa	cga	ttc	gtc	gaa	agg	1027	
Ala	Glu	Arg	Met	Val	Gly	Glu	Glu	Ile	Gly	Gln	Arg	Phe	Val	Glu	Arg		

295	300	305	
cat ttt cct gca agc tcc aag gag cac atg ctt gag ctc gtc gac tac His Phe Pro Ala Ser Lys Glu His Met Leu Glu Leu Val Asp Tyr 310 315 320 325			1075
ctg gtt gcc gcc tac cgt gat cgc att tcc aac ctc gaa tgg atg acg Leu Val Ala Ala Tyr Arg Asp Arg Ile Ser Asn Leu Glu Trp Met Thr 330 335 340			1123
ccc gcc acc cgc gag cgt gcc ctg gaa aag ttg ggc aaa ttc aac gcg Pro Ala Thr Arg Glu Arg Ala Leu Glu Lys Leu Gly Lys Phe Asn Ala 345 350 355			1171
aaa atc ggc tac ccc gac aag tgg cgc tcc tac gaa ggc ctc gaa ttc Lys Ile Gly Tyr Pro Asp Lys Trp Arg Ser Tyr Glu Gly Leu Glu Phe 360 365 370			1219
ggc tcc gac ctg gtg gac aac tcc cgc aag ggc tcc gcg ttc ctc cat Gly Ser Asp Leu Val Asp Asn Ser Arg Lys Gly Ser Ala Phe Leu His 375 380 385			1267
gac tat gag ctg ggc aag atc ggc aaa cca gcc gac cgc gac gaa tgg Asp Tyr Glu Leu Gly Lys Ile Gly Lys Pro Ala Asp Arg Asp Glu Trp 390 395 400 405			1315
gtc acc acc cca caa acc gtc aac gcc ttc tac aac ccc gtg gtc aac Val Thr Thr Pro Gln Thr Val Asn Ala Phe Tyr Asn Pro Val Val Asn 410 415 420			1363
gac atc acc ttc ccc gca gcc atc ctg cgc gca cca ttc ttc gac ccc Asp Ile Thr Phe Pro Ala Ala Ile Leu Arg Ala Pro Phe Phe Asp Pro 425 430 435			1411
gaa gca gaa gcc gca gaa aac ttc ggt gca atc ggt gct gtg atc gga Glu Ala Glu Ala Ala Glu Asn Phe Gly Ala Ile Gly Ala Val Ile Gly 440 445 450			1459
cac gaa atc ggc cac ggc ttt gac gat caa ggc agc caa tac gac ggc His Glu Ile Gly His Gly Phe Asp Asp Gln Gly Ser Gln Tyr Asp Gly 455 460 465			1507
gac ggc aac ctc aac tcc tgg tgg acc gac gaa gac cgc tcc gca ttc Asp Gly Asn Leu Asn Ser Trp Trp Thr Asp Glu Asp Arg Ser Ala Phe 470 475 480 485			1555
gag cag ctc acc tca cgt ctg gtc acc caa ttc agc gga ctc gtc cct Glu Gln Leu Thr Ser Arg Leu Val Thr Gln Phe Ser Gly Leu Val Pro 490 495 500			1603
gcc gtc ctg acc tct gaa gga atc gac acc gac ggc gtc aac ggt gaa Ala Val Leu Thr Ser Glu Gly Ile Asp Thr Asp Gly Val Asn Gly Glu 505 510 515			1651
ttc act ctc ggc gaa aac atc ggt gac ctc ggc gga ttg ggc atc gct Phe Thr Leu Gly Glu Asn Ile Gly Asp Leu Gly Gly Leu Gly Ile Ala 520 525 530			1699
gtc gtt gcc tac gaa aag tac ctc gca gac cgt ggc caa acc ttt gaa Val Val Ala Tyr Glu Lys Tyr Leu Ala Asp Arg Gly Gln Thr Phe Glu 535 540 545			1747

acc tca cca gtc caa aaa ttc gaa gca gaa ggc gcc gag gaa ggc ctg 1795
 Thr Ser Pro Val Gln Lys Phe Glu Ala Glu Gly Ala Glu Glu Gly Leu
 550 555 560 565

gcc gag caa gaa ttc aac ggt ctc caa cgc ctc ttc ctg tcc tgg gct 1843
 Ala Glu Gln Glu Phe Asn Gly Leu Gln Arg Leu Phe Leu Ser Trp Ala
 570 575 580

cgc gtg tgg cgc acc aaa atc cgc cca cag atg gcc gtc caa tac ctg 1891
 Arg Val Trp Arg Thr Lys Ile Arg Pro Gln Met Ala Val Gln Tyr Leu
 585 590 595

gcc atc gac cca cac tcc cct gca gaa ttc cgc tgc aat gtc atc gcc 1939
 Ala Ile Asp Pro His Ser Pro Ala Glu Phe Arg Cys Asn Val Ile Ala
 600 605 610

gga aac gtc gct gaa ttc tac gaa gca ttc gac gtc ccc gaa gat gca 1987
 Gly Asn Val Ala Glu Phe Tyr Glu Ala Phe Asp Val Pro Glu Asp Ala
 615 620 625

cct gtg tac atc aag cca gaa gag cgc cta gct atc tgg tagttgttag 2036
 Pro Val Tyr Ile Lys Pro Glu Glu Arg Leu Ala Ile Trp
 630 635 640

ttggtattga aaa 2049

<210> 44

<211> 642

<212> PRT

<213> *Corynebacterium glutamicum*

<400> 44

Met Lys Asp Leu Tyr Arg Phe Val Asn Gly Leu Trp Leu Asp Thr His
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Ile Ile Pro Asp Asp Arg Ala Val Asp Gly Thr Phe His Lys Leu Arg
 20 25 30

Asp Asp Ala Glu Glu Asp Val His Glu Ile Val Lys Glu Asp Thr Gly
 35 40 45

Arg Ala Gly Thr Leu Tyr Ala Ser Phe Met Asp Thr Asp Ala Ile Asn
 50 55 60

Ala Ala Gly Val Ala Pro Leu Asp Ala Asp Leu Asn Arg Leu Ser Val
 65 70 75 80

Ala Asn Ser Ser Phe Phe Ala Ala Ala Leu Gly Glu Leu Asp Arg Glu
 85 90 95

Gly Val Gly Ala Pro Val Gly Phe Trp Val Glu Lys Asp Ser Ser Ser
 100 105 110

Asn Glu Ser Val Ala Tyr Val Ile Gln Ser Gly Leu Gly Leu Pro Asp
 115 120 125

Glu Ala Tyr Tyr Arg Glu Glu Ala His Ala Glu Thr Leu Ala Ala Tyr
 130 135 140

Lys Glu His Val Glu Arg Met Leu Gly Tyr Leu Asp Asn Ser Arg Leu
 145 150 155 160
 Phe Gly Leu Ser Ala Ala Ser Ala Ala Ala Arg Ile Val Ala Leu Glu
 165 170 175
 Thr Glu Ile Ala Ala Gly His Trp Asp Val Val Lys Thr Arg Asp Ala
 180 185 190
 Val Ala Thr Tyr Asn Pro Thr Glu Leu Gly Ala Leu Pro Pro Lys Val
 195 200 205
 Arg Thr Leu Leu Ser Ser Ala Gly Leu Pro Asp Gln Arg Leu Val Ser
 210 215 220
 Met Met Pro Ser Tyr Leu Asp His Leu Asn Gly Leu Leu Val Asp Asp
 225 230 235 240
 Arg Leu Pro Asp Trp Gln Leu Trp Ala Thr Trp His Ile Leu Arg Ser
 245 250 255
 Arg Ala Gly Leu Leu Thr Glu Glu Ile Ser Gln Ala Asn Phe Asp Phe
 260 265 270
 Tyr Gly Thr Lys Leu Ser Gly Ala Thr Glu Gln Lys Asp Arg Trp Lys
 275 280 285
 Arg Ala Val Gly Leu Ala Glu Arg Met Val Gly Glu Glu Ile Gly Gln
 290 295 300
 Arg Phe Val Glu Arg His Phe Pro Ala Ser Ser Lys Glu His Met Leu
 305 310 315 320
 Glu Leu Val Asp Tyr Leu Val Ala Ala Tyr Arg Asp Arg Ile Ser Asn
 325 330 335
 Leu Glu Trp Met Thr Pro Ala Thr Arg Glu Arg Ala Leu Glu Lys Leu
 340 345 350
 Gly Lys Phe Asn Ala Lys Ile Gly Tyr Pro Asp Lys Trp Arg Ser Tyr
 355 360 365
 Glu Gly Leu Glu Phe Gly Ser Asp Leu Val Asp Asn Ser Arg Lys Gly
 370 375 380
 Ser Ala Phe Leu His Asp Tyr Glu Leu Gly Lys Ile Gly Lys Pro Ala
 385 390 395 400
 Asp Arg Asp Glu Trp Val Thr Thr Pro Gln Thr Val Asn Ala Phe Tyr
 405 410 415
 Asn Pro Val Val Asn Asp Ile Thr Phe Pro Ala Ala Ile Leu Arg Ala
 420 425 430
 Pro Phe Phe Asp Pro Glu Ala Glu Ala Ala Glu Asn Phe Gly Ala Ile
 435 440 445
 Gly Ala Val Ile Gly His Glu Ile Gly His Gly Phe Asp Asp Gln Gly
 450 455 460
 Ser Gln Tyr Asp Gly Asp Gly Asn Leu Asn Ser Trp Trp Thr Asp Glu

465		470		475		480
Asp Arg Ser Ala Phe Glu Gln Leu Thr Ser Arg Leu Val Thr Gln Phe	485		490		495	
Ser Gly Leu Val Pro Ala Val Leu Thr Ser Glu Gly Ile Asp Thr Asp	500		505		510	
Gly Val Asn Gly Glu Phe Thr Leu Gly Glu Asn Ile Gly Asp Leu Gly	515		520		525	
Gly Leu Gly Ile Ala Val Val Ala Tyr Glu Lys Tyr Leu Ala Asp Arg	530		535		540	
Gly Gln Thr Phe Glu Thr Ser Pro Val Gln Lys Phe Glu Ala Glu Gly	545		550		555	560
Ala Glu Glu Gly Leu Ala Glu Gln Glu Phe Asn Gly Leu Gln Arg Leu	565		570		575	
Phe Leu Ser Trp Ala Arg Val Trp Arg Thr Lys Ile Arg Pro Gln Met	580		585		590	
Ala Val Gln Tyr Leu Ala Ile Asp Pro His Ser Pro Ala Glu Phe Arg	595		600		605	
Cys Asn Val Ile Ala Gly Asn Val Ala Glu Phe Tyr Glu Ala Phe Asp	610		615		620	
Val Pro Glu Asp Ala Pro Val Tyr Ile Lys Pro Glu Glu Arg Leu Ala	625		630		635	640
Ile Trp						

<210> 45
 <211> 1734
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(1711)
 <223> FRXA01868

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 actggaccgt gaaggcgttg gcgcgccagt aggtttcttg gtg gag aag gat tct 115
 Val Glu Lys Asp Ser 5
 1
 tcc tcc aac gaa tcc gtc gcc tat gtc atc cag tcc ggc ctc ggc ctg 163
 Ser Ser Asn Glu Ser Val Ala Tyr Val Ile Gln Ser Gly Leu Gly Leu 20
 10 15
 ccc gat gag gct tat tac cgc gag gag gca cac gcc gaa act ctc gcg 211
 Pro Asp Glu Ala Tyr Tyr Arg Glu Glu Ala His Ala Glu Thr Leu Ala 35
 25 30

gcc tac aaa gag cac gtt gag cgc atg ctc ggc tac ttg gat aac agc	259
Ala Tyr Lys Glu His Val Glu Arg Met Leu Gly Tyr Leu Asp Asn Ser	
40 45 50	
cgc ctc ttc ggt ctg tcg gct gct tcc gct gcc gca cga att gtc gcc	307
Arg Leu Phe Gly Leu Ser Ala Ala Ser Ala Ala Arg Ile Val Ala	
55 60 65	
ctg gaa acg gaa atc gct gct ggc cac tgg gat gtc gtg aag acc cgc	355
Leu Glu Thr Glu Ile Ala Ala Gly His Trp Asp Val Val Lys Thr Arg	
70 75 80 85	
gac gcc gta gcc acc tac aac ccc acc gaa ctc ggc gcg ctg cca cca	403
Asp Ala Val Ala Thr Tyr Asn Pro Thr Glu Leu Gly Ala Leu Pro Pro	
90 95 100	
aag gtc cgc acg ctg ctc agt tcc gca ggc ctc ccg gac cag cgc ctg	451
Lys Val Arg Thr Leu Leu Ser Ser Ala Gly Leu Pro Asp Gln Arg Leu	
105 110 115	
gta tcg atg atg ccg tca tac ctc gac cac ctc aac ggc ttg ctt gtc	499
Val Ser Met Met Pro Ser Tyr Leu Asp His Leu Asn Gly Leu Leu Val	
120 125 130	
gac gac cgc ctc ccc gat tgg cag cta tgg gca acc tgg cac atc ttg	547
Asp Asp Arg Leu Pro Asp Trp Gln Leu Trp Ala Thr Trp His Ile Leu	
135 140 145	
agg tct cga gca gga ctg ttg acc gag gaa att agc caa gca aac ttc	595
Arg Ser Arg Ala Gly Leu Leu Thr Glu Glu Ile Ser Gln Ala Asn Phe	
150 155 160 165	
gac ttc tat ggc acc aaa ctg tcc ggc gcc acc gag caa aaa gat cga	643
Asp Phe Tyr Gly Thr Lys Leu Ser Gly Ala Thr Glu Gln Lys Asp Arg	
170 175 180	
tgg aag cgt gct gtc ggc ctg gca gag cgc atg gtg ggc gag gaa atc	691
Trp Lys Arg Ala Val Gly Leu Ala Glu Arg Met Val Gly Glu Ile	
185 190 195	
ggg caa cga ttc gtc gaa agg cat ttt cct gca agc tcc aag gag cac	739
Gly Gln Arg Phe Val Glu Arg His Phe Pro Ala Ser Ser Lys Glu His	
200 205 210	
atg ctt gag ctc gtc gac tac ctg gtt gcc gcc tac cgt gat cgc att	787
Met Leu Glu Leu Val Asp Tyr Leu Val Ala Ala Tyr Arg Asp Arg Ile	
215 220 225	
tcc aac ctc gaa tgg atg acg ccc gcc acc cgc gag cgt gcc ctg gaa	835
Ser Asn Leu Glu Trp Met Thr Pro Ala Thr Arg Glu Arg Ala Leu Glu	
230 235 240 245	
aag ttg ggc aaa ttc aac gcg aaa atc ggc tac ccc gac aag tgg cgc	883
Lys Leu Gly Lys Phe Asn Ala Lys Ile Gly Tyr Pro Asp Lys Trp Arg	
250 255 260	
tcc tac gaa ggc ctc gaa ttc ggc tcc gac ctg gtg gac aac tcc cgc	931
Ser Tyr Glu Gly Leu Glu Phe Gly Ser Asp Leu Val Asp Asn Ser Arg	
265 270 275	
aag ggc tcc gcg ttc ctc cat gac tat gag ctg ggc aag atc ggc aaa	979

Lys	Gly	Ser	Ala	Phe	Leu	His	Asp	Tyr	Glu	Leu	Gly	Lys	Ile	Gly	Lys		
	280						285					290					
cca	gcc	gac	cgc	gac	gaa	tgg	gtc	acc	acc	cca	caa	acc	gtc	aac	gcc	1027	
Pro	Ala	Asp	Arg	Asp	Glu	Trp	Val	Thr	Thr	Pro	Gln	Thr	Val	Asn	Ala		
	295					300					305						
ttc	tac	aac	ccc	gtg	gtc	aac	gac	atc	acc	ttc	ccc	gca	gcc	atc	ctg	1075	
Phe	Tyr	Asn	Pro	Val	Val	Asn	Asp	Ile	Thr	Phe	Pro	Ala	Ala	Ile	Leu		
	310				315					320					325		
cgc	gca	cca	ttc	ttc	gac	ccc	gaa	gca	gaa	gcc	gca	gaa	aac	ttc	ggt	1123	
Arg	Ala	Pro	Phe	Phe	Asp	Pro	Glu	Ala	Glu	Ala	Ala	Glu	Asn	Phe	Gly		
				330					335					340			
gca	atc	ggt	gct	gtg	atc	gga	cac	gaa	atc	ggc	cac	ggc	ttt	gac	gat	1171	
Ala	Ile	Gly	Ala	Val	Ile	Gly	His	Glu	Ile	Gly	His	Gly	Phe	Asp	Asp		
			345					350						355			
caa	ggc	agc	caa	tac	gac	ggc	gac	ggc	aac	ctc	aac	tcc	tgg	tgg	acc	1219	
Gln	Gly	Ser	Gln	Tyr	Asp	Gly	Asp	Gly	Asn	Leu	Asn	Ser	Trp	Trp	Thr		
		360					365					370					
gac	gaa	gac	cgc	tcc	gca	ttc	gag	cag	ctc	acc	tca	cgt	ctg	gtc	acc	1267	
Asp	Glu	Asp	Arg	Ser	Ala	Phe	Glu	Gln	Leu	Thr	Ser	Arg	Leu	Val	Thr		
		375				380					385						
caa	ttc	agc	gga	ctc	gtc	cct	gcc	gtc	ctg	acc	tct	gaa	gga	atc	gac	1315	
Gln	Phe	Ser	Gly	Leu	Val	Pro	Ala	Val	Leu	Thr	Ser	Glu	Gly	Ile	Asp		
	390				395				400						405		
acc	gac	ggc	gtc	aac	ggt	gaa	ttc	act	ctc	ggc	gaa	aac	atc	ggt	gac	1363	
Thr	Asp	Gly	Val	Asn	Gly	Glu	Phe	Thr	Leu	Gly	Glu	Asn	Ile	Gly	Asp		
			410					415						420			
ctc	ggc	gga	ttg	ggc	atc	gct	gtc	gtt	gcc	tac	gaa	aag	tac	ctc	gca	1411	
Leu	Gly	Gly	Leu	Gly	Ile	Ala	Val	Val	Ala	Tyr	Glu	Lys	Tyr	Leu	Ala		
			425				430						435				
gac	cgt	ggc	caa	acc	ttt	gaa	acc	tca	cca	gtc	caa	aaa	ttc	gaa	gca	1459	
Asp	Arg	Gly	Gln	Thr	Phe	Glu	Thr	Ser	Pro	Val	Gln	Lys	Phe	Glu	Ala		
		440					445					450					
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Glu	Gly	Ala	Glu	Glu	Gly	Leu	Ala	Glu	Gln	Glu	Phe	Asn	Gly	Leu	Gln		
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cgc	ctc	ttc	ctg	tcc	tgg	gct	cgc	gtg	tgg	cgc	acc	aaa	atc	cgc	cca	1555	
Arg	Leu	Phe	Leu	Ser	Trp	Ala	Arg	Val	Trp	Arg	Thr	Lys	Ile	Arg	Pro		
	470				475					480				485			
cag	atg	gcc	gtc	caa	tac	ctg	gcc	atc	gac	cca	cac	tcc	cct	gca	gaa	1603	
Gln	Met	Ala	Val	Gln	Tyr	Leu	Ala	Ile	Asp	Pro	His	Ser	Pro	Ala	Glu		
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ttc	cgc	tgc	aat	gtc	atc	gcc	gga	aac	gtc	gct	gaa	ttc	tac	gaa	gca	1651	
Phe	Arg	Cys	Asn	Val	Ile	Ala	Gly	Asn	Val	Ala	Glu	Phe	Tyr	Glu	Ala		
			505					510					515				
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525

530

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1734

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 Tyr Leu Asp Asn Ser Arg Leu Phe Gly Leu Ser Ala Ala Ser Ala Ala
 50 55 60
 Ala Arg Ile Val Ala Leu Glu Thr Glu Ile Ala Ala Gly His Trp Asp
 65 70 75 80
 Val Val Lys Thr Arg Asp Ala Val Ala Thr Tyr Asn Pro Thr Glu Leu
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 Gly Ala Leu Pro Lys Val Arg Thr Leu Leu Ser Ser Ala Gly Leu
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 Pro Asp Gln Arg Leu Val Ser Met Met Pro Ser Tyr Leu Asp His Leu
 115 120 125
 Asn Gly Leu Leu Val Asp Asp Arg Leu Pro Asp Trp Gln Leu Trp Ala
 130 135 140
 Thr Trp His Ile Leu Arg Ser Arg Ala Gly Leu Leu Thr Glu Glu Ile
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 Ser Gln Ala Asn Phe Asp Phe Tyr Gly Thr Lys Leu Ser Gly Ala Thr
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 Glu Gln Lys Asp Arg Trp Lys Arg Ala Val Gly Leu Ala Glu Arg Met
 180 185 190
 Val Gly Glu Glu Ile Gly Gln Arg Phe Val Glu Arg His Phe Pro Ala
 195 200 205
 Ser Ser Lys Glu His Met Leu Glu Leu Val Asp Tyr Leu Val Ala Ala
 210 215 220
 Tyr Arg Asp Arg Ile Ser Asn Leu Glu Trp Met Thr Pro Ala Thr Arg
 225 230 235 240
 Glu Arg Ala Leu Glu Lys Leu Gly Lys Phe Asn Ala Lys Ile Gly Tyr
 245 250 255

Pro Asp Lys Trp Arg Ser Tyr Glu Gly Leu Glu Phe Gly Ser Asp Leu
 260 265 270
 Val Asp Asn Ser Arg Lys Gly Ser Ala Phe Leu His Asp Tyr Glu Leu
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 Gly Lys Ile Gly Lys Pro Ala Asp Arg Asp Glu Trp Val Thr Thr Pro
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 Gln Thr Val Asn Ala Phe Tyr Asn Pro Val Val Asn Asp Ile Thr Phe
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 Pro Ala Ala Ile Leu Arg Ala Pro Phe Phe Asp Pro Glu Ala Glu Ala
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 Ala Glu Asn Phe Gly Ala Ile Gly Ala Val Ile Gly His Glu Ile Gly
 340 345 350
 His Gly Phe Asp Asp Gln Gly Ser Gln Tyr Asp Gly Asp Gly Asn Leu
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 Asn Ser Trp Trp Thr Asp Glu Asp Arg Ser Ala Phe Glu Gln Leu Thr
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 Ser Arg Leu Val Thr Gln Phe Ser Gly Leu Val Pro Ala Val Leu Thr
 385 390 395 400
 Ser Glu Gly Ile Asp Thr Asp Gly Val Asn Gly Glu Phe Thr Leu Gly
 405 410 415
 Glu Asn Ile Gly Asp Leu Gly Gly Leu Gly Ile Ala Val Val Ala Tyr
 420 425 430
 Glu Lys Tyr Leu Ala Asp Arg Gly Gln Thr Phe Glu Thr Ser Pro Val
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 Gln Lys Phe Glu Ala Glu Gly Ala Glu Glu Gly Leu Ala Glu Gln Glu
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 Phe Asn Gly Leu Gln Arg Leu Phe Leu Ser Trp Ala Arg Val Trp Arg
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 Thr Lys Ile Arg Pro Gln Met Ala Val Gln Tyr Leu Ala Ile Asp Pro
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 His Ser Pro Ala Glu Phe Arg Cys Asn Val Ile Ala Gly Asn Val Ala
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                                         Met Lys Asp Leu Tyr
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Arg Phe Val Asn Gly Leu Trp Leu Asp Thr His Ile Ile Pro Asp Asp
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Arg Ala Val Asp Gly Thr Phe His Lys Leu Arg Asp Asp Ala Glu Glu
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gac gtc cat gag atc gtc aag gaa gac act gga cgc gca ggc aca ctt 259
Asp Val His Glu Ile Val Lys Glu Asp Thr Gly Arg Ala Gly Thr Leu
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tat gcc tca ttt atg gat act gac gcc atc aac gct gct ggt gtt gca 307
Tyr Ala Ser Phe Met Asp Thr Asp Ala Ile Asn Ala Ala Gly Val Ala
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ccg ctc gat gcg gat ctg aac agg ctg tct gtt gct aac tca tcg ttt 355
Pro Leu Asp Ala Asp Leu Asn Arg Leu Ser Val Ala Asn Ser Ser Phe
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Arg Ala Gly Thr Leu Tyr Ala Ser Phe Met Asp Thr Asp Ala Ile Asn
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Ala Ala Gly Val Ala Pro Leu Asp Ala Asp Leu Asn Arg Leu Ser Val
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Met Phe Glu Arg Phe
1 5
acc gat cgt gca cgc cgc gtg att gtg ctc gcg cag gaa gag gcg cgc 163
Thr Asp Arg Ala Arg Val Ile Val Leu Ala Gln Glu Glu Ala Arg
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atg ctc aac cac aat tac atc ggc acg gag cac att ctc ctc ggc ctc 211
Met Leu Asn His Asn Tyr Ile Gly Thr Glu His Ile Leu Leu Gly Leu
25 30 35
att cac gag ggc gag ggc gtt gca gcc aag gct ttg gaa tcc atg gga 259
Ile His Glu Gly Glu Gly Val Ala Ala Lys Ala Leu Glu Ser Met Gly
40 45 50
att tcc ctg gac gcc gtc cgc cag gaa gtc gaa gag att atc ggc cag 307
Ile Ser Leu Asp Ala Val Arg Gln Glu Val Glu Glu Ile Ile Gly Gln
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ggc tcc cag ccc acc acc ggc cac att cct ttt act cca cgt gcc aag 355
Gly Ser Gln Pro Thr Thr Gly His Ile Pro Phe Thr Pro Arg Ala Lys
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aag gtc ctg gag ctc agc ctc cgc gaa ggc cta caa atg gga cac aag 403
Lys Val Leu Glu Leu Ser Leu Arg Glu Gly Leu Gln Met Gly His Lys
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tac atc ggt act gag ttc ctg ctt ctc ggt ttg atc cgt gag ggc gag 451
Tyr Ile Gly Thr Glu Phe Leu Leu Leu Gly Leu Ile Arg Glu Gly Glu
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Gly Val Ala Ala Gln Val Leu Val Lys Leu Gly Ala Asp Leu Pro Arg
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Val Arg Gln Gln Val Ile Gln Leu Leu Ser Gly Tyr Glu Gly Gly Gln
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Gly Gly Ser Pro Glu Gly Gly Gln Gly Ala Pro Thr Gly Gly Asp Ala
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Val Gly Ala Gly Ala Ala Pro Gly Gly Arg Pro Ser Ser Gly Ser Pro	
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Gly Glu Arg Ser Thr Ser Leu Val Leu Asp Gln Phe Gly Arg Asn Leu	
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Thr Gln Ala Ala Lys Asp Gly Lys Leu Asp Pro Val Val Gly Arg Asp	
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Lys Glu Ile Glu Arg Ile Met Gln Val Leu Ser Arg Arg Thr Lys Asn	
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Asn Pro Val Leu Ile Gly Glu Pro Gly Val Gly Lys Thr Ala Val Val	
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Glu Gly Leu Ala Leu Asp Ile Val Asn Gly Lys Val Pro Glu Thr Leu	
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Lys Asp Lys Gln Val Tyr Ser Leu Asp Leu Gly Ser Leu Val Ala Gly	
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Ser Arg Tyr Arg Gly Asp Phe Glu Glu Arg Leu Lys Lys Val Leu Lys	
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Glu Ile Asn Gln Arg Gly Asp Ile Ile Leu Phe Ile Asp Glu Ile His	
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Thr Leu Val Gly Ala Gly Ala Ala Glu Gly Ala Ile Asp Ala Ala Ser	
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Leu Leu Lys Pro Lys Leu Ala Arg Gly Glu Leu Gln Thr Ile Gly Ala	
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Glu Arg Arg Phe Gln Pro Val Gln Val Pro Glu Pro Ser Val Asp Leu	
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Thr Val Glu Ile Leu Lys Gly Leu Arg Asp Arg Tyr Glu Ala His His	
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390 395 400 405	

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atc gag aag gcc cac aag gag atc tac aac acc ttg ctg cag gtg ttg	2083

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aac	acc	gtc	ctg	atc	ttc	acc	tcc	aac	ctg	ggc	acc	gct	gac	atc	tcc	2179
Asn	Thr	Val	Leu	Ile	Phe	Thr	Ser	Asn	Leu	Gly	Thr	Ala	Asp	Ile	Ser	
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Lys	Ala	Val	Gly	Leu	Gly	Phe	Ser	Gly	Ser	Ser	Glu	Thr	Asp	Ser	Asp	
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Ala	Gln	Tyr	Asp	Arg	Met	Lys	Asn	Lys	Val	His	Asp	Glu	Leu	Lys	Lys	
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His	Phe	Arg	Pro	Glu	Phe	Leu	Asn	Arg	Ile	Asp	Glu	Ile	Val	Val	Phe	
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His	Gln	Leu	Thr	Lys	Asp	Gln	Ile	Val	Gln	Met	Val	Asp	Leu	Leu	Ile	
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Thr	Glu	Lys	Ala	Lys	Asp	Leu	Leu	Ala	Asn	Arg	Gly	Phe	Asp	Pro	Val	
775						780					785					
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Gln	Met	Ser	Glu	Lys	Ile	Leu	Phe	Gly	Glu	Ile	Gly	Ala	Gly	Glu	Ile	
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Val	Thr	Val	Asp	Val	Glu	Gly	Trp	Asp	Gly	Glu	Ser	Lys	Asp	Thr	Asp	
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Lys	Phe	Ser	Glu	Ile	Ser	Val	Glu	Ala	Ala	Glu	Ala	Ile	Gln	Asp	Val	
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gat	tct	gca	gct	gac	ggc	gat	gtc	cca	gaa	acc	gat	tca	ctt	tcc	gac	2755
Asp	Ser	Ala	Ala	Asp	Gly	Asp	Val	Pro	Glu	Thr	Asp	Ser	Leu	Ser	Asp	
870					875				880					885		
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Leu Glu Ser Met Gly Ile Ser Leu Asp Ala Val Arg Gln Glu Val Glu
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Glu Ile Ile Gly Gln Gly Ser Gln Pro Thr Thr Gly His Ile Pro Phe
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Thr Pro Arg Ala Lys Lys Val Leu Glu Leu Ser Leu Arg Glu Gly Leu
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Gln Met Gly His Lys Tyr Ile Gly Thr Glu Phe Leu Leu Leu Gly Leu
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Ile Arg Glu Gly Glu Gly Val Ala Ala Gln Val Leu Val Lys Leu Gly
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Thr Gly Gly Asp Ala Val Gly Ala Gly Ala Ala Pro Gly Gly Arg Pro
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Ser Ser Gly Ser Pro Gly Glu Arg Ser Thr Ser Leu Val Leu Asp Gln
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 Lys Lys Val Leu Lys Glu Ile Asn Gln Arg Gly Asp Ile Ile Leu Phe
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 Met Glu Glu Glu Leu His Lys Arg Ile Ile Gly Gln Asp Glu Ala Val
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Pro Lys Arg Pro Ser Gly Ser Phe Ile Phe Ala Gly Pro Ser Gly Val
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 Thr Ala Ser Arg Leu Phe Gly Ala Pro Pro Gly Tyr Val Gly Tyr Glu
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 Asp Glu Leu Lys Lys His Phe Arg Pro Glu Phe Leu Asn Arg Ile Asp
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 Glu Ile Val Val Phe His Gln Leu Thr Lys Asp Gln Ile Val Gln Met
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 Val Asp Leu Leu Ile Gly Arg Val Ser Asn Ala Leu Ala Glu Lys Asp
 755 760 765
 Met Ser Ile Glu Leu Thr Glu Lys Ala Lys Asp Leu Leu Ala Asn Arg
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 805 810 815
 Gly Ala Gly Glu Ile Val Thr Val Asp Val Glu Gly Trp Asp Gly Glu
 820 825 830
 Ser Lys Asp Thr Asp Arg Ala Lys Phe Thr Phe Thr Pro Arg Pro Lys
 835 840 845
 Pro Met Pro Glu Gly Lys Phe Ser Glu Ile Ser Val Glu Ala Ala Glu
 850 855 860
 Ala Ile Gln Asp Val Asp Ser Ala Ala Asp Gly Asp Val Pro Glu Thr
 865 870 875 880
 Asp Ser Leu Ser Asp Ile Asp Leu Glu Thr Leu Glu Lys Phe Glu Glu

72

ggc gga tcc cca gag ggc ggc cag ggc gcc cct act ggc ggt gac gct 595
 Gly Gly Ser Pro Glu Gly Gly Gln Gly Ala Pro Thr Gly Gly Asp Ala 165
 150 155 160

gtt ggt gca gga gct gct cct ggc ggt cgt cca tct tcg ggc agc cca 643
 Val Gly Ala Gly Ala Ala Pro Gly Gly Arg Pro Ser Ser Gly Ser Pro 180
 170 175

ggc gag cgt tct acc tct ttg gtc ctt gac cag ttc ggc cgc aac ctc 691
 Gly Glu Arg Ser Thr Ser Leu Val Leu Asp Gln Phe Gly Arg Asn Leu 195
 185 190

acc cag gct gca aag gac ggc aag ctg gat cca gtt gtt ggt cgc gat 739
 Thr Gln Ala Ala Lys Asp Gly Lys Leu Asp Pro Val Val Gly Arg Asp 210
 200 205

aag gaa atc gag cgc atc atg cag gtg ctc tcc cgt cgt acc aag aac 787
 Lys Glu Ile Glu Arg Ile Met Gln Val Leu Ser Arg Arg Thr Lys Asn 225
 215 220

aac cca gtt ctt att ggt gag cca ggt gtt ggt aag acc gca gtt gtt 835
 Asn Pro Val Leu Ile Gly Glu Pro Gly Val Gly Lys Thr Ala Val Val 245
 230 235 240

gaa ggt ctt gca cta gac att gtt aac ggc aag gtt cca gag acc ctc 883
 Glu Gly Leu Ala Leu Asp Ile Val Asn Gly Lys Val Pro Glu Thr Leu 260
 250 255

aag gac aag cag gtt tac tcc ctt gac tta ggt tcc ctg gtt gca ggt 931
 Lys Asp Lys Gln Val Tyr Ser Leu Asp Leu Gly Ser Leu Val Ala Gly 275
 265 270

tcc cgt tac cgc ggt gac ttc gaa gag cga ctg aag aag gtc ctc aag 979
 Ser Arg Tyr Arg Gly Asp Phe Glu Glu Arg Leu Lys Lys Val Leu Lys 290
 280 285

gag att aac cag cgc ggc gac atc atc ctg ttt atc gat gag atc cac 1027
 Glu Ile Asn Gln Arg Gly Asp Ile Ile Leu Phe Ile Asp Glu Ile His 305
 295 300

acc ctc gtg ggt gca ggt gca gca cga agg cgc aat cga cgc tgc ctc 1075
 Thr Leu Val Gly Ala Gly Ala Ala Arg Arg Arg Asn Arg Arg Cys Leu 325
 310 315 320

cct gct taagccaaag cttgcccgcg gtg 1104
 Pro Ala

<210> 52
 <211> 327
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 52
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 1 5 10 15
 Gln Glu Glu Ala Arg Met Leu Asn His Asn Tyr Ile Gly Thr Glu His
 20 25 30

Ile Leu Leu Gly Leu Ile His Glu Gly Glu Gly Val Ala Ala Lys Ala
 35 40 45
 Leu Glu Ser Met Gly Ile Ser Leu Asp Ala Val Arg Gln Glu Val Glu
 50 55 60
 Glu Ile Ile Gly Gln Gly Ser Gln Pro Thr Thr Gly His Ile Pro Phe
 65 70 75 80
 Thr Pro Arg Ala Lys Lys Val Leu Glu Leu Ser Leu Arg Glu Gly Leu
 85 90 95
 Gln Met Gly His Lys Tyr Ile Gly Thr Glu Phe Leu Leu Leu Gly Leu
 100 105 110
 Ile Arg Glu Gly Glu Gly Val Ala Ala Gln Val Leu Val Lys Leu Gly
 115 120 125
 Ala Asp Leu Pro Arg Val Arg Gln Gln Val Ile Gln Leu Leu Ser Gly
 130 135 140
 Tyr Glu Gly Gly Gln Gly Gly Ser Pro Glu Gly Gly Gln Gly Ala Pro
 145 150 155 160
 Thr Gly Gly Asp Ala Val Gly Ala Gly Ala Ala Pro Gly Gly Arg Pro
 165 170 175
 Ser Ser Gly Ser Pro Gly Glu Arg Ser Thr Ser Leu Val Leu Asp Gln
 180 185 190
 Phe Gly Arg Asn Leu Thr Gln Ala Ala Lys Asp Gly Lys Leu Asp Pro
 195 200 205
 Val Val Gly Arg Asp Lys Glu Ile Glu Arg Ile Met Gln Val Leu Ser
 210 215 220
 Arg Arg Thr Lys Asn Asn Pro Val Leu Ile Gly Glu Pro Gly Val Gly
 225 230 235 240
 Lys Thr Ala Val Val Glu Gly Leu Ala Leu Asp Ile Val Asn Gly Lys
 245 250 255
 Val Pro Glu Thr Leu Lys Asp Lys Gln Val Tyr Ser Leu Asp Leu Gly
 260 265 270
 Ser Leu Val Ala Gly Ser Arg Tyr Arg Gly Asp Phe Glu Glu Arg Leu
 275 280 285
 Lys Lys Val Leu Lys Glu Ile Asn Gln Arg Gly Asp Ile Ile Leu Phe
 290 295 300
 Ile Asp Glu Ile His Thr Leu Val Gly Ala Gly Ala Ala Arg Arg Arg
 305 310 315 320
 Asn Arg Arg Cys Leu Pro Ala
 325

<210> 53
 <211> 1956

<212> DNA
 <213> *Corynebacterium glutamicum*

<220>
 <221> CDS
 <222> (101)..(1933)
 <223> FRXA02471

<400> 53
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 atcctgttta tcgatgagat ccacaccctc gtgggtgcag gtg cag cac gaa ggc 115
 Val Gln His Glu Gly
 1 5
 gca atc gac gct gcc tcc ctg ctt aag cca aag ctt gcc cgc ggt gaa 163
 Ala Ile Asp Ala Ala Ser Leu Leu Lys Pro Lys Leu Ala Arg Gly Glu
 10 15 20
 ctg cag acc att ggt gca acc acc ctg gat gag tac cgt aag cac att 211
 Leu Gln Thr Ile Gly Ala Thr Thr Leu Asp Glu Tyr Arg Lys His Ile
 25 30 35
 gaa aag gac gca gct ctt gag cgt cgt ttc cag cca gtg cag gtt cca 259
 Glu Lys Asp Ala Ala Leu Glu Arg Arg Phe Gln Pro Val Gln Val Pro
 40 45 50
 gag cct tcg gtt gat ctc acc gtt gag atc ttg aag ggt ctg cgc gac 307
 Glu Pro Ser Val Asp Leu Thr Val Glu Ile Leu Lys Gly Leu Arg Asp
 55 60 65
 cgc tac gaa gct cac cac cgc gta tcc atc acc gat ggt gct ctt act 355
 Arg Tyr Glu Ala His His Arg Val Ser Ile Thr Asp Gly Ala Leu Thr
 70 75 80 85
 gca gca gct cag ctt gct gat cgc tac atc aac gac cgc ttc ttg cca 403
 Ala Ala Ala Gln Leu Ala Asp Arg Tyr Ile Asn Asp Arg Phe Leu Pro
 90 95 100
 gat aag gcc gtt gac ctc atc gat gag gct ggc gcc cgc atg cgc atc 451
 Asp Lys Ala Val Asp Leu Ile Asp Glu Ala Gly Ala Arg Met Arg Ile
 105 110 115
 aag cgc atg acc gca cct tcc tcc ctc cgc gag gtt gat gag cgt atc 499
 Lys Arg Met Thr Ala Pro Ser Ser Leu Arg Glu Val Asp Glu Arg Ile
 120 125 130
 gct gat gtt cgc cgt gag aag gaa gca gcg atc gat gct cag gac ttt 547
 Ala Asp Val Arg Arg Glu Lys Glu Ala Ala Ile Asp Ala Gln Asp Phe
 135 140 145
 gag aag gca gca ggt ctt cgc gat aag gag cgc aag ctc ggc gaa gag 595
 Glu Lys Ala Ala Gly Leu Arg Asp Lys Glu Arg Lys Leu Gly Glu Glu
 150 155 160 165
 cgt tca gag aag gaa aag cag tgg cgc tcc ggc gac ctc gag gac atc 643
 Arg Ser Glu Lys Glu Lys Gln Trp Arg Ser Gly Asp Leu Glu Asp Ile
 170 175 180
 gct gag gtt ggc gaa gag cag atc gca gaa gta ctg gcc aac tgg act 691
 Ala Glu Val Gly Glu Glu Gln Ile Ala Glu Val Leu Ala Asn Trp Thr

185	190	195	
ggt att cct gtc ttc aag ctc acc Gly Ile Pro Val Phe Lys Leu 200	gaa gct gaa tct tca cgc ctg ctc Glu Ala Glu Ser Ser Arg Leu Leu 205		739
aac atg gaa gaa gag ttg cac aag cgc atc atc Asn Met Glu Glu Glu Leu His Lys Arg Ile Ile 215	gga cag gat gaa gct Gly Gln Asp Glu Ala 225		787
gtc aag gct gtc tcc cgt gcg atc cgt cgt acc cgt gca ggt ctg aag Val Lys Ala Val Ser Arg Ala Ile Arg Arg Thr Arg Ala Gly Leu Lys 230 235 240 245			835
gat cct aag cgt cct tcc gcc tcc ttc atc ttc gct ggt cca tcc gcc Asp Pro Lys Arg Pro Ser Gly Ser Phe Ile Phe Ala Gly Pro Ser Gly 250 255 260			883
gtt ggt aag acc gag ctg tcc aag gct ctc gca gga ttc ctc ttc ggt Val Gly Lys Thr Glu Leu Ser Lys Ala Leu Ala Gly Phe Leu Phe Gly 265 270 275			931
gac gat gat tcc ctc atc caa atc gac atg ggt gag ttc cac gac cgc Asp Asp Asp Ser Leu Ile Gln Ile Asp Met Gly Glu Phe His Asp Arg 280 285 290			979
ttc acc gcg tcc cga ctt ttc ggt gcc cct ccg gga tac gtt gcc tac Phe Thr Ala Ser Arg Leu Phe Gly Ala Pro Pro Gly Tyr Val Gly Tyr 295 300 305			1027
gaa gaa ggt ggc cag ctg acc gag aag gtt cgc cgt aag cca ttc tcc Glu Glu Gly Gly Gln Leu Thr Glu Lys Val Arg Arg Lys Pro Phe Ser 310 315 320 325			1075
gtt gtg ctt ttc gac gaa atc gag aag gcc cac aag gag atc tac aac Val Val Leu Phe Asp Glu Ile Glu Lys Ala His Lys Glu Ile Tyr Asn 330 335 340			1123
acc ttg ctg cag gtg ttg gaa gat ggt cgc ctt acc gat ggt cag gga Thr Leu Leu Gln Val Leu Glu Asp Gly Arg Leu Thr Asp Gly Gln Gly 345 350 355			1171
cgc atc gtg gac ttc aag aac acc gtc ctg atc ttc acc tcc aac ctg Arg Ile Val Asp Phe Lys Asn Thr Val Leu Ile Phe Thr Ser Asn Leu 360 365 370			1219
ggc acc gct gac atc tcc aag gct gtt ggc ctg ggc ttc tcc gga tcc Gly Thr Ala Asp Ile Ser Lys Ala Val Gly Leu Gly Phe Ser Gly Ser 375 380 385			1267
tcc gag act gac agc gat gct cag tac gac cgc atg aag aac aag gtc Ser Glu Thr Asp Ser Asp Ala Gln Tyr Asp Arg Met Lys Asn Lys Val 390 395 400 405			1315
cac gac gag ctg aag aag cac ttc cgc cct gag ttc ctg aac cgt att His Asp Glu Leu Lys Lys His Phe Arg Pro Glu Phe Leu Asn Arg Ile 410 415 420			1363
gat gag atc gtg gtc ttc cac cag ctc acc aag gat cag atc gtt cag Asp Glu Ile Val Val Phe His Gln Leu Thr Lys Asp Gln Ile Val Gln 425 430 435			1411

atg gtc gac ctt ctt atc ggt cgc gtt tcc aac gca ctg gct gag aag 1459
 Met Val Asp Leu Leu Ile Gly Arg Val Ser Asn Ala Leu Ala Glu Lys
 440 445 450

gac atg agc atc gaa ctg act gag aag gcc aag gac ctc ctg gct aac 1507
 Asp Met Ser Ile Glu Leu Thr Glu Lys Ala Lys Asp Leu Leu Ala Asn
 455 460 465

cga ggc ttc gat cca gtt ctg ggt gca cga cca ttg cgt cgc acc atc 1555
 Arg Gly Phe Asp Pro Val Leu Gly Ala Arg Pro Leu Arg Arg Thr Ile
 470 475 480 485

cag cgc gaa att gaa gac cag atg tcc gag aag atc ctc ttc ggt gaa 1603
 Gln Arg Glu Ile Glu Asp Gln Met Ser Glu Lys Ile Leu Phe Gly Glu
 490 495 500

atc ggc gca ggc gag atc gtc acc gtt gac gtc gaa ggc tgg gac ggc 1651
 Ile Gly Ala Gly Glu Ile Val Thr Val Asp Val Glu Gly Trp Asp Gly
 505 510 515

gag tcc aag gac acc gac cgt cgc aag ttc acc ttc aca cca cgt cca 1699
 Glu Ser Lys Asp Thr Asp Arg Ala Lys Phe Thr Phe Thr Pro Arg Pro
 520 525 530

aag cca atg cca gaa ggt aag ttc tct gag atc tct gtc gag gct gcg 1747
 Lys Pro Met Pro Glu Gly Lys Phe Ser Glu Ile Ser Val Glu Ala Ala
 535 540 545

gaa gca att caa gat gta gat tct gca gct gac ggc gat gtc cca gaa 1795
 Glu Ala Ile Gln Asp Val Asp Ser Ala Ala Asp Gly Asp Val Pro Glu
 550 555 560 565

acc gat tca ctt tcc gac att gac ctt gaa acc ctt gaa aag ttt gag 1843
 Thr Asp Ser Leu Ser Asp Ile Asp Leu Glu Thr Leu Glu Lys Phe Glu
 570 575 580

gaa gat gta gaa aac ggc acc gac att gat cag gtg tcc ggt gac tac 1891
 Glu Asp Val Glu Asn Gly Thr Asp Ile Asp Gln Val Ser Gly Asp Tyr
 585 590 595

tac ggc acc gat gat cag gga ggc act gct cca agc aag gag 1933
 Tyr Gly Thr Asp Asp Gln Gly Gly Thr Ala Pro Ser Lys Glu
 600 605 610

tagcaacctt ttgaaaaagg gcc 1956

<210> 54
 <211> 611
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 54
 Val Gln His Glu Gly Ala Ile Asp Ala Ala Ser Leu Leu Lys Pro Lys
 1 5 10 15
 Leu Ala Arg Gly Glu Leu Gln Thr Ile Gly Ala Thr Thr Leu Asp Glu
 20 25 30
 Tyr Arg Lys His Ile Glu Lys Asp Ala Ala Leu Glu Arg Arg Phe Gln

35	40	45
Pro Val Gln Val Pro Glu Pro Ser Val Asp Leu Thr Val Glu Ile Leu		
50	55	60
Lys Gly Leu Arg Asp Arg Tyr Glu Ala His His Arg Val Ser Ile Thr		
65	70	75
Asp Gly Ala Leu Thr Ala Ala Ala Gln Leu Ala Asp Arg Tyr Ile Asn		
85	90	95
Asp Arg Phe Leu Pro Asp Lys Ala Val Asp Leu Ile Asp Glu Ala Gly		
100	105	110
Ala Arg Met Arg Ile Lys Arg Met Thr Ala Pro Ser Ser Leu Arg Glu		
115	120	125
Val Asp Glu Arg Ile Ala Asp Val Arg Arg Glu Lys Glu Ala Ala Ile		
130	135	140
Asp Ala Gln Asp Phe Glu Lys Ala Ala Gly Leu Arg Asp Lys Glu Arg		
145	150	155
Lys Leu Gly Glu Glu Arg Ser Glu Lys Glu Lys Gln Trp Arg Ser Gly		
165	170	175
Asp Leu Glu Asp Ile Ala Glu Val Gly Glu Glu Gln Ile Ala Glu Val		
180	185	190
Leu Ala Asn Trp Thr Gly Ile Pro Val Phe Lys Leu Thr Glu Ala Glu		
195	200	205
Ser Ser Arg Leu Leu Asn Met Glu Glu Glu Leu His Lys Arg Ile Ile		
210	215	220
Gly Gln Asp Glu Ala Val Lys Ala Val Ser Arg Ala Ile Arg Arg Thr		
225	230	235
Arg Ala Gly Leu Lys Asp Pro Lys Arg Pro Ser Gly Ser Phe Ile Phe		
245	250	255
Ala Gly Pro Ser Gly Val Gly Lys Thr Glu Leu Ser Lys Ala Leu Ala		
260	265	270
Gly Phe Leu Phe Gly Asp Asp Asp Ser Leu Ile Gln Ile Asp Met Gly		
275	280	285
Glu Phe His Asp Arg Phe Thr Ala Ser Arg Leu Phe Gly Ala Pro Pro		
290	295	300
Gly Tyr Val Gly Tyr Glu Glu Gly Gly Gln Leu Thr Glu Lys Val Arg		
305	310	315
Arg Lys Pro Phe Ser Val Val Leu Phe Asp Glu Ile Glu Lys Ala His		
325	330	335
Lys Glu Ile Tyr Asn Thr Leu Leu Gln Val Leu Glu Asp Gly Arg Leu		
340	345	350
Thr Asp Gly Gln Gly Arg Ile Val Asp Phe Lys Asn Thr Val Leu Ile		
355	360	365

Phe Thr Ser Asn Leu Gly Thr Ala Asp Ile Ser Lys Ala Val Gly Leu
 370 375 380
 Gly Phe Ser Gly Ser Ser Glu Thr Asp Ser Asp Ala Gln Tyr Asp Arg
 385 390 395 400
 Met Lys Asn Lys Val His Asp Glu Leu Lys Lys His Phe Arg Pro Glu
 405 410 415
 Phe Leu Asn Arg Ile Asp Glu Ile Val Val Phe His Gln Leu Thr Lys
 420 425 430
 Asp Gln Ile Val Gln Met Val Asp Leu Leu Ile Gly Arg Val Ser Asn
 435 440 445
 Ala Leu Ala Glu Lys Asp Met Ser Ile Glu Leu Thr Glu Lys Ala Lys
 450 455 460
 Asp Leu Leu Ala Asn Arg Gly Phe Asp Pro Val Leu Gly Ala Arg Pro
 465 470 475 480
 Leu Arg Arg Thr Ile Gln Arg Glu Ile Glu Asp Gln Met Ser Glu Lys
 485 490 495
 Ile Leu Phe Gly Glu Ile Gly Ala Gly Glu Ile Val Thr Val Asp Val
 500 505 510
 Glu Gly Trp Asp Gly Glu Ser Lys Asp Thr Asp Arg Ala Lys Phe Thr
 515 520 525
 Phe Thr Pro Arg Pro Lys Pro Met Pro Glu Gly Lys Phe Ser Glu Ile
 530 535 540
 Ser Val Glu Ala Ala Glu Ala Ile Gln Asp Val Asp Ser Ala Ala Asp
 545 550 555 560
 Gly Asp Val Pro Glu Thr Asp Ser Leu Ser Asp Ile Asp Leu Glu Thr
 565 570 575
 Leu Glu Lys Phe Glu Glu Asp Val Glu Asn Gly Thr Asp Ile Asp Gln
 580 585 590
 Val Ser Gly Asp Tyr Tyr Gly Thr Asp Asp Gln Gly Gly Thr Ala Pro
 595 600 605
 Ser Lys Glu .
 610

<210> 55
 <211> 1446
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(1423)
 <223> RXA02630

<400> 55

gcgggttcga aaatgtcgtat gattaaacca ctaaagagct cacaggaagt gttcagacta 60

cttagagtga cgccccagcc acagggttca taatcaaata atg aca aat caa ttc 115
Met Thr Asn Gln Phe
1 5

ccc aca aac aac ggt gag aac ccg gac cgt gca tcg gaa act cca tca 163
Pro Thr Asn Asn Gly Glu Asn Pro Asp Arg Ala Ser Glu Thr Pro Ser
10 15 20

gaa acc aac tcc ttc gaa cat gtg cgt agt tca tat ccg cag tgg ggt 211
Glu Thr Asn Ser Phe Glu His Val Arg Ser Ser Tyr Pro Gln Trp Gly
25 30 35

aac act gct tcc aat caa aac ccc tat cct ggt gcg gcc ttc gcc tct 259
Asn Thr Ala Ser Asn Gln Asn Pro Tyr Pro Gly Ala Gly Phe Gly Ser
40 45 50

gaa caa aac act caa caa gga aat gag caa caa gct cca gcc tgg acc 307
Glu Gln Asn Thr Gln Gln Gly Asn Glu Gln Gln Ala Pro Ala Trp Thr
55 60 65

agt tgg gat aat cag cct cta agc aca gat gta aag cca gct aaa gaa 355
Ser Trp Asp Asn Gln Pro Leu Ser Thr Asp Val Lys Pro Ala Lys Glu
70 75 80 85

aag cga aaa gtt gcc atc gga acg gca etc gcg tta atg ctt gtt ggt 403
Lys Arg Lys Val Gly Ile Gly Thr Ala Leu Ala Leu Met Leu Val Gly
90 95 100

tct att gct acc ggt agc gtt gtg ggt gtt gca gca acc cag ctt ggt 451
Ser Ile Ala Thr Gly Ser Val Val Gly Val Ala Ala Thr Gln Leu Gly
105 110 115

tcg gac tct tca acc cca gtt aat gct ctt gag cag ccc agc gtg cag 499
Ser Asp Ser Ser Thr Pro Val Asn Ala Leu Glu Gln Pro Ser Val Gln
120 125 130

cgc acc act aat gct gaa cca ggt tca gcg gaa cag gtt gct gcc gca 547
Arg Thr Thr Asn Ala Glu Pro Gly Ser Ala Glu Gln Val Ala Ala Ala
135 140 145

gtt ttg cct tct gtc gtc tct att cag gcc att act agg acg tct gct 595
Val Leu Pro Ser Val Val Ser Ile Gln Ala Ile Thr Arg Thr Ser Ala
150 155 160 165

tct gag gcc tct gga tcc att att tcc tct gat ggt tac gtc atg acc 643
Ser Glu Gly Ser Gly Ser Ile Ile Ser Ser Asp Gly Tyr Val Met Thr
170 175 180

aat aat cac gtc gtg gca gcc att gaa caa tct ggt gtg tta gaa gta 691
Asn Asn His Val Val Ala Gly Ile Glu Gln Ser Gly Val Leu Glu Val
185 190 195

agt ttc tcc gat gga act aca gcg caa gct gat ttt att gct ggt gat 739
Ser Phe Ser Asp Gly Thr Thr Ala Gln Ala Asp Phe Ile Ala Gly Asp
200 205 210

cct tcc aca gat att gct gtg atc aag att agg gat gtg tcc aac ctt 787
Pro Ser Thr Asp Ile Ala Val Ile Lys Ile Arg Asp Val Ser Asn Leu
215 220 225

cca gtt atg agc ttt gga gat tcg gac gca tta ggc gtt gga caa agt	835
Pro Val Met Ser Phe Gly Asp Ser Asp Ala Leu Gly Val Gly Gln Ser	
230 235 240 245	
gtg atg gct gtt ggt tct cca ctg ggt ctg agc tcc act gtg acc acc	883
Val Met Ala Val Gly Ser Pro Leu Gly Leu Ser Ser Thr Val Thr Thr	
250 255 260	
ggg att gtg tcg gcc gtg aac cgt cct gtg cga gct tct ggt gat ggc	931
Gly Ile Val Ser Ala Val Asn Arg Pro Val Arg Ala Ser Gly Asp Gly	
265 270 275	
gga gag tcg tcc ctc atc gat gct atc cag acc gat gct gcg atc aac	979
Gly Glu Ser Ser Leu Ile Asp Ala Ile Gln Thr Asp Ala Ala Ile Asn	
280 285 290	
cct ggt aac tct ggt ggt ccg ctg gtt gat atg gat ggc aac ctc att	1027
Pro Gly Asn Ser Gly Gly Pro Leu Val Asp Met Asp Gly Asn Leu Ile	
295 300 305	
ggc atg aat tcg gta att gca tcg att tcg agc acc agc gat tcc gca	1075
Gly Met Asn Ser Val Ile Ala Ser Ile Ser Ser Thr Ser Asp Ser Ala	
310 315 320 325	
ggg tcc att ggt ctt ggt ttt tct atc cca tcc aac ttt gcc aag cgc	1123
Gly Ser Ile Gly Leu Gly Phe Ser Ile Pro Ser Asn Phe Ala Lys Arg	
330 335 340	
gtg gcc gat caa ttg atc agc acc ggc cag gta act cag ccg atg atc	1171
Val Ala Asp Gln Leu Ile Ser Thr Gly Gln Val Thr Gln Pro Met Ile	
345 350 355	
ggg gtg cag gtt ggc act gac aac tca gtg aca ggc gct gtg att gcc	1219
Gly Val Gln Val Gly Thr Asp Asn Ser Val Thr Gly Ala Val Ile Ala	
360 365 370	
agt gtt caa gat ggt gga ccg gcc gca gat gct gga ctt cag cca ggc	1267
Ser Val Gln Asp Gly Gly Pro Ala Ala Asp Ala Gly Leu Gln Pro Gly	
375 380 385	
gat atc gtg acc aag ctc aat gat cga gtt att gat agc cca gac tcc	1315
Asp Ile Val Thr Lys Leu Asn Asp Arg Val Ile Asp Ser Pro Asp Ser	
390 395 400 405	
ttg atc gct gct gtt cgt tcg cat gat ttt ggc gaa acc gtc act tta	1363
Leu Ile Ala Ala Val Arg Ser His Asp Phe Gly Glu Thr Val Thr Leu	
410 415 420	
aca att aca cag cca gat acc tcg cag agc cgg gag gta gag gtt act	1411
Thr Ile Thr Gln Pro Asp Thr Ser Gln Ser Arg Glu Val Glu Val Thr	
425 430 435	
ctg acg agt gag taggtttaaa agagttaatc tgc	1446
Leu Thr Ser Glu	
440	

<210> 56

<211> 441

<212> PRT

<213> Corynebacterium glutamicum

<400> 56

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 Ser Glu Thr Pro Ser Glu Thr Asn Ser Phe Glu His Val Arg Ser Ser
 20 25 30
 Tyr Pro Gln Trp Gly Asn Thr Ala Ser Asn Gln Asn Pro Tyr Pro Gly
 35 40 45
 Ala Gly Phe Gly Ser Glu Gln Asn Thr Gln Gln Gly Asn Glu Gln Gln
 50 55 60
 Ala Pro Ala Trp Thr Ser Trp Asp Asn Gln Pro Leu Ser Thr Asp Val
 65 70 75 80
 Lys Pro Ala Lys Glu Lys Arg Lys Val Gly Ile Gly Thr Ala Leu Ala
 85 90 95
 Leu Met Leu Val Gly Ser Ile Ala Thr Gly Ser Val Val Gly Val Ala
 100 105 110
 Ala Thr Gln Leu Gly Ser Asp Ser Ser Thr Pro Val Asn Ala Leu Glu
 115 120 125
 Gln Pro Ser Val Gln Arg Thr Thr Asn Ala Glu Pro Gly Ser Ala Glu
 130 135 140
 Gln Val Ala Ala Ala Val Leu Pro Ser Val Val Ser Ile Gln Ala Ile
 145 150 155 160
 Thr Arg Thr Ser Ala Ser Glu Gly Ser Gly Ser Ile Ile Ser Ser Asp
 165 170 175
 Gly Tyr Val Met Thr Asn Asn His Val Val Ala Gly Ile Glu Gln Ser
 180 185 190
 Gly Val Leu Glu Val Ser Phe Ser Asp Gly Thr Thr Ala Gln Ala Asp
 195 200 205
 Phe Ile Ala Gly Asp Pro Ser Thr Asp Ile Ala Val Ile Lys Ile Arg
 210 215 220
 Asp Val Ser Asn Leu Pro Val Met Ser Phe Gly Asp Ser Asp Ala Leu
 225 230 235 240
 Gly Val Gly Gln Ser Val Met Ala Val Gly Ser Pro Leu Gly Leu Ser
 245 250 255
 Ser Thr Val Thr Thr Gly Ile Val Ser Ala Val Asn Arg Pro Val Arg
 260 265 270
 Ala Ser Gly Asp Gly Gly Glu Ser Ser Leu Ile Asp Ala Ile Gln Thr
 275 280 285
 Asp Ala Ala Ile Asn Pro Gly Asn Ser Gly Gly Pro Leu Val Asp Met
 290 295 300
 Asp Gly Asn Leu Ile Gly Met Asn Ser Val Ile Ala Ser Ile Ser Ser

305	310	315	320
Thr Ser Asp Ser Ala Gly Ser Ile Gly Leu Gly Phe Ser Ile Pro Ser			
	325	330	335
Asn Phe Ala Lys Arg Val Ala Asp Gln Leu Ile Ser Thr Gly Gln Val			
	340	345	350
Thr Gln Pro Met Ile Gly Val Gln Val Gly Thr Asp Asn Ser Val Thr			
	355	360	365
Gly Ala Val Ile Ala Ser Val Gln Asp Gly Gly Pro Ala Ala Asp Ala			
	370	375	380
Gly Leu Gln Pro Gly Asp Ile Val Thr Lys Leu Asn Asp Arg Val Ile			
	385	390	400
Asp Ser Pro Asp Ser Leu Ile Ala Ala Val Arg Ser His Asp Phe Gly			
	405	410	415
Glu Thr Val Thr Leu Thr Ile Thr Gln Pro Asp Thr Ser Gln Ser Arg			
	420	425	430
Glu Val Glu Val Thr Leu Thr Ser Glu			
	435	440	

<210> 57

<211> 518

<212> DNA

<213> *Corynebacterium glutamicum*

<220>

<221> CDS

<222> (1)..(495)

<223> RXA02834

<400> 57

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Asp Ser Lys Gly Arg Ser Val Asp Phe Lys Asn Thr Ile Ile Ile Met	
1 5 10 15	

act agt aat att ggt tca caa gta tta ctt gaa aat gta aaa gat gct	96
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Thr Ser Asn Ile Gly Ser Gln Val Leu Leu Glu Asn Val Lys Asp Ala	
20 25 30	

ggt gaa att agt gat gat aca gag aaa gca gtt atg gac agt cta cat	144
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Gly Glu Ile Ser Asp Asp Thr Glu Lys Ala Val Met Asp Ser Leu His	
35 40 45	

gca tac ttc aaa cct gaa ata tta aat cgt atg gat gac atc gtg tta	192
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Ala Tyr Phe Lys Pro Glu Ile Leu Asn Arg Met Asp Asp Ile Val Leu	
50 55 60	

ttt aaa cca tta tca gtt aat gat atg agt atg att gta gat aaa att	240
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Phe Lys Pro Leu Ser Val Asn Asp Met Ser Met Ile Val Asp Lys Ile	
65 70 75 80	

tta aca caa tta aat atg aga tta tta gat caa cat atc tca att gaa	288
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Leu Thr Gln Leu Asn Met Arg Leu Leu Asp Gln His Ile Ser Ile Glu	
85 90 95	

gtg aca gaa gaa gcg aaa aaa tgg cta ggt gaa gaa gcg tat gaa cca 336
 Val Thr Glu Glu Ala Lys Lys Trp Leu Gly Glu Glu Ala Tyr Glu Pro
 100 105 110

caa ttt ggt gca aga cca tta aaa cgc ttt gtt caa cga caa ata gaa 384
 Gln Phe Gly Ala Arg Pro Leu Lys Arg Phe Val Gln Arg Gln Ile Glu
 115 120 125

act cca att gca cgt atg atg att aaa gaa agt cta cct gaa ggt aca 432
 Thr Pro Ile Ala Arg Met Met Ile Lys Glu Ser Leu Pro Glu Gly Thr
 130 135 140

ata att aaa gta gat tta aat gac aat aaa gaa ctt gat ttt aag gtt 480
 Ile Ile Lys Val Asp Leu Asn Asp Asn Lys Glu Leu Asp Phe Lys Val
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 Val Lys Pro Thr Ser
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<212> PRT

<213> *Corynebacterium glutamicum*

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Gly Glu Ile Ser Asp Asp Thr Glu Lys Ala Val Met Asp Ser Leu His
 35 40 45

Ala Tyr Phe Lys Pro Glu Ile Leu Asn Arg Met Asp Asp Ile Val Leu
 50 55 60

Phe Lys Pro Leu Ser Val Asn Asp Met Ser Met Ile Val Asp Lys Ile
 65 70 75 80

Leu Thr Gln Leu Asn Met Arg Leu Leu Asp Gln His Ile Ser Ile Glu
 85 90 95

Val Thr Glu Glu Ala Lys Lys Trp Leu Gly Glu Glu Ala Tyr Glu Pro
 100 105 110

Gln Phe Gly Ala Arg Pro Leu Lys Arg Phe Val Gln Arg Gln Ile Glu
 115 120 125

Thr Pro Ile Ala Arg Met Met Ile Lys Glu Ser Leu Pro Glu Gly Thr
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Ile Ile Lys Val Asp Leu Asn Asp Asn Lys Glu Leu Asp Phe Lys Val
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Val Lys Pro Thr Ser
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<223> RXA00112
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gtg acc aac gcc cac gtt gtt gca ggt acc tcc acc gtc agc ctg gat Val Thr Asn Ala His Val Val Ala Gly Thr Ser Thr Val Ser Leu Asp 230 235 240 245	835
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gag gta cag gag cgg atc ggc gac atc acc gcg ctg act cag cct gtc Glu Val Gln Glu Arg Ile Gly Asp Ile Thr Ala Leu Thr Gln Pro Val 375 380 385	1267
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<213> Corynebacterium glutamicum

<400> 60

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 Pro Phe Val Met Gly Leu Thr Asp Ser Thr Ala Leu Arg Phe Leu Leu
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 Ala Ile Gly Thr Val Val Leu Leu Val Gly Leu Gly Asn Leu Ile Gly
 65 70 75 80
 Ala His Leu Gly Ala Ala Ile Arg Asp Asn Ile Lys Phe Arg Ser Ser
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 Arg Thr Leu Asp Ser Gly Leu Gly Ala Ile Phe Gln Val Leu Ala Thr
 100 105 110
 Leu Ile Val Val Trp Leu Val Ala Ile Pro Leu Ala Thr Gly Leu Pro
 115 120 125
 Gly Thr Val Ala Ser Gly Ile Arg Asp Ser Arg Ile Leu Gly Phe Val
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 Asp Lys Tyr Thr Pro Gln Gly Leu Asp Thr Leu Pro Ser Lys Ile Ala
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 Ala Met Leu Ser Glu Ser Gly Leu Pro Pro Leu Ile Ser Pro Phe Thr
 165 170 175
 Gly Gly Ser Ser Val Glu Val Asp Ala Pro Glu Ile Asn Val Thr Asn
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 Val Asp Leu Val Glu Ala Met Arg Pro Ser Val Ile His Val Met Gly
 195 200 205
 Asp Ala Gln Glu Cys Ser Arg Arg Leu Met Gly Ser Gly Phe Val Ala
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 Ser Pro Asp Tyr Val Val Thr Asn Ala His Val Val Ala Gly Thr Ser
 225 230 235 240
 Thr Val Ser Leu Asp Thr Met Ile Gly Thr Arg Ser Ala Glu Val Val
 245 250 255
 Phe Tyr Asp Pro Asn Leu Asp Ile Ala Val Leu Tyr Ser Pro Asp Leu
 260 265 270
 Gly Leu Asp Pro Leu Pro Trp Ala Ser Thr Pro Leu Asp Thr Gly Asp
 275 280 285
 Glu Ala Ile Val Met Gly Phe Pro Gln Ser Gly Pro Phe Asn Ala Ser
 290 295 300
 Pro Ala Arg Val Arg Glu Arg Ile Met Ile Thr Gly Ser Asn Ile Tyr

305	310	315	320
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325		330	335
Gln Ser Gly Asn Ser Gly Gly Pro Met Thr Asn Glu Met Gly Glu Val			
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Val Gly Val Val Phe Gly Ala Ala Ile Asp Gly Ser Asp Thr Gly Tyr			
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                                         Met Pro Thr Ser Arg
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tac gtg ctg cct tcc ttc att gag cag tcc gca tac ggc acc aaa gag 163
Tyr Val Leu Pro Ser Phe Ile Glu Gln Ser Ala Tyr Gly Thr Lys Glu
                               10                               15                               20

acc aac cct tac gca aaa ctc ttt gaa gag cgc atc atc ttc ctg ggc 211
Thr Asn Pro Tyr Ala Lys Leu Phe Glu Glu Arg Ile Ile Phe Leu Gly
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acc cag gtc gac gac acc tct gca aac gac atc atg gcg cag ctc ctt 259
Thr Gln Val Asp Asp Thr Ser Ala Asn Asp Ile Met Ala Gln Leu Leu
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gtc ctc gaa ggc atg gac cca gac cgc gat atc acc ctg tac atc aac 307
Val Leu Glu Gly Met Asp Pro Asp Arg Asp Ile Thr Leu Tyr Ile Asn
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Ser Pro Gly Gly Ser Phe Thr Ala Leu Met Ala Ile Tyr Asp Thr Met
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Gln Tyr Val Arg Pro Asp Val Gln Thr Val Cys Leu Gly Gln Ala Ala
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Ser Ala Ala Ala Val Leu Leu Ala Ala Gly Ala Pro Gly Lys Arg Ala

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Thr	Gln	Gly	Gln	Val	Ser	Asp	Leu	Glu	Ile	Gln	Ala	Ala	Glu	Ile	Glu															
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Arg	Lys	Leu	Lys	Arg																										
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<212> PRT

<213> Corynebacterium glutamicum

<400> 62

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Ile	Ile	Phe	Leu	Gly	Thr	Gln	Val	Asp	Asp	Thr	Ser	Ala	Asn	Asp	Ile
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Met	Ala	Gln	Leu	Leu	Val	Leu	Glu	Gly	Met	Asp	Pro	Asp	Arg	Asp	Ile
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Thr	Leu	Tyr	Ile	Asn	Ser	Pro	Gly	Gly	Ser	Phe	Thr	Ala	Leu	Met	Ala
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Ile	Tyr	Asp	Thr	Met	Gln	Tyr	Val	Arg	Pro	Asp	Val	Gln	Thr	Val	Cys
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Leu	Gly	Gln	Ala	Ala	Ser	Ala	Ala	Ala	Val	Leu	Leu	Ala	Ala	Gly	Ala
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Pro	Gly	Lys	Arg	Ala	Val	Leu	Pro	Asn	Ser	Arg	Val	Leu	Ile	His	Gln
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Pro	Ala	Thr	Gln	Gly	Thr	Gln	Gly	Gln	Val	Ser	Asp	Leu	Glu	Ile	Gln
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cag ctg atc gct gag cac acc ggc cag acc ttt gag cag atc tcc aag 595
 Gln Leu Ile Ala Glu His Thr Gly Gln Thr Phe Glu Gln Ile Ser Lys
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gat tcc gat cgt gac cgc tgg ttc act gca cag gaa gct aag gat tac 643
 Asp Ser Asp Arg Asp Arg Trp Phe Thr Ala Gln Glu Ala Lys Asp Tyr
 170 175 180

gga ctt gtt gac cac gtg att acc ttg gct gaa ggc cca atc agc aac 691
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Leu Ser Ala Glu Asp Pro Thr Arg Asp Ile Ser Leu Tyr Ile Asn Ser
 50 55 60

Pro Gly Gly Ser Val Thr Ala Gly Met Ala Ile Tyr Asp Thr Met Lys
 65 70 75 80

Tyr Ser Pro Cys Asp Ile Ala Thr Tyr Gly Met Gly Leu Ala Ala Ser
 85 90 95

Met Gly Gln Phe Leu Leu Ser Gly Gly Thr Lys Gly Lys Arg Phe Ala
 100 105 110

Leu Pro His Ala Arg Ile Met Met His Gln Pro Ser Ala Gly Val Gly
 115 120 125

Gly Thr Ala Ala Asp Ile Ala Ile Gln Ala Glu Gln Phe Ala Ala Thr
 130 135 140

Lys Arg Glu Met Ala Gln Leu Ile Ala Glu His Thr Gly Gln Thr Phe
 145 150 155 160

Glu Gln Ile Ser Lys Asp Ser Asp Arg Asp Arg Trp Phe Thr Ala Gln
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Gly Pro Ile Ser Asn
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<212> DNA
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<222> (101)..(1852)
<223> RXN03094

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Met Ser Ser Phe Asn
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cca act acc aaa acc aat gaa gcc atg cag gct gct ctt cag cag gca 163
Pro Thr Thr Lys Thr Asn Glu Ala Met Gln Ala Ala Leu Gln Gln Ala
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tcc tcg gct ggc aac cct gat att cgt cca gct cac ctg ttg gct gcc 211
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Ile Leu Glu Gln Thr Asp Gly Val Ala Ala Pro Val Leu Met Ala Thr
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Leu Gly Asp Glu Tyr Val Ser Thr Glu Val Leu Leu Ala Gly Ile Ala
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Tyr Asp Ala Ile Lys Glu Ala Phe Pro Ser Val Arg Gly Ser Gln Arg
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 Arg Arg Leu Glu Ile Glu Glu Met Ala Leu Ser Lys Glu Ser Asp Ala
 425 430 435
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 Ala Ser Lys Glu Arg Leu Glu Lys Leu Arg Ser Glu Leu Ala Asp Glu
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<211> 584

<212> PRT

<213> Corynebacterium glutamicum

<400> 66

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His Leu Leu Ala Ala Ile Leu Glu Gln Thr Asp Gly Val Ala Ala Pro

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Lys	Lys	Leu	Val	Ala	Ser	Tyr	Pro	Lys	Ala	Ser	Gly	Ala	Asn	Met	Ala
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Asn	Pro	Asn	Phe	Asn	Arg	Asp	Ala	Leu	Asn	Ala	Phe	Thr	Ala	Ala	Gln
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Glu	Leu	Ala	Gly	Glu	Leu	Gly	Asp	Glu	Tyr	Val	Ser	Thr	Glu	Val	Leu
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Leu	Ala	Gly	Ile	Ala	Arg	Gly	Lys	Ser	Asp	Ala	Ala	Asp	Leu	Leu	Thr
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Arg	Gly	Ser	Gln	Arg	Val	Thr	Thr	Gln	Asp	Pro	Glu	Gly	Gln	Phe	Gln
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Lys	Ile	Asp	Pro	Val	Ile	Gly	Arg	Asp	Gln	Glu	Ile	Arg	Arg	Val	Val
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Gln	Val	Leu	Ser	Arg	Arg	Thr	Lys	Asn	Asn	Pro	Val	Leu	Ile	Gly	Glu
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Leu	Asp	Leu	Gly	Ser	Met	Val	Ala	Gly	Ala	Lys	Tyr	Arg	Gly	Glu	Phe
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Glu	Val	Val	Thr	Phe	Ile	Asp	Glu	Leu	His	Thr	Ile	Val	Gly	Ala	Gly
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Tyr	Arg	Lys	Tyr	Ile	Glu	Lys	Asp	Ala	Ala	Leu	Glu	Arg	Arg	Phe	Gln
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Gln	Val	Tyr	Val	Gly	Glu	Pro	Thr	Val	Glu	Asp	Ala	Ile	Gly	Ile	Leu
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Arg	Gly	Leu	Lys	Glu	Arg	Tyr	Glu	Val	His	His	Gly	Val	Arg	Ile	Gln
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Asp Ser Ala Leu Val Ala Ala Ala Glu Leu Ser Asn Arg Tyr Ile Thr
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Ser Arg Phe Leu Pro Asp Lys Ala Ile Asp Leu Val Asp Glu Ala Ala
385 390 395 400

Ser Arg Leu Arg Met Glu Ile Asp Ser Ser Pro Gln Glu Ile Asp Glu
405 410 415

Leu Glu Arg Ile Val Arg Arg Leu Glu Ile Glu Glu Met Ala Leu Ser
420 425 430

Lys Glu Ser Asp Ala Ala Ser Lys Glu Arg Leu Glu Lys Leu Arg Ser
435 440 445

Glu Leu Ala Asp Glu Arg Glu Lys Leu Ser Glu Leu Lys Ala Arg Trp
450 455 460

Gln Asn Glu Lys Thr Ala Ile Asp Asp Val Arg Glu Met Lys Glu Glu
465 470 475 480

Leu Glu Ala Leu Arg Ser Glu Ser Asp Ile Ala Lys Arg Asp Gly Asn
485 490 495

Tyr Cys Arg Val Ala Lys Leu Arg Tyr Gly Arg Ile Pro Glu Leu Glu
500 505 510

Lys Gln Ile Glu Asp Ala Glu Ser Lys Val Glu Val Asn Glu Asn Ala
515 520 525

Met Leu Thr Glu Glu Val Thr Pro Asp Thr Ile Ala Asp Val Val Ser
530 535 540

Ala Trp Thr Gly Ile Pro Ala Gly Lys Met Met Gln Gly Glu Thr Glu
545 550 555 560

Lys Leu Leu Asn Met Glu Arg Val Leu Gly Asn Arg Val Val Gly Gln
565 570 575

Leu Glu Ser Gly Asn Cys Ser Val
580

<210> 67

<211> 1816

<212> DNA

<213> Corynebacterium glutamicum

<220>

<221> CDS

<222> (101)..(1816)

<223> FRXA01668

<400> 67

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ttgtgtaggt tggcgcccaa caaagaagg gcgttgaaag atg agt tca ttc aat 115
Met Ser Ser Phe Asn

1

5

cca act acc aaa acc aat gaa gcc atg cag gct gct ctt cag cag gca	163
Pro Thr Thr Lys Thr Asn Glu Ala Met Gln Ala Ala Leu Gln Gln Ala	
10 15 20	
tcc tcg gct ggc aac cct gat att cgt cca gct cac ctg ttg gct gcc	211
Ser Ser Ala Gly Asn Pro Asp Ile Arg Pro Ala His Leu Leu Ala Ala	
25 30 35	
atc ttg gag caa act gat ggc gta gca gcg cca gtc ctc atg gct act	259
Ile Leu Glu Gln Thr Asp Gly Val Ala Ala Pro Val Leu Met Ala Thr	
40 45 50	
ggg gtg gat cct aag gag atc ctc gca gag gcc aag aag ttg gtt gct	307
Gly Val Asp Pro Lys Glu Ile Leu Ala Glu Ala Lys Lys Leu Val Ala	
55 60 65	
tct tac ccc aag gct tct ggc gcc aat atg gct aat cca aac ttc aac	355
Ser Tyr Pro Lys Ala Ser Gly Ala Asn Met Ala Asn Pro Asn Phe Asn	
70 75 80 85	
cgg gat gcc ctc aat gcg ttc act gca gct cag gag ctt gcc ggt gag	403
Arg Asp Ala Leu Asn Ala Phe Thr Ala Ala Gln Glu Leu Ala Gly Glu	
90 95 100	
ttg ggc gat gag tac gtc tca acc gaa gta ctt ctt gcc ggt atc gct	451
Leu Gly Asp Glu Tyr Val Ser Thr Glu Val Leu Leu Ala Gly Ile Ala	
105 110 115	
cgc gga aag tct gat gct gcg gat ctg ttg acc aac aag ggt gca acc	499
Arg Gly Lys Ser Asp Ala Ala Asp Leu Leu Thr Asn Lys Gly Ala Thr	
120 125 130	
tat gac gcc atc aaa gag gct ttc cct tcg gtt cgt gga tct cag cgt	547
Tyr Asp Ala Ile Lys Glu Ala Phe Pro Ser Val Arg Gly Ser Gln Arg	
135 140 145	
gtc acc act cag gat cca gag gga cag ttc cag gct ttg gaa aag tac	595
Val Thr Thr Gln Asp Pro Glu Gly Gln Phe Gln Ala Leu Glu Lys Tyr	
150 155 160 165	
tcc act gac ctg acc aag ctt gct cgt gaa ggc aag att gat cct gtt	643
Ser Thr Asp Leu Thr Lys Leu Ala Arg Glu Gly Lys Ile Asp Pro Val	
170 175 180	
att ggc cgt gac cag gaa att cgt cgc gtc gtt cag gtg ctt agc cgt	691
Ile Gly Arg Asp Gln Glu Ile Arg Arg Val Val Gln Val Leu Ser Arg	
185 190 195	
cgt acc aag aac aac cct gtt ctg atc ggt gag cca ggt gtc ggt aaa	739
Arg Thr Lys Asn Asn Pro Val Leu Ile Gly Glu Pro Gly Val Gly Lys	
200 205 210	
acc gcc atc gtg gaa ggc ctt gca cgc cgc atc gtt gct ggt gac gtt	787
Thr Ala Ile Val Glu Gly Leu Ala Arg Arg Ile Val Ala Gly Asp Val	
215 220 225	
cca gaa tcc ctc aag ggc aaa act ctg atc agt ctt gat ctt ggt tcc	835
Pro Glu Ser Leu Lys Gly Lys Thr Leu Ile Ser Leu Asp Leu Gly Ser	
230 235 240 245	
atg gtt gcc ggc gct aag tat cgc ggt gaa ttc gag gag cga ctg aag	883

Met	Val	Ala	Gly	Ala	Lys	Tyr	Arg	Gly	Glu	Phe	Glu	Glu	Arg	Leu	Lys		
				250					255					260			
gct	gtt	ctg	gat	gag	atc	aag	gga	gct	aac	ggc	gaa	gtc	gtt	acc	ttc	931	
Ala	Val	Leu	Asp	Glu	Ile	Lys	Gly	Ala	Asn	Gly	Glu	Val	Val	Thr	Phe		
			265				270						275				
atc	gat	gag	ctg	cac	acc	atc	gtc	ggc	gct	ggt	gct	tcg	ggg	gaa	tcc	979	
Ile	Asp	Glu	Leu	His	Thr	Ile	Val	Gly	Ala	Gly	Ala	Ser	Gly	Glu	Ser		
		280					285					290					
gcc	atg	gat	gcc	gga	aac	atg	att	aag	cca	ctg	ctt	gcc	cgc	ggg	gag	1027	
Ala	Met	Asp	Ala	Gly	Asn	Met	Ile	Lys	Pro	Leu	Leu	Ala	Arg	Gly	Glu		
	295					300					305						
ctg	cgc	ttg	gtt	ggg	gcc	acc	acg	ctg	aat	gag	tac	cgc	aag	tac	atc	1075	
Leu	Arg	Leu	Val	Gly	Ala	Thr	Thr	Leu	Asn	Glu	Tyr	Arg	Lys	Tyr	Ile		
310				315						320					325		
gaa	aag	gac	gct	gcc	ctg	gag	cgt	agg	ttc	cag	cag	gtt	tat	gtc	ggg	1123	
Glu	Lys	Asp	Ala	Ala	Leu	Glu	Arg	Arg	Phe	Gln	Gln	Val	Tyr	Val	Gly		
			330					335						340			
gag	cca	acg	gta	gaa	gat	gcc	atc	ggg	att	ctt	cgt	gga	ttg	aag	gaa	1171	
Glu	Pro	Thr	Val	Glu	Asp	Ala	Ile	Gly	Ile	Leu	Arg	Gly	Leu	Lys	Glu		
			345					350					355				
cgc	tac	gag	gtc	cat	cac	ggg	gtc	cgc	atc	cag	gac	tcc	gca	ctg	gtc	1219	
Arg	Tyr		Val	His	His	Gly	Val	Arg	Ile	Gln	Asp	Ser	Ala	Leu	Val		
	360					365					370						
gcc	gca	gct	gaa	ctc	tca	aac	cgc	tat	atc	acc	agc	cgt	ttc	ctt	cct	1267	
Ala	Ala	Ala	Glu	Leu	Ser	Asn	Arg	Tyr	Ile	Thr	Ser	Arg	Phe	Leu	Pro		
	375					380					385						
gat	aag	gct	att	gac	tta	gtt	gat	gag	gca	gca	tca	cgc	ctg	cgc	atg	1315	
Asp	Lys	Ala	Ile	Asp	Leu	Val	Asp	Glu	Ala	Ala	Ser	Arg	Leu	Arg	Met		
390				395				400						405			
gag	att	gat	tct	tca	cct	cag	gaa	atc	gat	gag	ctg	gag	cgt	atc	gtc	1363	
Glu	Ile	Asp	Ser	Ser	Pro	Gln	Glu	Ile	Asp	Glu	Leu	Glu	Arg	Ile	Val		
			410					415						420			
cgc	cgc	ctc	gag	atc	gaa	gag	atg	gcg	ctg	tcc	aag	gaa	tcc	gat	gca	1411	
Arg	Arg	Leu	Glu	Ile	Glu	Glu	Met	Ala	Leu	Ser	Lys	Glu	Ser	Asp	Ala		
		425					430						435				
gct	tcc	aag	gaa	cgt	cta	gaa	aag	ctg	cgc	tcg	gaa	ctt	gct	gat	gaa	1459	
Ala	Ser	Lys	Glu	Arg	Leu	Glu	Lys	Leu	Arg	Ser	Glu	Leu	Ala	Asp	Glu		
		440				445					450						
cgc	gaa	aag	ctc	tct	gag	ttg	aag	gct	cgt	tgg	cag	aat	gag	aaa	act	1507	
Arg	Glu	Lys	Leu	Ser	Glu	Leu	Lys	Ala	Arg	Trp	Gln	Asn	Glu	Lys	Thr		
	455				460					465							
gct	att	gac	gat	gtc	cgg	gag	atg	aaa	gaa	gag	ctg	gaa	gcg	ctg	cgt	1555	
Ala	Ile	Asp	Asp	Val	Arg	Glu	Met	Lys	Glu	Glu	Leu	Glu	Ala	Leu	Arg		
470				475					480					485			
tct	gag	tcg	gat	att	gca	aaa	cgt	gac	ggc	aat	tat	tgt	cgt	gtc	gca	1603	
Ser	Glu	Ser	Asp	Ile	Ala	Lys	Arg	Asp	Gly	Asn	Tyr	Cys	Arg	Val	Ala		

490	495	500	
aag ctt cgc tac ggc cga atc cct	gag ctg gaa aag cag atc gag gat	1651	
Lys Leu Arg Tyr Gly Arg Ile Pro	Glu Leu Glu Lys Gln Ile Glu Asp		
505	510	515	
gca gaa tcc aag gtc gag gtc aat	gaa aat gcc atg ctc act gag gag	1699	
Ala Glu Ser Lys Val Glu Val Asn	Glu Asn Ala Met Leu Thr Glu Glu		
520	525	530	
gtc acg cca gac acg atc gcc gat	gtg gtt tcc gca tgg acg ggc att	1747	
Val Thr Pro Asp Thr Ile Ala Asp	Val Val Ser Ala Trp Thr Gly Ile		
535	540	545	
cct gca ggc aag atg atg cag ggt	gag acc gag aag ctg ctc aac atg	1795	
Pro Ala Gly Lys Met Met Gln Gly	Glu Thr Glu Lys Leu Leu Asn Met		
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gag cgc gtc ttg ggc aac ccg		1816	
Glu Arg Val Leu Gly Asn Pro			
570			

<210> 68

<211> 572

<212> PRT

<213> Corynebacterium glutamicum

<400> 68

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His Leu Leu Ala Ala Ile Leu Glu Gln Thr Asp Gly Val Ala Ala Pro	
35 40 45	
Val Leu Met Ala Thr Gly Val Asp Pro Lys Glu Ile Leu Ala Glu Ala	
50 55 60	
Lys Lys Leu Val Ala Ser Tyr Pro Lys Ala Ser Gly Ala Asn Met Ala	
65 70 75 80	
Asn Pro Asn Phe Asn Arg Asp Ala Leu Asn Ala Phe Thr Ala Ala Gln	
85 90 95	
Glu Leu Ala Gly Glu Leu Gly Asp Glu Tyr Val Ser Thr Glu Val Leu	
100 105 110	
Leu Ala Gly Ile Ala Arg Gly Lys Ser Asp Ala Ala Asp Leu Leu Thr	
115 120 125	
Asn Lys Gly Ala Thr Tyr Asp Ala Ile Lys Glu Ala Phe Pro Ser Val	
130 135 140	
Arg Gly Ser Gln Arg Val Thr Thr Gln Asp Pro Glu Gly Gln Phe Gln	
145 150 155 160	
Ala Leu Glu Lys Tyr Ser Thr Asp Leu Thr Lys Leu Ala Arg Glu Gly	
165 170 175	

Lys Ile Asp Pro Val Ile Gly Arg Asp Gln Glu Ile Arg Arg Val Val
 180 185 190
 Gln Val Leu Ser Arg Arg Thr Lys Asn Asn Pro Val Leu Ile Gly Glu
 195 200 205
 Pro Gly Val Gly Lys Thr Ala Ile Val Glu Gly Leu Ala Arg Arg Ile
 210 215 220
 Val Ala Gly Asp Val Pro Glu Ser Leu Lys Gly Lys Thr Leu Ile Ser
 225 230 235 240
 Leu Asp Leu Gly Ser Met Val Ala Gly Ala Lys Tyr Arg Gly Glu Phe
 245 250 255
 Glu Glu Arg Leu Lys Ala Val Leu Asp Glu Ile Lys Gly Ala Asn Gly
 260 265 270
 Glu Val Val Thr Phe Ile Asp Glu Leu His Thr Ile Val Gly Ala Gly
 275 280 285
 Ala Ser Gly Glu Ser Ala Met Asp Ala Gly Asn Met Ile Lys Pro Leu
 290 295 300
 Leu Ala Arg Gly Glu Leu Arg Leu Val Gly Ala Thr Thr Leu Asn Glu
 305 310 315 320
 Tyr Arg Lys Tyr Ile Glu Lys Asp Ala Ala Leu Glu Arg Arg Phe Gln
 325 330 335
 Gln Val Tyr Val Gly Glu Pro Thr Val Glu Asp Ala Ile Gly Ile Leu
 340 345 350
 Arg Gly Leu Lys Glu Arg Tyr Glu Val His His Gly Val Arg Ile Gln
 355 360 365
 Asp Ser Ala Leu Val Ala Ala Ala Glu Leu Ser Asn Arg Tyr Ile Thr
 370 375 380
 Ser Arg Phe Leu Pro Asp Lys Ala Ile Asp Leu Val Asp Glu Ala Ala
 385 390 395 400
 Ser Arg Leu Arg Met Glu Ile Asp Ser Ser Pro Gln Glu Ile Asp Glu
 405 410 415
 Leu Glu Arg Ile Val Arg Arg Leu Glu Ile Glu Glu Met Ala Leu Ser
 420 425 430
 Lys Glu Ser Asp Ala Ala Ser Lys Glu Arg Leu Glu Lys Leu Arg Ser
 435 440 445
 Glu Leu Ala Asp Glu Arg Glu Lys Leu Ser Glu Leu Lys Ala Arg Trp
 450 455 460
 Gln Asn Glu Lys Thr Ala Ile Asp Asp Val Arg Glu Met Lys Glu Glu
 465 470 475 480
 Leu Glu Ala Leu Arg Ser Glu Ser Asp Ile Ala Lys Arg Asp Gly Asn
 485 490 495

Tyr Cys Arg Val Ala Lys Leu Arg Tyr Gly Arg Ile Pro Glu Leu Glu
 500 505 510
 Lys Gln Ile Glu Asp Ala Glu Ser Lys Val Glu Val Asn Glu Asn Ala
 515 520 525
 Met Leu Thr Glu Glu Val Thr Pro Asp Thr Ile Ala Asp Val Val Ser
 530 535 540
 Ala Trp Thr Gly Ile Pro Ala Gly Lys Met Met Gln Gly Glu Thr Glu
 545 550 555 560
 Lys Leu Leu Asn Met Glu Arg Val Leu Gly Asn Pro
 565 570

<210> 69
 <211> 1401
 <212> DNA
 <213> *Corynebacterium glutamicum*

<220>
 <221> CDS
 <222> (101)..(1378)
 <223> RXN01120

<400> 69
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 agtaaattctt ttcaatcggtg gaagcgggtc tcacagtcta atg gca cgt atg cag 115
 Met Ala Arg Met Gln
 1 5
 gaa agc gcc gat ctg ctc aaa tgt tcc ttc tgc gga aag agc caa aag 163
 Glu Ser Ala Asp Leu Leu Lys Cys Ser Phe Cys Gly Lys Ser Gln Lys
 10 15 20
 cag gta aaa aaa ctc atc gcg ggt ggc gcc gta tat atc tgt gat gag 211
 Gln Val Lys Lys Leu Ile Ala Gly Gly Ala Val Tyr Ile Cys Asp Glu
 25 30 35
 tgc att gag ctg tgc aac gag att att gaa gaa gaa ctc ggt caa gct 259
 Cys Ile Glu Leu Cys Asn Glu Ile Ile Glu Glu Glu Leu Gly Gln Ala
 40 45 50
 caa cac gac gag cag gag cgc aac gag ctc ccc aag ccg tcg gag att 307
 Gln His Asp Glu Gln Glu Arg Asn Glu Leu Pro Lys Pro Ser Glu Ile
 55 60 65
 tca gcc ttc ctt gat act tat gtc atc ggg cag gac cca gca aaa cgt 355
 Ser Ala Phe Leu Asp Thr Tyr Val Ile Gly Gln Asp Pro Ala Lys Arg
 70 75 80 85
 atc ctg tcg gtt gcg gtg tac aac cat tac aag cgt ctc cgc gca tcg 403
 Ile Leu Ser Val Ala Val Tyr Asn His Tyr Lys Arg Leu Arg Ala Ser
 90 95 100
 gaa acc atc ggt cgt cgc agg aat gac gag cct gaa acc gaa ctg gtt 451
 Glu Thr Ile Gly Arg Arg Arg Asn Asp Glu Pro Glu Thr Glu Leu Val
 105 110 115

aag tcc aat att ttg atg ctc ggc ccc act ggc tcc ggc aag act ttc	499
Lys Ser Asn Ile Leu Met Leu Gly Pro Thr Gly Ser Gly Lys Thr Phe	
120 125 130	
ctt gcc cag act ttg gca aag ctg ctg gat gtt cct ttt gct atc gcg	547
Leu Ala Gln Thr Leu Ala Lys Leu Leu Asp Val Pro Phe Ala Ile Ala	
135 140 145	
gat gcc acc tca ctg acc gag gct ggt tat gtg ggc gag gat gtg gaa	595
Asp Ala Thr Ser Leu Thr Glu Ala Gly Tyr Val Gly Glu Asp Val Glu	
150 155 160 165	
aac atc ttg ctc aag ctg ctt cag gct gct gat ttt gat gtg gaa cgt	643
Asn Ile Leu Leu Lys Leu Leu Gln Ala Ala Asp Phe Asp Val Glu Arg	
170 175 180	
gca cag cgc ggc atc att tac atc gat gaa gtg gac aag att tcc cgc	691
Ala Gln Arg Gly Ile Ile Tyr Ile Asp Glu Val Asp Lys Ile Ser Arg	
185 190 195	
aag tct gaa aac cca tcc atc act cgc gat gtt tcc ggt gaa ggc gtg	739
Lys Ser Glu Asn Pro Ser Ile Thr Arg Asp Val Ser Gly Glu Gly Val	
200 205 210	
cag cag gca ctg ctg aaa att ttg gaa ggc act gtc gcc gca atc cca	787
Gln Gln Ala Leu Leu Lys Ile Leu Glu Gly Thr Val Ala Ala Ile Pro	
215 220 225	
ccg cag gga gga cgc aag cac ccc aac cag gat ttc atc cag ctg gat	835
Pro Gln Gly Gly Arg Lys His Pro Asn Gln Asp Phe Ile Gln Leu Asp	
230 235 240 245	
acc acc aac att ttg ttc atc gtt gct ggt gcg ttc tct ggt ctg gag	883
Thr Thr Asn Ile Leu Phe Ile Val Ala Gly Ala Phe Ser Gly Leu Glu	
250 255 260	
aag gtc atc gcg gac cgc aat ggc aag aaa ggc ttg ggc ttc ggt gtg	931
Lys Val Ile Ala Asp Arg Asn Gly Lys Lys Gly Leu Gly Phe Gly Val	
265 270 275	
gag gtc tct tcc aag aag gaa gaa gcc aac att gtg gat atc ttc aag	979
Glu Val Ser Ser Lys Lys Glu Glu Ala Asn Ile Val Asp Ile Phe Lys	
280 285 290	
gat gtc ctc cct gag gac ctg gtg aag ttt ggt ctc atc cca gaa ttc	1027
Asp Val Leu Pro Glu Asp Leu Val Lys Phe Gly Leu Ile Pro Glu Phe	
295 300 305	
att ggg cgt ctg cca gtc gtt gcc acc gta tcc aac ctg gat cag aaa	1075
Ile Gly Arg Leu Pro Val Val Ala Thr Val Ser Asn Leu Asp Gln Lys	
310 315 320 325	
tct ctg gtc aag gtt ctc acg gag cct cgt aac tca ttg gtg aag cag	1123
Ser Leu Val Lys Val Leu Thr Glu Pro Arg Asn Ser Leu Val Lys Gln	
330 335 340	
tat cga cgt ctg ttt gaa atg gat gac gct gtg ttg acc ttt act gat	1171
Tyr Arg Arg Lys Phe Glu Met Asp Asp Ala Val Leu Thr Phe Thr Asp	
345 350 355	
gat gct ttg gag gag atc gct aat cag gca ctc gag cgc aaa act ggc	1219

Asp Ala Leu Glu Glu Ile Ala Asn Gln Ala Leu Glu Arg Lys Thr Gly
 360 365 370
 gcc cgt ggc ctg cgc gcg atc atg gaa gag atc ctg gtt ccg atc atg 1267
 Ala Arg Gly Leu Arg Ala Ile Met Glu Glu Ile Leu Val Pro Ile Met
 375 380 385
 tat gac ctc cca gac cgt aaa gac gtt ggc gaa gtc atc atc aac ggt 1315
 Tyr Asp Leu Pro Asp Arg Lys Asp Val Gly Glu Val Ile Ile Asn Gly
 390 395 400 405
 gcc gtt gcc cgt ggc gaa gcc gaa cca gag atg ttg gaa gct gtc gca 1363
 Ala Val Ala Arg Gly Glu Ala Glu Pro Glu Met Leu Glu Ala Val Ala
 410 415 420
 gaa gaa aag acc gcg tagttggcag gagttatcac cgg 1401
 Glu Glu Lys Thr Ala
 425

<210> 70
 <211> 426
 <212> PRT
 <213> *Corynebacterium glutamicum*

<400> 70
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 Gly Lys Ser Gln Lys Gln Val Lys Lys Leu Ile Ala Gly Gly Ala Val
 20 25 30
 Tyr Ile Cys Asp Glu Cys Ile Glu Leu Cys Asn Glu Ile Ile Glu Glu
 35 40 45
 Glu Leu Gly Gln Ala Gln His Asp Glu Gln Glu Arg Asn Glu Leu Pro
 50 55 60
 Lys Pro Ser Glu Ile Ser Ala Phe Leu Asp Thr Tyr Val Ile Gly Gln
 65 70 75 80
 Asp Pro Ala Lys Arg Ile Leu Ser Val Ala Val Tyr Asn His Tyr Lys
 85 90 95
 Arg Leu Arg Ala Ser Glu Thr Ile Gly Arg Arg Arg Asn Asp Glu Pro
 100 105 110
 Glu Thr Glu Leu Val Lys Ser Asn Ile Leu Met Leu Gly Pro Thr Gly
 115 120 125
 Ser Gly Lys Thr Phe Leu Ala Gln Thr Leu Ala Lys Leu Leu Asp Val
 130 135 140
 Pro Phe Ala Ile Ala Asp Ala Thr Ser Leu Thr Glu Ala Gly Tyr Val
 145 150 155 160
 Gly Glu Asp Val Glu Asn Ile Leu Leu Lys Leu Leu Gln Ala Ala Asp
 165 170 175
 Phe Asp Val Glu Arg Ala Gln Arg Gly Ile Ile Tyr Ile Asp Glu Val
 180 185 190

Asp Lys Ile Ser Arg Lys Ser Glu Asn Pro Ser Ile Thr Arg Asp Val
 195 200 205
 Ser Gly Glu Gly Val Gln Gln Ala Leu Leu Lys Ile Leu Glu Gly Thr
 210 215 220
 Val Ala Ala Ile Pro Pro Gln Gly Gly Arg Lys His Pro Asn Gln Asp
 225 230 235 240
 Phe Ile Gln Leu Asp Thr Thr Asn Ile Leu Phe Ile Val Ala Gly Ala
 245 250 255
 Phe Ser Gly Leu Glu Lys Val Ile Ala Asp Arg Asn Gly Lys Lys Gly
 260 265 270
 Leu Gly Phe Gly Val Glu Val Ser Ser Lys Lys Glu Glu Ala Asn Ile
 275 280 285
 Val Asp Ile Phe Lys Asp Val Leu Pro Glu Asp Leu Val Lys Phe Gly
 290 295 300
 Leu Ile Pro Glu Phe Ile Gly Arg Leu Pro Val Val Ala Thr Val Ser
 305 310 315 320
 Asn Leu Asp Gln Lys Ser Leu Val Lys Val Leu Thr Glu Pro Arg Asn
 325 330 335
 Ser Leu Val Lys Gln Tyr Arg Arg Leu Phe Glu Met Asp Asp Ala Val
 340 345 350
 Leu Thr Phe Thr Asp Asp Ala Leu Glu Glu Ile Ala Asn Gln Ala Leu
 355 360 365
 Glu Arg Lys Thr Gly Ala Arg Gly Leu Arg Ala Ile Met Glu Glu Ile
 370 375 380
 Leu Val Pro Ile Met Tyr Asp Leu Pro Asp Arg Lys Asp Val Gly Glu
 385 390 395 400
 Val Ile Ile Asn Gly Ala Val Ala Arg Gly Glu Ala Glu Pro Glu Met
 405 410 415
 Leu Glu Ala Val Ala Glu Glu Lys Thr Ala
 420 425

<210> 71
 <211> 1401
 <212> DNA
 <213> *Corynebacterium glutamicum*

<220>
 <221> CDS
 <222> (101)..(1378)
 <223> FRXA01120

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																1 5			
gaa agc gcc gat ctg ctc aaa tgt tcc ttc tgc gga aag agc caa aag																163			
Glu Ser Ala Asp Leu Leu Lys Cys Ser Phe Cys Gly Lys Ser Gln Lys																10 15 20			
cag gta aaa aaa ctc atc gcg ggt ggc gcc gta tat atc tgt gat gag																211			
Gln Val Lys Lys Leu Ile Ala Gly Gly Ala Val Tyr Ile Cys Asp Glu																25 30 35			
tgc att gag ctg tgc aac gag att att gaa gaa gaa ctc ggt caa gct																259			
Cys Ile Glu Leu Cys Asn Glu Ile Ile Glu Glu Glu Leu Gly Gln Ala																40 45 50			
caa cac gac gag cag gag cgc aac gag ctc ccc aag ccg tcg gag att																307			
Gln His Asp Glu Gln Glu Arg Asn Glu Leu Pro Lys Pro Ser Glu Ile																55 60 65			
tca gcc ttc ctt gat act tat gtc atc ggg cag gac cca gca aaa cgt																355			
Ser Ala Phe Leu Asp Thr Tyr Val Ile Gly Gln Asp Pro Ala Lys Arg																70 75 80 85			
atc ctg tcg gtt gcg gtg tac aac cat tac aag cgt ctc cgc gca tcg																403			
Ile Leu Ser Val Ala Val Tyr Asn His Tyr Lys Arg Leu Arg Ala Ser																90 95 100			
gaa acc atc ggt cgt cgc agg aat gac gag cct gaa acc gaa ctg gtt																451			
Glu Thr Ile Gly Arg Arg Arg Asn Asp Glu Pro Glu Thr Glu Leu Val																105 110 115			
aag tcc aat att ttg atg ctc ggc ccc act ggc tcc ggc aag act ttc																499			
Lys Ser Asn Ile Leu Met Leu Gly Pro Thr Gly Ser Gly Lys Thr Phe																120 125 130			
ctt gcc cag act ttg gca aag ctg ctg gat gtt cct ttt gct atc gcg																547			
Leu Ala Gln Thr Leu Ala Lys Leu Leu Asp Val Pro Phe Ala Ile Ala																135 140 145			
gat gcc acc tca ctg acc gag gct ggt tat gtg ggc gag gat gtg gaa																595			
Asp Ala Thr Ser Leu Thr Glu Ala Gly Tyr Val Gly Glu Asp Val Glu																150 155 160 165			
aac atc ttg ctc aag ctg ctt cag gct gct gat ttt gat gtg gaa cgt																643			
Asn Ile Leu Leu Lys Leu Leu Gln Ala Ala Asp Phe Asp Val Glu Arg																170 175 180			
gca cag cgc ggc atc att tac atc gat gaa gtg gac aag att tcc cgc																691			
Ala Gln Arg Gly Ile Ile Tyr Ile Asp Glu Val Asp Lys Ile Ser Arg																185 190 195			
aag tct gaa aac cca tcg atc act cgc gat gtt tcc ggt gaa ggc gtg																739			
Lys Ser Glu Asn Pro Ser Ile Thr Arg Asp Val Ser Gly Glu Gly Val																200 205 210			
cag cag gca ctg ctg aaa att ttg gaa ggc act gtc gcc gca atc cca																787			
Gln Gln Ala Leu Leu Lys Ile Leu Glu Gly Thr Val Ala Ala Ile Pro																215 220 225			
ccg cag gga gga cgc aag cac ccc aac cag gat ttc atc cag ctg gat																835			
Pro Gln Gly Gly Arg Lys His Pro Asn Gln Asp Phe Ile Gln Leu Asp																			

230	235	240	245	
acc acc aac att ttg ttc atc gtt gct ggt gcg ttc tct ggt ctg gag	Thr Thr Asn Ile Leu Phe Ile Val Ala Gly Ala Phe Ser Gly Leu Glu	883		
	250	255	260	
aag gtc atc gcg gac cgc aat ggc aag aaa ggc ttg ggc ttc ggt gtg	Lys Val Ile Ala Asp Arg Asn Gly Lys Lys Gly Leu Gly Phe Gly Val	931		
	265	270	275	
gag gtc tct tcc aag aag gaa gaa gcc aac att gtg gat atc ttc aag	Glu Val Ser Ser Lys Lys Glu Glu Ala Asn Ile Val Asp Ile Phe Lys	979		
	280	285	290	
gat gtc ctc cct gag gac ctg gtg aag ttt ggt ctc atc cca gaa ttc	Asp Val Leu Pro Glu Asp Leu Val Lys Phe Gly Leu Ile Pro Glu Phe	1027		
	295	300	305	
att ggg cgt ctg cca gtc gtt gcc acc gta tcc aac ctg gat cag aaa	Ile Gly Arg Leu Pro Val Val Ala Thr Val Ser Asn Leu Asp Gln Lys	1075		
	310	315	320	325
tct ctg gtc aag gtt ctc acg gag cct cgt aac tca ttg gtg aag cag	Ser Leu Val Lys Val Leu Thr Glu Pro Arg Asn Ser Leu Val Lys Gln	1123		
	330	335	340	
tat cga cgt ctg ttt gaa atg gat gac gct gtg ttg acc ttt act gat	Tyr Arg Arg Leu Phe Glu Met Asp Ala Val Leu Thr Phe Thr Asp	1171		
	345	350	355	
gat gct ttg gag gag atc gct aat cag gca ctc gag cgc aaa act ggc	Asp Ala Leu Glu Glu Ile Ala Asn Gln Ala Leu Glu Arg Lys Thr Gly	1219		
	360	365	370	
gcc cgt ggc ctg cgc gcg atc atg gaa gag atc ctg gtt ccg atc atg	Ala Arg Gly Leu Arg Ala Ile Met Glu Glu Ile Leu Val Pro Ile Met	1267		
	375	380	385	
tat gac ctc cca gac cgt aaa gac gtt ggc gaa gtc atc atc aac ggt	Tyr Asp Leu Pro Asp Arg Lys Asp Val Gly Glu Val Ile Ile Asn Gly	1315		
	390	395	400	405
gcc gtt gcc cgt ggc gaa gcc gaa cca gag atg ttg gaa gct gtc gca	Ala Val Ala Arg Gly Glu Ala Glu Pro Glu Met Leu Glu Ala Val Ala	1363		
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	425			

<210> 72
 <211> 426
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 72
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 Gly Lys Ser Gln Lys Gln Val Lys Lys Leu Ile Ala Gly Gly Ala Val

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Tyr	Ile	Cys	Asp	Glu	Cys	Ile	Glu	Leu	Cys	Asn	Glu	Ile	Ile	Glu	Glu				
35			40							45									
Glu	Leu	Gly	Gln	Ala	Gln	His	Asp	Glu	Gln	Glu	Arg	Asn	Glu	Leu	Pro				
50			55							60									
Lys	Pro	Ser	Glu	Ile	Ser	Ala	Phe	Leu	Asp	Thr	Tyr	Val	Ile	Gly	Gln				
65			70							75			80						
Asp	Pro	Ala	Lys	Arg	Ile	Leu	Ser	Val	Ala	Val	Tyr	Asn	His	Tyr	Lys				
			85							90			95						
Arg	Leu	Arg	Ala	Ser	Glu	Thr	Ile	Gly	Arg	Arg	Arg	Asn	Asp	Glu	Pro				
			100							105			110						
Glu	Thr	Glu	Leu	Val	Lys	Ser	Asn	Ile	Leu	Met	Leu	Gly	Pro	Thr	Gly				
115			120							125									
Ser	Gly	Lys	Thr	Phe	Leu	Ala	Gln	Thr	Leu	Ala	Lys	Leu	Leu	Asp	Val				
130			135							140									
Pro	Phe	Ala	Ile	Ala	Asp	Ala	Thr	Ser	Leu	Thr	Glu	Ala	Gly	Tyr	Val				
145			150							155			160						
Gly	Glu	Asp	Val	Glu	Asn	Ile	Leu	Leu	Lys	Leu	Leu	Gln	Ala	Ala	Asp				
			165							170			175						
Phe	Asp	Val	Glu	Arg	Ala	Gln	Arg	Gly	Ile	Ile	Tyr	Ile	Asp	Glu	Val				
			180							185			190						
Asp	Lys	Ile	Ser	Arg	Lys	Ser	Glu	Asn	Pro	Ser	Ile	Thr	Arg	Asp	Val				
195			200							205									
Ser	Gly	Glu	Gly	Val	Gln	Gln	Ala	Leu	Leu	Lys	Ile	Leu	Glu	Gly	Thr				
210			215							220									
Val	Ala	Ala	Ile	Pro	Pro	Gln	Gly	Gly	Arg	Lys	His	Pro	Asn	Gln	Asp				
225			230							235			240						
Phe	Ile	Gln	Leu	Asp	Thr	Thr	Asn	Ile	Leu	Phe	Ile	Val	Ala	Gly	Ala				
			245							250			255						
Phe	Ser	Gly	Leu	Glu	Lys	Val	Ile	Ala	Asp	Arg	Asn	Gly	Lys	Lys	Gly				
260			265							270									
Leu	Gly	Phe	Gly	Val	Glu	Val	Ser	Ser	Lys	Lys	Glu	Glu	Ala	Asn	Ile				
275			280							285									
Val	Asp	Ile	Phe	Lys	Asp	Val	Leu	Pro	Glu	Asp	Leu	Val	Lys	Phe	Gly				
290			295							300									
Leu	Ile	Pro	Glu	Phe	Ile	Gly	Arg	Leu	Pro	Val	Val	Ala	Thr	Val	Ser				
305			310							315			320						
Asn	Leu	Asp	Gln	Lys	Ser	Leu	Val	Lys	Val	Leu	Thr	Glu	Pro	Arg	Asn				
			325							330			335						
Ser	Leu	Val	Lys	Gln	Tyr	Arg	Arg	Leu	Phe	Glu	Met	Asp	Asp	Ala	Val				
340			345							350									

Leu Thr Phe Thr Asp Asp Ala Leu Glu Glu Ile Ala Asn Gln Ala Leu
 355 360 365
 Glu Arg Lys Thr Gly Ala Arg Gly Leu Arg Ala Ile Met Glu Glu Ile
 370 375 380
 Leu Val Pro Ile Met Tyr Asp Leu Pro Asp Arg Lys Asp Val Gly Glu
 385 390 395 400
 Val Ile Ile Asn Gly Ala Val Ala Arg Gly Glu Ala Glu Pro Glu Met
 405 410 415
 Leu Glu Ala Val Ala Glu Glu Lys Thr Ala
 420 425

<210> 73
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 <213> Corynebacterium glutamicum

<220>
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 <223> RXA00744

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 Met Gly Asn Trp Ala
 1 5
 gag att act gat gaa att tct aag att tac caa gat aat cag tac aag 163
 Glu Ile Thr Asp Glu Ile Ser Lys Ile Tyr Gln Asp Asn Gln Tyr Lys
 10 15 20
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 Ile Arg Gln Ile Asn Asp Val Asp Ala Val Ser Asp Lys Arg Arg Glu
 25 30 35
 gcg cta caa gca ctg ttt gaa cat act ggt cga aat gta atc gtc tat 259
 Ala Leu Gln Ala Leu Phe Glu His Thr Gly Arg Asn Val Ile Val Tyr
 40 45 50
 tat tca gcg tgg tta gaa aat ggt cga cga ttt tcc ggg caa tct acg 307
 Tyr Ser Ala Trp Leu Glu Asn Gly Arg Arg Phe Ser Gly Gln Ser Thr
 55 60 65
 gat ttt tcg gta aat gat act gat aaa aac agt ttt atg act gcg ctc 355
 Asp Phe Ser Val Asn Asp Thr Asp Lys Asn Ser Phe Met Thr Ala Leu
 70 75 80 85
 cat aag ttg gat cag agt aaa ggt ctc gat ctt atc ctc cac act ccg 403
 His Lys Leu Asp Gln Ser Lys Gly Leu Asp Leu Ile Leu His Thr Pro
 90 95 100
 ggt gga gat gtt gct gcg aca gag tcg tta gta gat tac att cac gca 451
 Gly Gly Asp Val Ala Ala Thr Glu Ser Leu Val Asp Tyr Ile His Ala
 105 110 115

ctc ttt ggt caa gat ttc aga gtc att gtc ccc caa ctc gca atg tca 499
 Leu Phe Gly Gln Asp Phe Arg Val Ile Val Pro Gln Leu Ala Met Ser
 120 125 130

gca gga aca atg atc gca ctt tcg tcc aaa gag att gtt atg ggg aag 547
 Ala Gly Thr Met Ile Ala Leu Ser Ser Lys Glu Ile Val Met Gly Lys
 135 140 145

cat tct agt ctt ggc ccc att gat cct cag ttt aac ggc cta ccg gca 595
 His Ser Ser Leu Gly Pro Ile Asp Pro Gln Phe Asn Gly Leu Pro Ala
 150 155 160 165

cac ggg tta ttg gaa gaa ttt gag caa gcg aag aaa gag gtc tct gag 643
 His Gly Leu Leu Glu Glu Phe Glu Gln Ala Lys Lys Glu Val Ser Glu
 170 175 180

aat ccg cag act gct cat ata tgg cag gtg atc ttg aat aaa tac aac 691
 Asn Pro Gln Thr Ala His Ile Trp Gln Val Ile Leu Asn Lys Tyr Asn
 185 190 195

ccc acg atg ttg ggt gaa gct aaa aaa gct att cag tgg tcc aac tcg 739
 Pro Thr Met Leu Gly Glu Ala Lys Lys Ala Ile Gln Trp Ser Asn Ser
 200 205 210

atg gtt aag cag tgg ctt gaa aag ggt atg ttt tta gac gag cct gac 787
 Met Val Lys Gln Trp Leu Glu Lys Gly Met Phe Leu Asp Glu Pro Asp
 215 220 225

aaa gaa gaa aaa gcc act cgc gct atc aaa gag ctc gct gat cat tcc 835
 Lys Glu Glu Lys Ala Thr Arg Ala Ile Lys Glu Leu Ala Asp His Ser
 230 235 240 245

gtt act ctt gcg cat aat cga cac att tcg gtc agt aaa gca ctt gag 883
 Val Thr Leu Ala His Asn Arg His Ile Ser Val Ser Lys Ala Leu Glu
 250 255 260

ctg gga ttg aat atc aaa gaa ctt gag agc gat cca aag ctt caa gat 931
 Leu Gly Leu Asn Ile Lys Glu Leu Glu Ser Asp Pro Lys Leu Gln Asp
 265 270 275

tta gtt ctt act ctt cac cac ctg tcc gtt att gct gcg caa cga gga 979
 Leu Val Leu Thr Leu His His Leu Ser Val Ile Ala Ala Gln Arg Gly
 280 285 290

cca tta att aag ttt gtc gtc aat cat gac aac cgt ggc act ttt ctg 1027
 Pro Leu Ile Lys Phe Val Val Asn His Asp Asn Arg Gly Thr Phe Leu
 295 300 305

cag ggg cat gaa aac taattaagt atgcaatagt cta 1065
 Gln Gly His Glu Asn
 310

<210> 74

<211> 314

<212> PRT

<213> Corynebacterium glutamicum

<400> 74

Met Gly Asn Trp Ala Glu Ile Thr Asp Glu Ile Ser Lys Ile Tyr Gln

1	5	10	15
Asp Asn Gln Tyr Lys Ile Arg Gln Ile Asn Asp Val Asp Ala Val Ser	20	25	30
Asp Lys Arg Arg Glu Ala Leu Gln Ala Leu Phe Glu His Thr Gly Arg	35	40	45
Asn Val Ile Val Tyr Tyr Ser Ala Trp Leu Glu Asn Gly Arg Arg Phe	50	55	60
Ser Gly Gln Ser Thr Asp Phe Ser Val Asn Asp Thr Asp Lys Asn Ser	65	70	75
Phe Met Thr Ala Leu His Lys Leu Asp Gln Ser Lys Gly Leu Asp Leu	85	90	95
Ile Leu His Thr Pro Gly Gly Asp Val Ala Ala Thr Glu Ser Leu Val	100	105	110
Asp Tyr Ile His Ala Leu Phe Gly Gln Asp Phe Arg Val Ile Val Pro	115	120	125
Gln Leu Ala Met Ser Ala Gly Thr Met Ile Ala Leu Ser Ser Lys Glu	130	135	140
Ile Val Met Gly Lys His Ser Ser Leu Gly Pro Ile Asp Pro Gln Phe	145	150	155
Asn Gly Leu Pro Ala His Gly Leu Leu Glu Glu Phe Glu Gln Ala Lys	165	170	175
Lys Glu Val Ser Glu Asn Pro Gln Thr Ala His Ile Trp Gln Val Ile	180	185	190
Leu Asn Lys Tyr Asn Pro Thr Met Leu Gly Glu Ala Lys Lys Ala Ile	195	200	205
Gln Trp Ser Asn Ser Met Val Lys Gln Trp Leu Glu Lys Gly Met Phe	210	215	220
Leu Asp Glu Pro Asp Lys Glu Glu Lys Ala Thr Arg Ala Ile Lys Glu	225	230	235
Leu Ala Asp His Ser Val Thr Leu Ala His Asn Arg His Ile Ser Val	245	250	255
Ser Lys Ala Leu Glu Leu Gly Leu Asn Ile Lys Glu Leu Glu Ser Asp	260	265	270
Pro Lys Leu Gln Asp Leu Val Leu Thr Leu His His Leu Ser Val Ile	275	280	285
Ala Ala Gln Arg Gly Pro Leu Ile Lys Phe Val Val Asn His Asp Asn	290	295	300
Arg Gly Thr Phe Leu Gln Gly His Glu Asn	305	310	

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<211> 957
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
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 <223> RXA00844

<400> 75

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Met Ser Ser Ala Ser
1 5

ttt acc acc aaa gca ctg tcc gta ctc gca gct tta acg gct gcg tct 163
Phe Thr Thr Lys Ala Leu Ser Val Leu Ala Ala Leu Thr Ala Ala Ser
10 15 20

gcc ccc tta gtg gcg gcg tca cct gca cat gct ttg gca aat gct cgc 211
Ala Pro Leu Val Ala Ala Ser Pro Ala His Ala Leu Ala Asn Ala Arg
25 30 35

aac gtt acg ggt tca agc acc act tca gat tca att gtt cgt ctg cac 259
Asn Val Thr Gly Ser Ser Thr Thr Ser Asp Ser Ile Val Arg Leu His
40 45 50

atc ggt aac act gca tgt aca gga acc atg atc acc cca acg tgg gcg 307
Ile Gly Asn Thr Ala Cys Thr Gly Thr Met Ile Thr Pro Thr Trp Ala
55 60 65

atc acc gcc cgc cac tgt atc cct gag ggc ggt att gcc ggt gca gct 355
Ile Thr Ala Arg His Cys Ile Pro Glu Gly Gly Ile Ala Gly Ala Ala
70 75 80 85

att ggt tca agc act ttg agc caa ttt cag cag gtg tcc caa gcg atc 403
Ile Gly Ser Ser Thr Leu Ser Gln Phe Gln Val Ser Gln Ala Ile
90 95 100

ttg cac cct act gcg gac tta gct ctc gtt gag ctt ccc aat cag gca 451
Leu His Pro Thr Ala Asp Leu Ala Leu Val Glu Leu Pro Asn Gln Ala
105 110 115

agt tcc aac acg gtt gat ctc tac ggt gca cac gtg cag cct ggt gaa 499
Ser Ser Asn Thr Val Asp Leu Tyr Gly Ala His Val Gln Pro Gly Glu
120 125 130

aat ggt caa gca gcc ggc tgg ggt ggg tac tct gcc ttt ggc caa aat 547
Asn Gly Gln Ala Ala Gly Trp Gly Gly Tyr Ser Ala Phe Gly Gln Asn
135 140 145

gtt gca cag caa gcc gat gtg caa att caa cgc agg gta gtc aat gtg 595
Val Ala Gln Gln Ala Asp Val Gln Ile Gln Arg Val Val Asn Val
150 155 160 165

cca agc ccc gac cgc acc gct gtg ctg ctt gaa ggc act gtt tct aac 643
Pro Ser Pro Asp Arg Thr Ala Val Leu Leu Glu Gly Thr Val Ser Asn
170 175 180

ggt cgt ctc gta cca ggc gat tcc ggc gga cct ttg tac atc aat ggt 691

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Gly Arg Leu Val Pro Gly Asp Ser Gly Gly Pro Leu Tyr Ile Asn Gly
 185 190 195 739
 caa ctg gct ggt gtg ctc agc atg tcc act gac gta gaa aac gat gca
 Gln Leu Ala Gly Val Leu Ser Met Ser Thr Asp Val Glu Asn Asp Ala
 200 205 210
 cta gac ggc acc gtc ggc tgg tac atc ccc gtt gct gaa cac gcc gag 787
 Leu Asp Gly Thr Val Gly Trp Tyr Ile Pro Val Ala Glu His Ala Glu
 215 220 225
 tgg atc gcc tac tac acc ggc aag cac att gcc ccc att gct ggt gcg 835
 Trp Ile Ala Tyr Tyr Thr Gly Lys His Ile Ala Pro Ile Ala Gly Ala
 230 235 240 245
 ccc gca gaa ctt gtt gac gcc acc gcc aac ccc acc ttc atc cct gct 883
 Pro Ala Glu Leu Val Asp Ala Thr Ala Asn Pro Thr Phe Ile Pro Ala
 250 255 260
 cca cag cct ttc acc ggt tca tcc atc ggt ggt tgg gcg ctg gcc agc 931
 Pro Gln Pro Phe Thr Gly Ser Ser Ile Gly Gly Trp Ala Leu Gly Ser
 265 270 275
 tcc tagaatatgc tgatctccct gct 957
 Ser

<210> 76

<211> 278

<212> PRT

<213> Corynebacterium glutamicum

<400> 76

Met Ser Ser Ala Ser Phe Thr Thr Lys Ala Leu Ser Val Leu Ala Ala
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 Leu Ala Asn Ala Arg Asn Val Thr Gly Ser Ser Thr Thr Ser Asp Ser
 35 40 45
 Ile Val Arg Leu His Ile Gly Asn Thr Ala Cys Thr Gly Thr Met Ile
 50 55 60
 Thr Pro Thr Trp Ala Ile Thr Ala Arg His Cys Ile Pro Glu Gly Gly
 65 70 75 80
 Ile Ala Gly Ala Ala Ile Gly Ser Ser Thr Leu Ser Gln Phe Gln Gln
 85 90 95
 Val Ser Gln Ala Ile Leu His Pro Thr Ala Asp Leu Ala Leu Val Glu
 100 105 110
 Leu Pro Asn Gln Ala Ser Ser Asn Thr Val Asp Leu Tyr Gly Ala His
 115 120 125
 Val Gln Pro Gly Glu Asn Gly Gln Ala Ala Gly Trp Gly Gly Tyr Ser
 130 135 140

Ala Phe Gly Gln Asn Val Ala Gln Gln Ala Asp Val Gln Ile Gln Arg
 145 150 155 160

Arg Val Val Asn Val Pro Ser Pro Asp Arg Thr Ala Val Leu Leu Glu
 165 170 175

Gly Thr Val Ser Asn Gly Arg Leu Val Pro Gly Asp Ser Gly Gly Pro
 180 185 190

Leu Tyr Ile Asn Gly Gln Leu Ala Gly Val Leu Ser Met Ser Thr Asp
 195 200 205

Val Glu Asn Asp Ala Leu Asp Gly Thr Val Gly Trp Tyr Ile Pro Val
 210 215 220

Ala Glu His Ala Glu Trp Ile Ala Tyr Tyr Thr Gly Lys His Ile Ala
 225 230 235 240

Pro Ile Ala Gly Ala Pro Ala Glu Leu Val Asp Ala Thr Ala Asn Pro
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Thr Phe Ile Pro Ala Pro Gln Pro Phe Thr Gly Ser Ser Ile Gly Gly
 260 265 270

Trp Ala Leu Gly Ser Ser
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 <211> 958
 <212> DNA
 <213> Corynebacterium glutamicum

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 <222> (101)..(958)
 <223> RXA01151

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 Met Ser Ser Pro Thr
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gat tct tcg ccc tct aat tct ttt agc gac ttc aac cgg gag gaa cag 163
 Asp Ser Ser Pro Ser Asn Ser Phe Ser Asp Phe Asn Arg Glu Glu Gln
 10 15 20

tcc cgg tta tct gat gag gtg cgc cag ctc aag cgc acc aac tct gat 211
 Ser Arg Leu Ser Asp Glu Val Arg Gln Leu Lys Arg Thr Asn Ser Asp
 25 30 35

ctt ggg gca cgt aat gcc aag ctc gcg gag atg ctg aag tcg tct cgg 259
 Leu Gly Ala Arg Asn Ala Lys Leu Ala Glu Met Leu Lys Ser Ser Arg
 40 45 50

gat aaa ttg tct gtg ctg ttt tct cag ttg gag gat atg gct cag cgg 307
 Asp Lys Leu Ser Val Leu Phe Ser Gln Leu Glu Asp Met Ala Gln Pro
 55 60 65

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Pro	Ser	Val	Tyr	Gly	Thr	Phe	Leu	Glu	Thr	Ala	Lys	Asp	Gly	Ser	Asn	
70				75					80						85	
gcg	gag	atc	ttt	gct	ggt	gga	cgt	cgc	atg	cgt	gtg	gct	gtt	tct	cct	403
Ala	Glu	Ile	Phe	Ala	Gly	Gly	Arg	Arg	Met	Arg	Val	Ala	Val	Ser	Pro	
			90					95						100		
atg	ctg	tgt	gcc	gcg	gat	ttg	atg	ccg	ggt	gtg	cag	gtt	cgt	ttg	ggt	451
Met	Leu	Cys	Ala	Ala	Asp	Leu	Met	Pro	Gly	Val	Gln	Val	Arg	Leu	Gly	
			105					110					115			
gaa	ggc	aat	caa	gtt	ctt	gag	gcc	tgt	gat	ttt	gaa	caa	acc	ggt	gaa	499
Glu	Gly	Asn	Gln	Val	Leu	Glu	Ala	Cys	Asp	Phe	Glu	Gln	Thr	Gly	Glu	
		120					125						130			
tta	gcc	acg	ttg	atg	gaa	atg	att	ggc	cgg	gat	cgt	gct	ttg	gtt	tca	547
Leu	Ala	Thr	Leu	Met	Glu	Met	Ile	Gly	Arg	Asp	Arg	Ala	Leu	Val	Ser	
		135					140					145				
gat	cgc	tcg	ggg	gag	gag	cgc	gtc	gtc	aag	ctt	gct	ggt	ccg	ttg	atg	595
Asp	Arg	Ser	Gly	Glu	Glu	Arg	Val	Val	Lys	Leu	Ala	Gly	Pro	Leu	Met	
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gat	cgc	acc	gca	aag	ctg	ccg	cgc	ccc	ggt	gac	acc	ctg	ctt	gtt	gac	643
Asp	Arg	Thr	Ala	Lys	Leu	Pro	Arg	Pro	Gly	Asp	Thr	Leu	Leu	Val	Asp	
			170					175						180		
cgc	aaa	gcg	ggc	tac	gct	ttt	gag	gcg	att	gcc	aag	acg	gaa	att	tcg	691
Arg	Lys	Ala	Gly	Tyr	Ala	Phe	Glu	Ala	Ile	Ala	Lys	Thr	Glu	Ile	Ser	
			185					190					195			
agg	ctt	gcg	ctg	gaa	gag	gcg	cca	gat	gtg	tct	tat	cag	gat	att	ggt	739
Arg	Leu	Ala	Leu	Glu	Glu	Ala	Pro	Asp	Val	Ser	Tyr	Gln	Asp	Ile	Gly	
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ggc	ttg	gat	gat	cag	att	gaa	ttg	att	caa	gat	gcc	gtt	gag	ctg	cca	787
Gly	Leu	Asp	Asp	Gln	Ile	Glu	Leu	Ile	Gln	Asp	Ala	Val	Glu	Leu	Pro	
		215				220					225					
ttt	ttg	cac	ccg	gag	atg	tac	cgc	gcc	tac	aac	ctg	cat	cca	cca	aag	835
Phe	Leu	His	Pro	Glu	Met	Tyr	Arg	Ala	Tyr	Asn	Leu	His	Pro	Pro	Lys	
230				235						240					245	
ggc	gtg	ctg	ctg	tac	ggc	cct	ccc	ggc	tgt	gga	aag	acg	ctg	att	gct	883
Gly	Val	Leu	Leu	Tyr	Gly	Pro	Pro	Gly	Cys	Gly	Lys	Thr	Leu	Ile	Ala	
			250					255						260		
aag	gct	gtg	gct	aat	tct	ttg	gcc	aac	cgc	atc	ggt	gag	act	ggc	acc	931
Lys	Ala	Val	Ala	Asn	Ser	Leu	Ala	Asn	Arg	Ile	Gly	Glu	Thr	Gly	Thr	
			265					270					275			
tcg	tac	ttc	atc	aac	gtc	aag	ggg	cca								958
Ser	Tyr	Phe	Ile	Asn	Val	Lys	Gly	Pro								
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<211> 286

<212> PRT

<213> Corynebacterium glutamicum

<400> 78

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Arg Thr Asn Ser Asp Leu Gly Ala Arg Asn Ala Lys Leu Ala Glu Met
 35           40           45

Leu Lys Ser Ser Arg Asp Lys Leu Ser Val Leu Phe Ser Gln Leu Glu
 50           55           60

Asp Met Ala Gln Pro Pro Ser Val Tyr Gly Thr Phe Leu Glu Thr Ala
 65           70           75           80

Lys Asp Gly Ser Asn Ala Glu Ile Phe Ala Gly Gly Arg Arg Met Arg
 85           90           95

Val Ala Val Ser Pro Met Leu Cys Ala Ala Asp Leu Met Pro Gly Val
 100          105          110

Gln Val Arg Leu Gly Glu Gly Asn Gln Val Leu Glu Ala Cys Asp Phe
 115          120          125

Glu Gln Thr Gly Glu Leu Ala Thr Leu Met Glu Met Ile Gly Arg Asp
 130          135          140

Arg Ala Leu Val Ser Asp Arg Ser Gly Glu Glu Arg Val Val Lys Leu
 145          150          155          160

Ala Gly Pro Leu Met Asp Arg Thr Ala Lys Leu Pro Arg Pro Gly Asp
 165          170          175

Thr Leu Leu Val Asp Arg Lys Ala Gly Tyr Ala Phe Glu Ala Ile Ala
 180          185          190

Lys Thr Glu Ile Ser Arg Leu Ala Leu Glu Glu Ala Pro Asp Val Ser
 195          200          205

Tyr Gln Asp Ile Gly Gly Leu Asp Asp Gln Ile Glu Leu Ile Gln Asp
 210          215          220

Ala Val Glu Leu Pro Phe Leu His Pro Glu Met Tyr Arg Ala Tyr Asn
 225          230          235          240

Leu His Pro Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro Gly Cys Gly
 245          250          255

Lys Thr Leu Ile Ala Lys Ala Val Ala Asn Ser Leu Ala Asn Arg Ile
 260          265          270

Gly Glu Thr Gly Thr Ser Tyr Phe Ile Asn Val Lys Gly Pro
 275          280          285

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<210> 79

<211> 735

<212> DNA

<213> *Corynebacterium glutamicum*

<220>

<221> CDS

<222> (101)..(712)

<223> RXA02317

<400> 79

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gacgtcgaaa agcgaattca tggcccatc ttgccttaaa atg gcg cac atg cgc 115
Met Ala His Met Arg
1 5
tta ctg ctg acc tcc ttt ggc cat gat cat att cgg gat ttt gta cgc 163
Leu Leu Leu Thr Ser Phe Gly His Asp His Ile Arg Asp Phe Val Arg
10 15 20
ggt acc gtg gcg tat atc cct gat gcg acc agg ctt ttt gct gat agt 211
Gly Thr Val Ala Tyr Ile Pro Asp Ala Thr Arg Leu Phe Ala Asp Ser
25 30 35
ccc gag gct gct cct ttt atg gag acg gag cga aat atg ctg cgc gag 259
Pro Glu Ala Ala Pro Phe Met Glu Thr Glu Arg Asn Met Leu Arg Glu
40 45 50
cac ggc ttg agc att cgt gag ctg ccg att tcc acg tcg act ccg gag 307
His Gly Leu Ser Ile Arg Glu Leu Pro Ile Ser Thr Ser Thr Pro Glu
55 60 65
gaa gtg gat ccg gtg ctt ggt gag gtt gat ggg gtg tat gtg gcg ggc 355
Glu Val Asp Arg Val Leu Gly Glu Val Asp Gly Val Tyr Val Ala Gly
70 75 80 85
ggt gag act ttt gat ctg atg tgg ctg ctg cgt tcc aca ggc aat gat 403
Gly Glu Thr Phe Asp Leu Met Trp Leu Leu Arg Ser Thr Gly Asn Asp
90 95 100
gag gtg ttg att aag cat gtt cgc gct ggt cta ccg tat att gga acg 451
Glu Val Leu Ile Lys His Val Arg Ala Gly Leu Pro Tyr Ile Gly Thr
105 110 115
agc gcc ggc gcg gta att gca ggt cct tcg att gaa ccg atc agc ttt 499
Ser Ala Gly Ala Val Ile Ala Gly Pro Ser Ile Glu Pro Ile Ser Phe
120 125 130
ttg gat agc ccc gat gtc gcg ccg aat tta agc gac tat tca ggt cta 547
Leu Asp Ser Pro Asp Val Ala Pro Asn Leu Ser Asp Tyr Ser Gly Leu
135 140 145
ggc ctg tgc gag cat gtc gtg gtg ccc cat gct ggt ggc acg atc ccg 595
Gly Leu Cys Glu His Val Val Val Pro His Ala Gly Gly Thr Ile Pro
150 155 160 165
caa ttt ccc atc gat gtg ttt gcg gaa acc gtg cgc acc tac ggc gcc 643
Gln Phe Pro Ile Asp Val Phe Ala Glu Thr Val Arg Thr Tyr Gly Ala
170 175 180
gaa ttc ccg ctg gtc ctg ctt aaa gat gga cag gca ctg ctt atc gac 691
Glu Phe Pro Leu Val Leu Leu Lys Asp Gly Gln Ala Leu Leu Ile Asp
185 190 195

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gac cac ggc gtc cac cta att taggatgggtt ccccatgagc acc
 Asp His Gly Val His Leu Ile
 200

735

<210> 80
 <211> 204
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 80
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 Arg Asp Phe Val Arg Gly Thr Val Ala Tyr Ile Pro Asp Ala Thr Arg
 20 25 30
 Leu Phe Ala Asp Ser Pro Glu Ala Ala Pro Phe Met Glu Thr Glu Arg
 35 40 45
 Asn Met Leu Arg Glu His Gly Leu Ser Ile Arg Glu Leu Pro Ile Ser
 50 55 60
 Thr Ser Thr Pro Glu Glu Val Asp Arg Val Leu Gly Glu Val Asp Gly
 65 70 75 80
 Val Tyr Val Ala Gly Gly Glu Thr Phe Asp Leu Met Trp Leu Leu Arg
 85 90 95
 Ser Thr Gly Asn Asp Glu Val Leu Ile Lys His Val Arg Ala Gly Leu
 100 105 110
 Pro Tyr Ile Gly Thr Ser Ala Gly Ala Val Ile Ala Gly Pro Ser Ile
 115 120 125
 Glu Pro Ile Ser Phe Leu Asp Ser Pro Asp Val Ala Pro Asn Leu Ser
 130 135 140
 Asp Tyr Ser Gly Leu Gly Leu Cys Glu His Val Val Val Pro His Ala
 145 150 155 160
 Gly Gly Thr Ile Pro Gln Phe Pro Ile Asp Val Phe Ala Glu Thr Val
 165 170 175
 Arg Thr Tyr Gly Ala Glu Phe Pro Leu Val Leu Leu Lys Asp Gly Gln
 180 185 190
 Ala Leu Leu Ile Asp Asp His Gly Val His Leu Ile
 195 200

<210> 81
 <211> 774
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(751)
 <223> RXA02644

215

<210> 82
 <211> 217
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 82

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Glu His Glu Leu Glu Ile Lys Arg Ser Arg Phe Leu Thr Tyr Ile Thr
      20              25              30

Arg Val Gln Asp Gln Glu Gln Ala Arg Glu Phe Ile His Ser Ile Lys
      35              40              45

Glu Leu Tyr Pro Asp Ala Arg His His Cys Ser Ala Phe Ile Phe His
 50              55              60

Val Asp Gly Ser Asn Asp Val Glu Arg Ser Ser Asp Asp Gly Glu Pro
 65              70              75              80

Ser Gly Thr Ala Gly Lys Pro Met Leu Glu Ala Leu Arg Gly Ser Gly
      85              90              95

Met Lys Asp Ile Ala Ala Val Val Val Arg Tyr Phe Gly Gly Val Lys
      100              105              110

Leu Gly Thr Gly Gly Leu Val Asn Ala Tyr Thr Asn Ala Val Thr Glu
 115              120              125

Leu Leu Pro Glu Val Leu Gln Val Thr Arg Ser Val Arg Glu Ile Phe
 130              135              140

Lys Ile Asp Leu Pro His Ser Asp Ala Gly Arg Ile Glu Ala Asn Leu
 145              150              155              160

Arg Gly Met Gly Ile Ile Ile Thr Asp Thr Glu Tyr Gly Ala Glu Val
      165              170              175

Thr Tyr Thr Leu Ala Leu Leu Pro Gly Glu Gln Ala Ala Val Glu Ser
      180              185              190

Gln Leu Ser Ser Met Met Gly Ala Glu Ile Glu Leu Lys Glu Ser Gly
 195              200              205

His Met Trp Val Glu Ser Pro Ser Asp
 210              215

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<210> 83
 <211> 1411
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(1411)
 <223> RXN02820

<400> 83

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                                         Met Lys Val Thr Gln
                                         1           5

agc aca ttc ctt aaa tcg gta gct gcg ttc act gtc gca gcc tta acc 163
Ser Thr Phe Leu Lys Ser Val Ala Ala Phe Thr Val Ala Ala Leu Thr
              10              15              20

ctg acc atc tct tcg tgt tcc agc ggt gaa gac acc tcc gca agc tcc 211
Leu Thr Ile Ser Ser Cys Ser Ser Gly Glu Asp Thr Ser Ala Ser Ser
              25              30              35

acg gat act gaa aac tcc tca acc caa gca gca gcg tct ccc cca ctt 259
Thr Asp Thr Glu Asn Ser Ser Thr Gln Ala Ala Ala Ser Pro Pro Leu
              40              45              50

gcg cct tgt gaa ctt occ gcc gac gct tct gct gaa gag gaa gta gaa 307
Ala Pro Cys Glu Leu Pro Ala Asp Ala Ser Ala Glu Glu Glu Val Glu
              55              60              65

ggc act cac aca ggt gaa gat att tct gtt gcc ccg gaa atc ggt acc 355
Gly Thr His Thr Gly Glu Asp Ile Ser Val Ala Pro Glu Ile Gly Thr
              70              75              80              85

ggc tac cgc gag ggc atg acc cct gtt caa acc caa ggt tat gcg gtg 403
Gly Tyr Arg Glu Glu Met Thr Pro Val Gln Thr Gln Gly Tyr Ala Val
              90              95              100

gca act gca aac ccc atc gct tct gaa gca gcc tgc gcg gtg tta aga 451
Ala Thr Ala Asn Pro Ile Ala Ser Glu Ala Ala Cys Ala Val Leu Arg
              105              110              115

gaa ggc ggc act gca gct gat gct ctt gtc acc gcg cag ttt gtt ttg 499
Glu Gly Gly Thr Ala Ala Asp Ala Leu Val Thr Ala Gln Phe Val Leu
              120              125              130

gga ctg acg gaa ccg cag tcg tct ggc ctt ggt ggt ggc gga tac att 547
Gly Leu Thr Glu Pro Gln Ser Ser Gly Leu Gly Gly Gly Tyr Ile
              135              140              145

ctg tac tac gac gcc gaa gcc aat gcg gtg aca gcc att gat ggc cgt 595
Leu Tyr Tyr Asp Ala Glu Ala Asn Ala Val Thr Ala Ile Asp Gly Arg
              150              155              160              165

gaa aca gcg cca gtt gct gct gat gaa aac tat ctc att cat gtt tct 643
Glu Thr Ala Pro Val Ala Ala Asp Glu Asn Tyr Leu Ile His Val Ser
              170              175              180

gca gag gat caa acg gca cct gtt cct gat gcc cga cgt tcc ggc agg 691
Ala Glu Asp Gln Thr Ala Pro Val Pro Asp Ala Arg Arg Ser Gly Arg
              185              190              195

tca att ggt gtg cca gga atc gtg gca gcc ctt gga cag ctg cat gat 739
Ser Ile Gly Val Pro Gly Ile Val Ala Ala Leu Gly Gln Leu His Asp
              200              205              210

tca ttc gga aag acc tcc tgg cag gac gtg ctg aca act ccg cag cag 787

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Ser	Phe	Gly	Lys	Thr	Ser	Trp	Gln	Asp	Val	Leu	Thr	Thr	Pro	Gln	Gln		
215						220					225						
ctc	gca	act	gat	ggg	ttt	tcc	atc	agc	cct	cgc	atg	tca	gca	tca	att	835	
Leu	Ala	Thr	Asp	Gly	Phe	Ser	Ile	Ser	Pro	Arg	Met	Ser	Ala	Ser	Ile		
230					235					240					245		
gct	aac	tcc	gct	gag	gat	ctc	tcc	cac	gat	ccg	gaa	gct	gcc	gca	tat	883	
Ala	Asn	Ser	Ala	Glu	Asp	Leu	Ser	His	Asp	Pro	Glu	Ala	Ala	Ala	Tyr		
				250					255					260			
ttc	ctt	gat	gaa	aac	ggg	gat	gcg	aag	gca	ccc	ggc	aca	ctt	tta	caa	931	
Phe	Leu	Asp	Glu	Asn	Gly	Asp	Ala	Lys	Ala	Pro	Gly	Thr	Leu	Leu	Gln		
			265					270					275				
aac	cct	gac	tat	gca	gaa	acg	att	cgt	ctc	atc	tct	gaa	ggg	ggc	ccc	979	
Asn	Pro	Asp	Tyr	Ala	Glu	Thr	Ile	Arg	Leu	Ile	Ser	Glu	Gly	Gly	Pro		
		280					285					290					
gat	gcg	ttc	tac	acg	ggg	gag	att	gca	gca	gac	atc	gtg	gaa	cgc	gcc	1027	
Asp	Ala	Phe	Tyr	Thr	Gly	Glu	Ile	Ala	Ala	Asp	Ile	Val	Glu	Arg	Ala		
	295					300					305						
acc	cgt	gag	gtt	gac	ggg	ttc	aca	cca	tca	ctg	atg	agc	acg	gca	gat	1075	
Thr	Arg	Glu	Val	Asp	Gly	Phe	Thr	Pro	Ser	Leu	Met	Ser	Thr	Ala	Asp		
310					315					320					325		
ttg	gct	gcc	tac	act	ccg	gaa	act	cgt	gaa	gct	ttg	tgt	gct	ccc	tac	1123	
Leu	Ala	Ala	Tyr	Thr	Pro	Glu	Thr	Arg	Glu	Ala	Leu	Cys	Ala	Pro	Tyr		
				330					335					340			
cgc	gac	aag	att	gtt	tgt	ggc	atg	cca	ccg	tca	tca	tcg	ggg	ggc	gtc	1171	
Arg	Asp	Lys	Ile	Val	Cys	Gly	Met	Pro	Pro	Ser	Ser	Ser	Gly	Gly	Val		
		345						350					355				
aca	gtg	atg	gaa	acc	ctg	ggg	atc	ttg	aac	aac	ttt	gat	ctc	gcc	caa	1219	
Thr	Val	Met	Glu	Thr	Leu	Gly	Ile	Leu	Asn	Asn	Phe	Asp	Leu	Ala	Gln		
		360				365						370					
tac	cca	ccc	act	gag	gtt	ggg	ttg	gat	ggc	gga	ttg	cca	aat	gcg	gaa	1267	
Tyr	Pro	Pro	Thr	Glu	Val	Gly	Leu	Asp	Gly	Gly	Leu	Pro	Asn	Ala	Glu		
		375				380					385						
gct	gtt	cac	ctg	att	tca	gag	gct	gag	cgc	ctg	gct	tat	gct	gat	cgc	1315	
Ala	Val	His	Leu	Ile	Ser	Glu	Ala	Glu	Arg	Leu	Ala	Tyr	Ala	Asp	Arg		
390					395				400						405		
gat	gct	tac	atc	ggg	gat	cct	gct	ttc	gtg	gaa	gtt	cca	gca	ggg	ggg	1363	
Asp	Ala	Tyr	Ile	Gly	Asp	Pro	Ala	Phe	Val	Glu	Val	Pro	Ala	Gly	Gly		
				410				415						420			
gtc	caa	cag	tgg	atc	aac	cat	gtc	cac	acg	ggc	gaa	cac	tcc	aaa	ctt	1411	
Val	Gln	Gln	Trp	Ile	Asn	His	Val	His	Thr	Gly	Glu	His	Ser	Lys	Leu		
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<210> 84

<211> 437

<212> PRT

<213> Corynebacterium glutamicum

<400> 84

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 20 25 30
 Thr Ser Ala Ser Ser Thr Asp Thr Glu Asn Ser Ser Thr Gln Ala Ala
 35 40 45
 Ala Ser Pro Pro Leu Ala Pro Cys Glu Leu Pro Ala Asp Ala Ser Ala
 50 55 60
 Glu Glu Glu Val Glu Gly Thr His Thr Gly Glu Asp Ile Ser Val Ala
 65 70 75 80
 Pro Glu Ile Gly Thr Gly Tyr Arg Glu Gly Met Thr Pro Val Gln Thr
 85 90 95
 Gln Gly Tyr Ala Val Ala Thr Ala Asn Pro Ile Ala Ser Glu Ala Ala
 100 105 110
 Cys Ala Val Leu Arg Glu Gly Gly Thr Ala Ala Asp Ala Leu Val Thr
 115 120 125
 Ala Gln Phe Val Leu Gly Leu Thr Glu Pro Gln Ser Ser Gly Leu Gly
 130 135 140
 Gly Gly Gly Tyr Ile Leu Tyr Tyr Asp Ala Glu Ala Asn Ala Val Thr
 145 150 155 160
 Ala Ile Asp Gly Arg Glu Thr Ala Pro Val Ala Ala Asp Glu Asn Tyr
 165 170 175
 Leu Ile His Val Ser Ala Glu Asp Gln Thr Ala Pro Val Pro Asp Ala
 180 185 190
 Arg Arg Ser Gly Arg Ser Ile Gly Val Pro Gly Ile Val Ala Ala Leu
 195 200 205
 Gly Gln Leu His Asp Ser Phe Gly Lys Thr Ser Trp Gln Asp Val Leu
 210 215 220
 Thr Thr Pro Gln Gln Leu Ala Thr Asp Gly Phe Ser Ile Ser Pro Arg
 225 230 235 240
 Met Ser Ala Ser Ile Ala Asn Ser Ala Glu Asp Leu Ser His Asp Pro
 245 250 255
 Glu Ala Ala Ala Tyr Phe Leu Asp Glu Asn Gly Asp Ala Lys Ala Pro
 260 265 270
 Gly Thr Leu Leu Gln Asn Pro Asp Tyr Ala Glu Thr Ile Arg Leu Ile
 275 280 285
 Ser Glu Gly Gly Pro Asp Ala Phe Tyr Thr Gly Glu Ile Ala Ala Asp
 290 295 300
 Ile Val Glu Arg Ala Thr Arg Glu Val Asp Gly Phe Thr Pro Ser Leu
 305 310 315 320

Met Ser Thr Ala Asp Leu Ala Ala Tyr Thr Pro Glu Thr Arg Glu Ala
 325 330 335

Leu Cys Ala Pro Tyr Arg Asp Lys Ile Val Cys Gly Met Pro Pro Ser
 340 345 350

Ser Ser Gly Gly Val Thr Val Met Glu Thr Leu Gly Ile Leu Asn Asn
 355 360 365

Phe Asp Leu Ala Gln Tyr Pro Pro Thr Glu Val Gly Leu Asp Gly Gly
 370 375 380

Leu Pro Asn Ala Glu Ala Val His Leu Ile Ser Glu Ala Glu Arg Leu
 385 390 395 400

Ala Tyr Ala Asp Arg Asp Ala Tyr Ile Gly Asp Pro Ala Phe Val Glu
 405 410 415

Val Pro Ala Gly Gly Val Gln Gln Trp Ile Asn His Val His Thr Gly
 420 425 430

Glu His Ser Lys Leu
 435

<210> 85

<211> 507

<212> DNA

<213> *Corynebacterium glutamicum*

<220>

<221> CDS

<222> (1)..(507)

<223> FRXA02820

<400> 85

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 Ala Asn Ser Ala Glu Asp Leu Ser His Asp Pro Glu Ala Ala Ala Tyr
 1 5 10 15

ttc ctt gat gaa aac ggt gat gcg aag gca ccc ggc aca ctt tta caa 96
 Phe Leu Asp Glu Asn Gly Asp Ala Lys Ala Pro Gly Thr Leu Leu Gln
 20 25 30

aac cct gac tat gca gaa acg att cgt ctc atc tct gaa ggt ggc ccc 144
 Asn Pro Asp Tyr Ala Glu Thr Ile Arg Leu Ile Ser Glu Gly Gly Pro
 35 40 45

gat gcg ttc tac acg ggt gag att gca gca gac atc gtg gaa cgc gcc 192
 Asp Ala Phe Tyr Thr Gly Glu Ile Ala Ala Asp Ile Val Glu Arg Ala
 50 55 60

acc cgt gag gtt gac ggt ttc aca cca tca ctg atg agc acg gca gat 240
 Thr Arg Glu Val Asp Gly Phe Thr Pro Ser Leu Met Ser Thr Ala Asp
 65 70 75 80

ttg gct gcc tac act ccg gaa act cgt gaa gct ttg tgt gct ccc tac 288
 Leu Ala Ala Tyr Thr Pro Glu Thr Arg Glu Ala Leu Cys Ala Pro Tyr
 85 90 95

cgc gac aag att gtt tgt ggc atg cca ccg tca tca tcg ggt ggc gtc 336

Arg Asp Lys Ile Val Cys Gly Met Pro Pro Ser Ser Ser Gly Gly Val
 100 105 110

aca gtg atg gaa acc ctg ggt atc ttg aac aac ttt gat ctc gcc caa 384
 Thr Val Met Glu Thr Leu Gly Ile Leu Asn Asn Phe Asp Leu Ala Gln
 115 120 125

tac cca ccc act gag gtt ggt ttg gat ggc gga ttg cca aat gcg gaa 432
 Tyr Pro Pro Thr Glu Val Gly Leu Asp Gly Gly Leu Pro Asn Ala Glu
 130 135 140

gct gtt cac ctg att tca gag gct gag cgc ctg gct tat gct gat cgc 480
 Ala Val His Leu Ile Ser Glu Ala Glu Arg Leu Ala Tyr Ala Asp Arg
 145 150 155 160

gat gct tac atc ggt gat cct gct ttc 507
 Asp Ala Tyr Ile Gly Asp Pro Ala Phe
 165

<210> 86
 <211> 169
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 86
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Phe Leu Asp Glu Asn Gly Asp Ala Lys Ala Pro Gly Thr Leu Leu Gln
 20 25 30

Asn Pro Asp Tyr Ala Glu Thr Ile Arg Leu Ile Ser Glu Gly Gly Pro
 35 40 45

Asp Ala Phe Tyr Thr Gly Glu Ile Ala Ala Asp Ile Val Glu Arg Ala
 50 55 60

Thr Arg Glu Val Asp Gly Phe Thr Pro Ser Leu Met Ser Thr Ala Asp
 65 70 75 80

Leu Ala Ala Tyr Thr Pro Glu Thr Arg Glu Ala Leu Cys Ala Pro Tyr
 85 90 95

Arg Asp Lys Ile Val Cys Gly Met Pro Pro Ser Ser Ser Gly Gly Val
 100 105 110

Thr Val Met Glu Thr Leu Gly Ile Leu Asn Asn Phe Asp Leu Ala Gln
 115 120 125

Tyr Pro Pro Thr Glu Val Gly Leu Asp Gly Gly Leu Pro Asn Ala Glu
 130 135 140

Ala Val His Leu Ile Ser Glu Ala Glu Arg Leu Ala Tyr Ala Asp Arg
 145 150 155 160

Asp Ala Tyr Ile Gly Asp Pro Ala Phe
 165

<210> 87

<211> 604
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (101)..(604)
 <223> FRXA02000

<400> 87

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aatatctccc cacataaaag ttccttgata ggctcgagag atg aaa gtg acc caa 115
Met Lys Val Thr Gln
1 5

agc aca ttc ctt aaa tcg gta gct gcg ttc act gtc gca gcc tta acc 163
Ser Thr Phe Leu Lys Ser Val Ala Ala Phe Thr Val Ala Ala Leu Thr
10 15 20

ctg acc atc tct tcg tgt tcc agc ggt gaa gac acc tcc gca agc tcc 211
Leu Thr Ile Ser Ser Cys Ser Ser Gly Glu Asp Thr Ser Ala Ser Ser
25 30 35

acg gat act gaa aac tcc tca acc caa gca gca gcg tct ccc cca ctt 259
Thr Asp Thr Glu Asn Ser Ser Thr Thr Gln Ala Ala Ala Ser Pro Pro Leu
40 45 50

gcg cct tgt gaa ctt ccc gcc gac gct tct gct gaa gag gaa gta gaa 307
Ala Pro Cys Glu Leu Pro Ala Asp Ala Ser Ala Glu Glu Val Glu
55 60 65

ggc act cac aca ggt gaa gat att tct gtt gcc ccg gaa atc ggt acc 355
Gly Thr His Thr Gly Glu Asp Ile Ser Val Ala Pro Glu Ile Gly Thr
70 75 80 85

ggc tac cgc gag ggc atg acc cct gtt caa acc caa ggt tat gcg gtg 403
Gly Tyr Arg Glu Gly Met Thr Pro Val Gln Thr Gln Gly Tyr Ala Val
90 95 100

gca act gca aac ccc atc gct tct gaa gca gcc tgc gcg gtg tta aga 451
Ala Thr Ala Asn Pro Ile Ala Ser Glu Ala Ala Cys Ala Val Leu Arg
105 110 115

gaa ggc ggc act gca gct gat gct ctt gtc acc gcg cag ttt gtt ttg 499
Glu Gly Gly Thr Ala Ala Asp Ala Leu Val Thr Ala Gln Phe Val Leu
120 125 130

gga ctg acg gaa ccg cag tcg tct ggc ctt ggt ggt ggc gga tac att 547
Gly Leu Thr Glu Pro Gln Ser Ser Gly Leu Gly Gly Gly Tyr Ile
135 140 145

ctg tac tac gac gcc gaa gcc aat gcg gtg aca gcc att gat ggc cgt 595
Leu Tyr Tyr Asp Ala Glu Ala Asn Ala Val Thr Ala Ile Asp Gly Arg
150 155 160 165

gaa aca gcg
Glu Thr Ala 604

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<210> 88
 <211> 168
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 88
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 20 25 30
 Thr Ser Ala Ser Ser Thr Asp Thr Glu Asn Ser Ser Thr Gln Ala Ala
 35 40 45
 Ala Ser Pro Pro Leu Ala Pro Cys Glu Leu Pro Ala Asp Ala Ser Ala
 50 55 60
 Glu Glu Glu Val Glu Gly Thr His Thr Gly Glu Asp Ile Ser Val Ala
 65 70 75 80
 Pro Glu Ile Gly Thr Gly Tyr Arg Glu Gly Met Thr Pro Val Gln Thr
 85 90 95
 Gln Gly Tyr Ala Val Ala Thr Ala Asn Pro Ile Ala Ser Glu Ala Ala
 100 105 110
 Cys Ala Val Leu Arg Glu Gly Gly Thr Ala Ala Asp Ala Leu Val Thr
 115 120 125
 Ala Gln Phe Val Leu Gly Leu Thr Glu Pro Gln Ser Ser Gly Leu Gly
 130 135 140
 Gly Gly Gly Tyr Ile Leu Tyr Tyr Asp Ala Glu Ala Asn Ala Val Thr
 145 150 155 160
 Ala Ile Asp Gly Arg Glu Thr Ala
 165

<210> 89
 <211> 824
 <212> DNA
 <213> Corynebacterium glutamicum

<220>
 <221> CDS
 <222> (1)..(801)
 <223> RXN03178

<400> 89
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 Pro Thr Thr Val Val Thr Gly Thr Met Glu Ala Ala Asn Ile Glu Gly
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 Ser Arg Val Gly Val Gly Glu Ala Gly Gln Tyr Thr Val Asp Gln Leu
 20 25 30
 ctg cac ggt ctt ctt tta gcc agc ggt aac gat gcg gcg tat atg ttg 144
 Leu His Gly Leu Leu Leu Ala Ser Gly Asn Asp Ala Ala Tyr Met Leu

35	40	45	
gct cag gaa ctt ggt ggg gat caa gca acc ctg gag aaa gta aac gcg 192			
Ala Gln Glu Leu Gly Gly Asp Gln Ala Thr Leu Glu Lys Val Asn Ala			
50	55	60	
ctg gcc aag gag ttg ggc act caa gac acc ttc gtt gcc act tat tcc 240			
Leu Ala Lys Glu Leu Gly Thr Gln Asp Thr Phe Val Ala Thr Tyr Ser			
65	70	75	80
ggt ttg gat gcg ccg gga atg tcg acc tcc gca tac gac atg tca ttg 288			
Gly Leu Asp Ala Pro Gly Met Ser Thr Ser Ala Tyr Asp Met Ser Leu			
85	90	95	
att tat cag cat gcg tgg cag aac ccg gtt ttc gag tcg att atc tcc 336			
Ile Tyr Gln His Ala Trp Gln Asn Pro Val Phe Glu Ser Ile Ile Ser			
100	105	110	
acc gat cac att gat ttc cct ggt tgg ggc gac aat gag ggt ttc caa 384			
Thr Asp His Ile Asp Phe Pro Gly Trp Gly Asp Asn Glu Gly Phe Gln			
115	120	125	
gtc tgg aac gat aac gcc ttg ttc atg aac gat cct gat ggc atc ggc 432			
Val Trp Asn Asp Asn Ala Leu Phe Met Asn Asp Pro Asp Gly Ile Gly			
130	135	140	
ggc aag acc ggc tac acc gac gac gcg aac cac acc ttt gtc ggc ggt 480			
Gly Lys Thr Gly Tyr Thr Asp Asp Ala Asn His Thr Phe Val Gly Gly			
145	150	155	160
ctc gat cgg ggt ggt cgc cgc ctc gcc gcc gta ctc ttg gat tcc acc 528			
Leu Asp Arg Gly Gly Arg Arg Leu Ala Ala Val Leu Leu Asp Ser Thr			
165	170	175	
gtc agc gac att cgt ccg tgg gaa caa gca cga ttg ctt atc gac gcc 576			
Val Ser Asp Ile Arg Pro Trp Glu Gln Ala Arg Leu Leu Ile Asp Ala			
180	185	190	
tcc ctc ccc atc acg ccg ggg tcc ggc gtg ggc cag ctg ggc tcc ggc 624			
Ser Leu Pro Ile Thr Pro Gly Ser Gly Val Gly Gln Leu Gly Ser Gly			
195	200	205	
agc gcg aac gat gtg gca ccg gcg acc cca gaa tta cca gaa ccc acc 672			
Ser Ala Asn Asp Val Ala Pro Ala Thr Pro Glu Leu Pro Glu Pro Thr			
210	215	220	
gac aac ctg act tca ggt gag ggt ggg tcg cag aac acg ctt ctt aag 720			
Asp Asn Leu Thr Ser Gly Glu Gly Ser Gln Asn Thr Leu Leu Lys			
225	230	235	240
ctc gtg gtg ccc atc gga atc atc gtg ctg ttg cta atc gcc gca cta 768			
Leu Val Val Pro Ile Gly Ile Ile Val Leu Leu Leu Ile Ala Ala Leu			
245	250	255	
gcg tgg aca ttc aga tct ccc aag aaa aag aac taggtgttct tcttcacgac 821			
Ala Trp Thr Phe Arg Ser Pro Lys Lys Lys Asn			
260	265		
ctc			824

<210> 90
 <211> 267
 <212> PRT
 <213> Corynebacterium glutamicum

<400> 90

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Pro Thr Val Val Thr Gly Thr Met Glu Ala Ala Asn Ile Glu Gly
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Ser Arg Val Gly Val Gly Glu Ala Gly Gln Tyr Thr Val Asp Gln Leu
          20           25           30

Leu His Gly Leu Leu Leu Ala Ser Gly Asn Asp Ala Ala Tyr Met Leu
      35           40           45

Ala Gln Glu Leu Gly Gly Asp Gln Ala Thr Leu Glu Lys Val Asn Ala
      50           55           60

Leu Ala Lys Glu Leu Gly Thr Gln Asp Thr Phe Val Ala Thr Tyr Ser
 65           70           75           80

Gly Leu Asp Ala Pro Gly Met Ser Thr Ser Ala Tyr Asp Met Ser Leu
          85           90           95

Ile Tyr Gln His Ala Trp Gln Asn Pro Val Phe Glu Ser Ile Ile Ser
      100           105           110

Thr Asp His Ile Asp Phe Pro Gly Trp Gly Asp Asn Glu Gly Phe Gln
      115           120           125

Val Trp Asn Asp Asn Ala Leu Phe Met Asn Asp Pro Asp Gly Ile Gly
      130           135           140

Gly Lys Thr Gly Tyr Thr Asp Asp Ala Asn His Thr Phe Val Gly Gly
 145           150           155           160

Leu Asp Arg Gly Gly Arg Arg Leu Ala Ala Val Leu Leu Asp Ser Thr
      165           170           175

Val Ser Asp Ile Arg Pro Trp Glu Gln Ala Arg Leu Leu Ile Asp Ala
      180           185           190

Ser Leu Pro Ile Thr Pro Gly Ser Gly Val Gly Gln Leu Gly Ser Gly
      195           200           205

Ser Ala Asn Asp Val Ala Pro Ala Thr Pro Glu Leu Pro Glu Pro Thr
      210           215           220

Asp Asn Leu Thr Ser Gly Glu Gly Gly Ser Gln Asn Thr Leu Leu Lys
 225           230           235           240

Leu Val Val Pro Ile Gly Ile Ile Val Leu Leu Leu Ile Ala Ala Leu
      245           250           255

Ala Trp Thr Phe Arg Ser Pro Lys Lys Lys Asn
      260           265

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<210> 91
 <211> 749
 <212> DNA

<213> Corynebacterium glutamicum

<220>

<221> CDS

<222> (1)..(726)

<223> FRXA02859

<400> 91

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Gln Tyr Thr Val Asp Gln Leu Leu His Gly Leu Leu Leu Ala Ser Gly	
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aac gat gcg gcg tat ctg ttg gct cag gaa ctt ggt ggg gat caa gca	96
Asn Asp Ala Ala Tyr Leu Leu Ala Gln Glu Leu Gly Gly Asp Gln Ala	
20 25 30	
acc ctg gag aaa gta aac gcg ctg gcc aag gag ttg ggc act caa gac	144
Thr Leu Glu Lys Val Asn Ala Leu Ala Lys Glu Leu Gly Thr Gln Asp	
35 40 45	
acc ttc gtt gcc act tat tcc ggt ttg gat gcg ccg gga atg tcg acc	192
Thr Phe Val Ala Thr Tyr Ser Gly Leu Asp Ala Pro Gly Met Ser Thr	
50 55 60	
tcc gca tac gac atg tca ttg att tat cag cat gcg tgg cag aac ccg	240
Ser Ala Tyr Asp Met Ser Leu Ile Tyr Gln His Ala Trp Gln Asn Pro	
65 70 75 80	
gtt ttc gag tcg att atc tcc acc gat cac att gat ttc cct ggt tgg	288
Val Phe Glu Ser Ile Ile Ser Thr Asp His Ile Asp Phe Pro Gly Trp	
85 90 95	
ggc gac aat gag ggt ttc caa gtc tgg aac gat aac gcc ttg ttc atg	336
Gly Asp Asn Glu Gly Phe Gln Val Trp Asn Asp Asn Ala Leu Phe Met	
100 105 110	
aac gat cct gat ggc atc ggc ggc aag acc ggc tac acc gac gac gcg	384
Asn Asp Pro Asp Gly Ile Gly Gly Lys Thr Gly Tyr Thr Asp Asp Ala	
115 120 125	
aac cac acc ttt gtc ggc ggt ctc gat cgg ggt ggt cgc cgc ctc gcc	432
Asn His Thr Phe Val Gly Gly Leu Asp Arg Gly Gly Arg Arg Leu Ala	
130 135 140	
gcc gta ctc ttg gat tcc acc gtc agc gac att cgt ccg tgg gaa caa	480
Ala Val Leu Leu Asp Ser Thr Val Ser Asp Ile Arg Pro Trp Glu Gln	
145 150 155 160	
gca cga ttg ctt atc gac gcc tcc ctc ccc atc acg ccg ggg tcc ggc	528
Ala Arg Leu Leu Ile Asp Ala Ser Leu Pro Ile Thr Pro Gly Ser Gly	
165 170 175	
gtg ggc cag ctg ggc tcc ggc agc gcg aac gat gtg gca ccg gcg acc	576
Val Gly Gln Leu Gly Ser Gly Ser Ala Asn Asp Val Ala Pro Ala Thr	
180 185 190	
cca gaa tta cca gaa ccc acc gac aac ctg act tca ggt gag ggt ggg	624
Pro Glu Leu Pro Glu Pro Thr Asp Asn Leu Thr Ser Gly Glu Gly Gly	
195 200 205	
tcg cag aac acg ctg ctt aag ctc gtg gtg ccc atc gga atc atc gtg	672

Ser Gln Asn Thr Leu Leu Lys Leu Val Val Pro Ile Gly Ile Ile Val
 210 215 220

ctg ttg cta atc gcc gca cta gcg tgg aca ttc aga tct ccc aag aaa 720
 Leu Leu Leu Ile Ala Ala Leu Ala Trp Thr Phe Arg Ser Pro Lys Lys
 225 230 235 240

aag aac taggtgttct tcttcacgac etc 749
 Lys Asn

<210> 92

<211> 242

<212> PRT

<213> Corynebacterium glutamicum

<400> 92

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Thr Leu Glu Lys Val Asn Ala Leu Ala Lys Glu Leu Gly Thr Gln Asp
 35 40 45

Thr Phe Val Ala Thr Tyr Ser Gly Leu Asp Ala Pro Gly Met Ser Thr
 50 55 60

Ser Ala Tyr Asp Met Ser Leu Ile Tyr Gln His Ala Trp Gln Asn Pro
 65 70 75 80

Val Phe Glu Ser Ile Ile Ser Thr Asp His Ile Asp Phe Pro Gly Trp
 85 90 95

Gly Asp Asn Glu Gly Phe Gln Val Trp Asn Asp Asn Ala Leu Phe Met
 100 105 110

Asn Asp Pro Asp Gly Ile Gly Gly Lys Thr Gly Tyr Thr Asp Asp Ala
 115 120 125

Asn His Thr Phe Val Gly Gly Leu Asp Arg Gly Gly Arg Arg Leu Ala
 130 135 140

Ala Val Leu Leu Asp Ser Thr Val Ser Asp Ile Arg Pro Trp Glu Gln
 145 150 155 160

Ala Arg Leu Leu Ile Asp Ala Ser Leu Pro Ile Thr Pro Gly Ser Gly
 165 170 175

Val Gly Gln Leu Gly Ser Gly Ser Ala Asn Asp Val Ala Pro Ala Thr
 180 185 190

Pro Glu Leu Pro Glu Pro Thr Asp Asn Leu Thr Ser Gly Glu Gly Gly
 195 200 205

Ser Gln Asn Thr Leu Leu Lys Leu Val Val Pro Ile Gly Ile Ile Val
 210 215 220

Leu Leu Leu Ile Ala Ala Leu Ala Trp Thr Phe Arg Ser Pro Lys Lys

240

ttacagagt	gcttatgagg	caatcagcca	ctaagtgttg	agtaatctac	tagtttggac	60
tagaagttac	ccactttcag	tgaattttta	aggagagaac	atg Met	gct Ala	115
				1	5	
acc cga ttt gcc act cgt cgt cgc gca ctt gcc gca aaa ctg gca gct	163					
Thr Arg Phe Ala 10 Arg Arg Arg Ala 15 Ala Ala Lys Leu Ala 20						
caa cgg atc gac tca att ttg gtg aca agc ccg atc cat gtt cgc tat	211					
Gln Arg Ile Asp 25 Ser Ile Leu Val Thr 30 Ser Pro Ile His Val Arg Tyr						
ctc agc gga ttc acc ggc tcc aac ggc gca ctg atc gtg aac aaa gat	259					
Leu Ser 40 Gly Phe Thr Gly Ser Asn 45 Gly Ala Leu Ile 50 Val Asn Lys Asp						
ctc tcc cgc cag atc tgc acc gac ggt cgc tac acc acc cag atc gca	307					
Leu Ser 55 Ala Gln Ile Cys Thr 60 Asp Gly Arg Tyr Thr 65 Thr Thr Gln Ile Ala						
gaa gaa gtc ccg gac atc gag cgc ctg att gag cgt gcc tcg gca acg	355					
Glu Glu Val Pro Asp 75 Ile Glu Ala Leu Ile 80 Glu Arg Ala Ser Ala Thr 85						
acg ctg cta cgc cag gtc gaa ggg ccg cgt cgt ata gca atc gaa gcc	403					
Thr Leu Leu Ala Gln 90 Val Glu Gly Pro Arg 95 Arg Ala Ile Ile 100 Glu Ala						
gca caa acc acc ctg gac cag cta gac agc ctg cgt gaa gca acc cag	451					
Ala Gln Thr Thr 105 Leu Asp Gln Leu Asp 110 Ser Leu Arg Glu Ala Thr Gln 115						
gaa gac gtc gag ctg atc ccc gtc tca ggt gtt gtg gaa tcc att cgc	499					
Glu Asp Val 120 Glu Leu Ile Pro Val 125 Ser Gly Val Val 130 Glu Ser Ile Arg						
ctg acc aaa gac agc ttc gaa ctc gac cgc ctc cgc gat gtc gca gcg	547					
Leu Thr 135 Lys Asp Ser Phe Glu 140 Leu Asp Arg Leu Arg 145 Asp Val Ala Ala						
ctg gct tcc caa gca ttc gaa gat tta ctc gca gga gaa ctc gcc	595					
Leu Ala Ser Gln Ala Phe Glu Asp Leu Leu Ala Ala Gly Glu Leu Ala						

150	155	160	165	
gaa ggc cga tca gag cgc caa gtc gcc gcc gat ctg gaa tac cgc atg Glu Gly Arg Ser Glu Arg Gln Val Ala Ala Asp Leu Glu Tyr Arg Met	170	175	180	643
cgc ctg ttg gga gca gaa cgc ccc agc ttc gac acc atc gtg gcc tct Arg Leu Leu Gly Ala Glu Arg Pro Ser Phe Asp Thr Ile Val Ala Ser	185	190	195	691
gga cct aac tcc gcg aaa cca cac cac ggc gca ggc gac cgc atc ctc Gly Pro Asn Ser Ala Lys Pro His His Gly Ala Gly Asp Arg Ile Leu	200	205	210	739
cag cgc ggc gat cta gtc acc atc gat ttc ggc gca cac gca cgc gga Gln Arg Gly Asp Leu Val Thr Ile Asp Phe Gly Ala His Ala Arg Gly	215	220	225	787
ttc aac tcc gat atg acc cgc acc ctc gtt atg ggc gaa gca ggg gag Phe Asn Ser Asp Met Thr Arg Thr Leu Val Met Gly Glu Ala Gly Glu	230	235	240	835
ttc gaa gca gaa atc tac gac atc gtc ctg cgc tcc caa ctc gct ggt Phe Glu Ala Glu Tyr Asp Ile Val Leu Arg Ser Gln Leu Ala Gly	250	255	260	883
gtt gaa gca gcc tac tca ggc gcc aac ctc ttc gac atc gac gca gca Val Glu Ala Ala Tyr Ser Gly Ala Asn Leu Phe Asp Ile Asp Ala Ala	265	270	275	931
tgc cgc aaa atc atc gaa gac gca ggc tac ggc gaa tac ttc gtg cac Cys Arg Lys Ile Ile Glu Asp Ala Gly Tyr Gly Glu Tyr Phe Val His	280	285	290	979
tcc acc ggc cac ggc atc gga ctt gaa gtc cac gaa gcc cca agc gca Ser Thr Gly His Gly Ile Gly Leu Glu Val His Glu Ala Pro Ser Ala	295	300	305	1027
tcc aaa acc tca caa gga gtc cta gaa acc ggc tcc aca ctg acc atc Ser Lys Thr Ser Gln Gly Val Leu Glu Thr Gly Ser Thr Leu Thr Ile	310	315	320	1075
gaa ccc gga att tac gtc ccc gga aag ggc ggc gta cgc atc gaa gac Glu Pro Gly Ile Tyr Val Pro Gly Lys Gly Gly Val Arg Ile Glu Asp	330	335	340	1123
acc ctg att att acc tca gga gca ccg gaa atc atc acc aag gtg agt Thr Leu Ile Ile Thr Ser Gly Ala Pro Glu Ile Ile Thr Lys Val Ser	345	350	355	1171
aag gac ctc atc gtg gtg taatctaggt gagctaatacg gtc Lys Asp Leu Ile Val Val	360			1212

<210> 94

<211> 363

<212> PRT

<213> Corynebacterium glutamicum

<400> 94

Met Ala Leu Ala Asp Thr Arg Phe Ala Thr Arg Arg Arg Ala Leu Ala
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 Ala Lys Leu Ala Ala Gln Arg Ile Asp Ser Ile Leu Val Thr Ser Pro
 20 25 30
 Ile His Val Arg Tyr Leu Ser Gly Phe Thr Gly Ser Asn Gly Ala Leu
 35 40 45
 Ile Val Asn Lys Asp Leu Ser Ala Gln Ile Cys Thr Asp Gly Arg Tyr
 50 55 60
 Thr Thr Gln Ile Ala Glu Glu Val Pro Asp Ile Glu Ala Leu Ile Glu
 65 70 75 80
 Arg Ala Ser Ala Thr Thr Leu Leu Ala Gln Val Glu Gly Pro Arg Arg
 85 90 95
 Ile Ala Ile Glu Ala Ala Gln Thr Thr Leu Asp Gln Leu Asp Ser Leu
 100 105 110
 Arg Glu Ala Thr Gln Glu Asp Val Glu Leu Ile Pro Val Ser Gly Val
 115 120 125
 Val Glu Ser Ile Arg Leu Thr Lys Asp Ser Phe Glu Leu Asp Arg Leu
 130 135 140
 Arg Asp Val Ala Ala Leu Ala Ser Gln Ala Phe Glu Asp Leu Leu Ala
 145 150 155 160
 Ala Gly Glu Leu Ala Glu Gly Arg Ser Glu Arg Gln Val Ala Ala Asp
 165 170 175
 Leu Glu Tyr Arg Met Arg Leu Leu Gly Ala Glu Arg Pro Ser Phe Asp
 180 185 190
 Thr Ile Val Ala Ser Gly Pro Asn Ser Ala Lys Pro His His Gly Ala
 195 200 205
 Gly Asp Arg Ile Leu Gln Arg Gly Asp Leu Val Thr Ile Asp Phe Gly
 210 215 220
 Ala His Ala Arg Gly Phe Asn Ser Asp Met Thr Arg Thr Leu Val Met
 225 230 235 240
 Gly Glu Ala Gly Glu Phe Glu Ala Glu Ile Tyr Asp Ile Val Leu Arg
 245 250 255
 Ser Gln Leu Ala Gly Val Glu Ala Ala Tyr Ser Gly Ala Asn Leu Phe
 260 265 270
 Asp Ile Asp Ala Ala Cys Arg Lys Ile Ile Glu Asp Ala Gly Tyr Gly
 275 280 285
 Glu Tyr Phe Val His Ser Thr Gly His Gly Ile Gly Leu Glu Val His
 290 295 300
 Glu Ala Pro Ser Ala Ser Lys Thr Ser Gln Gly Val Leu Glu Thr Gly
 305 310 315 320
 Ser Thr Leu Thr Ile Glu Pro Gly Ile Tyr Val Pro Gly Lys Gly Gly

	325		330		335	
Val Arg Ile Glu Asp Thr Leu Ile Ile Thr Ser Gly Ala Pro Glu Ile						
	340		345		350	
Ile Thr Lys Val Ser Lys Asp Leu Ile Val Val						
	355		360			
 <210> 95						
<211> 1404						
<212> DNA						
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<221> CDS						
<222> (101)..(1381)						
<223> RXN00499						
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gcagatatgtg ttcttgctgc tctaaggatga tttttgggca				gtg gtg ggg gtg gtg		115
				Val Val Gly Val Val		
				1	5	
tcc acc cct gcg cgt aac ctg gga agc atg act aaa aca ctt ggt tcc						163
Ser Thr Pro Ala Arg Asn Leu Gly Ser Met Thr Lys Thr Leu Gly Ser						
	10		15		20	
ctt cag ctg gaa gaa atc acg ctg acc ctc cct ctg act gaa gat gtg						211
Leu Gln Leu Glu Glu Ile Thr Leu Thr Leu Pro Leu Thr Glu Asp Val						
	25		30		35	
gcc gat gaa cgc acc att gat gtg ttc gca cgc att gcc aca cgc gtc						259
Ala Asp Glu Arg Thr Ile Asp Val Phe Ala Arg Ile Ala Thr Arg Val						
	40		45		50	
ggg gaa gac ctt cca tat tta gta ttc ctg cag ggt ggg cct ggc						307
Gly Gly Glu Asp Leu Pro Tyr Leu Val Phe Leu Gln Gly Gly Pro Gly						
	55		60		65	
aat gaa gct cca cgt cca agc ctt aat ccc ctc aac ccc aat tgg ttg						355
Asn Glu Ala Pro Arg Pro Ser Leu Asn Pro Leu Asn Pro Asn Trp Leu						
	70		75		80	85
ggc gtg gcc ttg gag gaa tac cgc gtg gtc atg ttg gat caa cgt ggc						403
Gly Val Ala Leu Glu Glu Tyr Arg Val Val Met Leu Asp Gln Arg Gly						
	90		95		100	
acc ggc cgt tcc acc cca gtg ggt aat gat att ttg gaa aaa ccc aca						451
Thr Gly Arg Ser Thr Pro Val Gly Asn Asp Ile Leu Glu Lys Pro Thr						
	105		110		115	
gca gaa gta gtg gag tac tta tcc cac ctg cgc gca gat ggc att gtg						499
Ala Glu Val Val Glu Tyr Leu Ser His Leu Arg Ala Asp Gly Ile Val						
	120		125		130	
cga gat gct gaa gcc ctg cgt aag cat ttg ggt gtg aat cag tgg aac						547
Arg Asp Ala Glu Ala Leu Arg Lys His Leu Gly Val Asn Gln Trp Asn						
	135		140		145	

ctt tta ggc cag tcc ttc gga ggt ttc acc acc ctg cat tac ttg tcc	595
Leu Leu Gly Gln Ser Phe Gly Gly Phe Thr Thr Leu His Tyr Leu Ser	
150 155 160 165	
cgg cac gcc gat tcc ttg gac aac gtg ttt att acc ggc ggt ctc agc	643
Arg His Ala Asp Ser Leu Asp Asn Val Phe Ile Thr Gly Gly Leu Ser	
170 175 180	
gct att gat cgc cca gca gaa gac gtg tat gcc aac tgt tac aac cgc	691
Ala Ile Asp Arg Pro Ala Glu Asp Val Tyr Ala Asn Cys Tyr Asn Arg	
185 190 195	
atg cgc cga aac tct gag gaa ttc tac cgt cgc ttc ccg caa tta cgg	739
Met Arg Arg Asn Ser Glu Glu Phe Tyr Arg Arg Phe Pro Gln Leu Arg	
200 205 210	
gaa act ttc cga ggg ttg gtt aat cgt gct cgc gcc ggg gag att gtg	787
Glu Thr Phe Arg Gly Leu Val Asn Arg Ala Arg Ala Gly Glu Ile Val	
215 220 225	
ctt ccc acc ggc gaa gtt gtg tca gaa acc agg ctg cga tcc ctt ggt	835
Leu Pro Thr Gly Glu Val Ser Glu Thr Arg Leu Arg Ser Leu Gly	
230 235 240 245	
cac ttg ttg ggt agc aat gac ggc tgg ttt gat ctg tac aac ctg ctg	883
His Leu Leu Gly Ser Asn Asp Gly Trp Phe Asp Leu Tyr Asn Leu Leu	
250 255 260	
gaa tta gat ccc acc tcc aac gct ttt gtc cat gac ctg gca gga ctt	931
Glu Leu Asp Pro Thr Ser Asn Ala Phe Val His Asp Leu Ala Gly Leu	
265 270 275	
ttg cct ttc ggc aac cgc aac cca att tat tac gtg ctc cat gag tcc	979
Leu Pro Phe Gly Asn Arg Asn Pro Ile Tyr Tyr Val Leu His Glu Ser	
280 285 290	
tct tac gcc gac ggt gtg gtg aca aat tgg gca gca gag cgt gtg ctt	1027
Ser Tyr Ala Asp Gly Val Val Thr Asn Trp Ala Ala Glu Arg Val Leu	
295 300 305	
cca gag gat ttc cgc gag gat cca aca ctg ctc acc ggt gag cac gtg	1075
Pro Glu Asp Phe Arg Glu Asp Pro Thr Leu Leu Thr Gly Glu His Val	
310 315 320 325	
ttc cag gag tgg aca gac acc gtg ccg tcg ctc aag ccg tgg aag gac	1123
Phe Gln Glu Trp Thr Asp Thr Val Pro Ser Leu Lys Pro Trp Lys Asp	
330 335 340	
gtt gcc ctg gca ttg gct cag cag gaa tgg ccc aag ctt tat gat gcg	1171
Val Ala Leu Ala Leu Ala Gln Gln Glu Trp Pro Lys Leu Tyr Asp Ala	
345 350 355	
aag gca ttg gaa aac tca cag gcc aag ggc gct gca gca gtg tat gcc	1219
Lys Ala Leu Glu Asn Ser Gln Ala Lys Gly Ala Ala Val Tyr Ala	
360 365 370	
aat gac gtt ttc gtc cca gtg gat tac tct ctg gaa acc gca caa cac	1267
Asn Asp Val Phe Val Pro Val Asp Tyr Ser Leu Glu Thr Ala Gln His	
375 380 385	

ctg ccc ggt gtg cag ctg ttt atc acc agc cag cat gaa cac aat gga 1315
 Leu Pro Gly Val Gln Leu Phe Ile Thr Ser Gln His Glu His Asn Gly
 390 395 400 405

ctt cgt gcc agc tca ggc gca gta ctg aag cac ctt ttc gat ctg gcc 1363
 Leu Arg Ala Ser Ser Gly Ala Val Leu Lys His Leu Phe Asp Leu Ala
 410 415 420

cac ggc cga gag gta cgc tgattcctcg tgtagtact agc 1404
 His Gly Arg Glu Val Arg
 425

<210> 96

<211> 427

<212> PRT

<213> Corynebacterium glutamicum

<400> 96

Val Val Gly Val Val Ser Thr Pro Ala Arg Asn Leu Gly Ser Met Thr
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Lys Thr Leu Gly Ser Leu Gln Leu Glu Glu Ile Thr Leu Thr Leu Pro
 20 25 30

Leu Thr Glu Asp Val Ala Asp Glu Arg Thr Ile Asp Val Phe Ala Arg
 35 40 45

Ile Ala Thr Arg Val Gly Gly Glu Asp Leu Pro Tyr Leu Val Phe Leu
 50 55 60

Gln Gly Gly Pro Gly Asn Glu Ala Pro Arg Pro Ser Leu Asn Pro Leu
 65 70 75 80

Asn Pro Asn Trp Leu Gly Val Ala Leu Glu Glu Tyr Arg Val Val Met
 85 90 95

Leu Asp Gln Arg Gly Thr Gly Arg Ser Thr Pro Val Gly Asn Asp Ile
 100 105 110

Leu Glu Lys Pro Thr Ala Glu Val Val Glu Tyr Leu Ser His Leu Arg
 115 120 125

Ala Asp Gly Ile Val Arg Asp Ala Glu Ala Leu Arg Lys His Leu Gly
 130 135 140

Val Asn Gln Trp Asn Leu Leu Gly Gln Ser Phe Gly Gly Phe Thr Thr
 145 150 155 160

Leu His Tyr Leu Ser Arg His Ala Asp Ser Leu Asp Asn Val Phe Ile
 165 170 175

Thr Gly Gly Leu Ser Ala Ile Asp Arg Pro Ala Glu Asp Val Tyr Ala
 180 185 190

Asn Cys Tyr Asn Arg Met Arg Arg Asn Ser Glu Glu Phe Tyr Arg Arg
 195 200 205

Phe Pro Gln Leu Arg Glu Thr Phe Arg Gly Leu Val Asn Arg Ala Arg
 210 215 220